



SDHA Germline Variants in Adult Patients With SDHA-Mutant Gastrointestinal Stromal Tumor

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Background: SDH-*deficient* gastrointestinal stromal tumors (GIST) account for 20–40% of all KIT/PDGFRA-negative GIST and are due to mutations in one of the four *SDH*-complex subunits, with *SDHA* mutations as the most frequent. Here we sought to evaluate the presence and prevalence of *SDHA* variants in the germline lineage in a population of *SDHA-deficient* GIST.

Methods: Germline *SDHA* status was assessed by Sanger sequencing on a series of 14 patients with gastric *SDHA-deficient* GIST.

Results: All patients carried a germline *SDHA* pathogenic variant, ranging from truncating, missense, or splicing variants. The second hit was the loss of the wild-type allele or an additional somatic mutation. One-third of the patients were over 50 years old. GIST was the only disease presentation in all cases except one, with no personal or familial cancer history. Seven metastatic cases received a multimodal treatment integrating surgery, locoregional and medical therapy. The mean follow-up time was of 10 years, confirming the indolent clinical course of the disease.

Conclusion: *SDHA* germline variants are highly frequent in SDHA-*deficient* GIST, and the disease may occur also in older adulthood. Genetic testing and surveillance of *SDHA*-mutation carriers and relatives should be performed.

Keywords: SDH-*deficient* GIST, SDHA, CT: Carney Triad, CSS: Carney-Stratakis syndrome, SDHA germinal mutations, gastrointestinal stromal tumors

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INTRODUCTION

SDH-deficient gastrointestinal stromal tumors (GIST) account for 20-40% of all KIT/PDGFRA-negative GIST (1). Several evidences have suggested that SDH-deficient GIST exclusively arise from the stomach with mainly multifocal primary localization, frequently present lymph node involvement, generally affect younger population, and above all, have an indolent behavior even with metastatic disease (2). SDH deficiency in GIST is defined by the loss of expression of SDHB protein at immunohistochemistry and is mainly due to mutations in the four SDH mitochondrial complex subunits: SDHA, SDHB, SDHC, and SDHD (3). SDHA mutations are the most frequent among SDH alterations in GIST, accounting approximately for half of the cases. Other rare epigenetic events involve the recurrent aberrant DNA methylation of SDHC seen in GIST associated to the Carney triad (CT), which is a rare condition with synchronous or metachronous occurrence of GIST, paragangliomas, and pulmonary chondromas (4-6).

Most of SDH mutations in GIST are germline, in particular germline mutations in SDHB, SDHC, and SDHD occur in about 20-30% of SDH-deficient disease and may be referred to as a Carney-Stratakis syndrome (CSS) (4). This syndrome was firstly described in 2002 as a hereditary condition characterized by the occurrence of GIST and paraganglioma. Germline SDHA pathogenic variants have been rarely described in apparently sporadic cases (7, 8). Currently, germline testing is recommended for all SDH-deficient GIST, but no clear guidelines for genetic counseling and follow-up of SDH mutation carriers and relatives have vet been released (9). Moreover, regarding SDHA mutations, many issues are still unclear such as the germline status and the clinical implications that are not yet linked to well-defined hereditary syndrome. The aim of this work is to evaluate the presence and the prevalence of SDHA variants in the germline lineage in an SDH-deficient GIST population harboring SDHA somatic mutations.

MATERIALS AND METHODS

Patients

Sixteen patients with a gastric *SDHA*-mutant GIST were studied. All cases were negative for SDHB immunohistochemistry, and *SDHA* mutations were assessed by Sanger sequencing of coding exons and exon-flanking regions as previously reported (10).

The tumor and patients' characteristics are reported in **Table 1**. The study was performed in accordance with the Declaration of Helsinki protocols. The study was reviewed and approved by the local Institutional Ethical Committee of Azienda Ospedaliero-Universitaria Policlinico S. Orsola-Malpighi, Bologna, Italy (approval number 113/2008/U/Tess), and informed consent was provided by all living patients.

Sanger Sequencing

For germline analysis, DNA was extracted from peripheral blood or FFPE normal tissue with the QiaAmp mini or micro kit (Qiagen). For somatic analysis, manual macrodissection of the tumor area was performed using a scalpel on areas selected by an expert pathologist on FFPE slides. At least 70% tumor enrichment was required for sample inclusion, and DNA was extracted using QiaAmp micro kit. SDHA variants were identified by Sanger sequencing. Exonic and flanking intronic regions of SDHA were amplified with FastStart TAQ polymerase (Roche) and sequenced on both strands using the Big Dye Terminator v1.1 Cycle Sequencing kit (Applied Biosystems) on ABI 3730 Genetic Analyzer (Applied Biosystems). Primer pairs were designed with Primer Express 3.0 Software (Applied Biosystems) to specifically amplify SDHA exons and not the related pseudogenes (10). Germline mutational analysis was performed on peripheral blood in nine cases and on matched normal tissue extracted from FFPE in five cases. Unfortunately, in two cases the matched normal counterpart was not available. Allele frequency in the general population was reported from the Genome Aggregation Database (gnomAD) v2.1.1, reporting data from 141,456 individuals. Variant classification was performed following ACMG recommendations using the VarSome shared data resource (https://varsome.com/).

RESULTS

Germline Mutational Analysis

Germline variants were identified in all 14 patients for which the normal counterpart was available (Table 2). Five cases harbored truncating non-sense variants (Ser384*, Arg31*, Trp119*, Arg210*), seven other cases carried missense variants (Gly233Val, Arg171His, Arg589Gln, Gly257Ala, Arg600Gln, Arg585Gln), and two harbored exon-flanking intronic variants predicted to affect splicing. In particular, the prediction with the tool Alternative Splice Site Predictor (ASSP) revealed that the c.457-2_457del is likely to generate an alternative acceptor splice site of exon 5, while the c.1663+3 G>C probably leads to the loss of donor splice site at exon 12 with consequent intron retention. All the two splice site alterations, as predicted, lead to a frameshift with a stop codon in the corresponding protein sequence. All the truncating variants, one splice-site, and two missense variants were classified as "Pathogenic" or "Likely Pathogenic" by the ACMG recommendations (Table 2). Conversely, the other five variants were classified as "Uncertain Significance" but were all predicted as damaging by the computational prediction algorithms implemented in Varsome. In eight cases, tumor DNA showed the loss of the corresponding wild-type allele, thus displaying homozygosity for the germline variant, while in the other six cases compound heterozygosity for an additional somatic mutation was detected (Arg589Trp, Arg451Cys, Arg171Cys, Arg585Gln, Thr308Met, and Gln176*) (Figure 1A). In both patients for which normal DNA was not available, tumors carried two mutational hits on SDHA. Germline SDHA variants identified in this study, along with those identified in previous reports on GIST patients, are

Abbreviations: GIST, gastrointestinal stromal tumor; CT, Carney triad; CSS, Carney-Stratakis syndrome; gnomAD, Genome Aggregation Database; PPGL, pheochromocytoma and paraganglioma; ExAC, Exome Aggregation Consortium.

TABLE 1 | Patients and tumor characteristics.

Pts N°	Age	Gender	Primary Site	Multifocality	Disease Status at Diagnosis	Follow-up time	Patient Status	
#1	28	F	Stomach	Yes	Metastatic	15.8 yrs	AWD	
#2	30	Μ	Stomach	No	Metastatic	13.0 yrs	AWOD	
#3	31	F	Stomach	No	Localized	7.4 yrs	AWOD	
#4	61	Μ	Stomach	No	Localized	3.3 yrs	AWOD	
#5	21	F	Stomach	Yes	Localized	22.8 yrs	AWOD	
#6	39	F	Stomach	No	Metastatic	14.7 yrs	AWD	
#7	37	F	Stomach	No	Localized	9.6 yrs	AWOD	
#8	38	Μ	Stomach	No	Localized	5.0 yrs	AWOD	
#9	70	F	Stomach	No	Localized	10.0 yrs	AWOD	
#10	66	F	Stomach	No	Localized	2.2 yrs	AWD	
#11	17	Μ	Stomach	No	Localized	22.7 yrs	AWD	
#12	55	F	Stomach	No	Metastatic	12.0 yrs	DOD	
#13	50	Μ	Stomach	No	Localized	9.6 yrs	AWOD	
#14	17	Μ	Stomach	No	Localized	4.8 yrs	AWOD	
#15	54	F	Stomach	NA	NA	NĂ	NA	
#16	18	F	Stomach	Yes	Localized	NA	NA	

AWD, alive with disease; AWOD, alive without disease; DOD, died of disease; NA, not available.

summarized in **Figure 1B**. As expected, mutations are scattered along the whole coding sequence, with a peak frequency of the Arg31* variant, recurrently identified in previous studies (3, 7, 11–18) and present in one case in our series.

Clinical Correlation

Average age at diagnosis was 39.5 ± 4.5 years (range 17-70). Ten cases were young-adults (range: 17-39 years old) and 6 patients were older adults (>50 years old; range: 50-70). Combining our series with the previously published reports (3, 7, 8, 11-18), it emerges that GIST can arise at very different age ranges in SDHAvariant carriers, since mean age at diagnosis is 36.0 ± 2.3 years, but up to 22% of patients are being diagnosed at more than 50 years old (Figure 2). In all cases of adult patients, the GIST was unifocal. In the whole series, four patients displayed metastases already at the time of diagnosis, while all other harbored localized disease. In all cases except one, the GIST was the only disease presentation, and no personal or familial cancer history was revealed. Only one case of 61 years of age at diagnosis reported having been affected by paraganglioma in the past (more than 10 years before), but unfortunately the biological material was not available for a pathologic revision and genetic testing. Except for two cases lost at follow-up, the mean follow-up time was 10.9 years, ranging between 2.2 to 22.8 years. Among localized cases, three developed a recurrence with multiple metastases. Altogether, seven metastatic cases received a high-complexity treatment with standard medical therapy integrated with surgery and/or other loco-regional therapy (radiofrequency of liver metastasis and also chemoembolization in one case). Among these seven metastatic cases, one died of disease and one is currently free of disease through complete surgical removal of liver metastases.

DISCUSSION

In a population of SDH-*deficient* GIST harboring SDHA somatic mutations, we found germline SDHA variants in all cases for

which normal DNA was available, and these findings underline that germline mutations in SDHA are highly frequent in SDHAdeficient GIST. Besides the non-sense and missense mutations, we also found two cases harboring damaging splice-site mutations (c.457-2_457 and c.1663+3 G>C) leading to a predicted premature protein sequence truncation. Overall, in our series we found seven truncating germline variants (five nonsense and two splice-site) present in all but one case in young patients. Conversely, four out of seven germline missense variants, whose effect on protein function and stability is harder to be assessed, were identified in older patients (more than 50 years old at diagnosis). Moreover, we did not find a high frequency of the Arg31* variant, whose recurrent identification in previous studies even led to the assumption of a putative founder effect (3, 7, 14, 16–18). Apart from the recurrent Arg31*, the high diversity of the variant type and position seen in our and in previous series supports a view in which SDHA can be inactivated by mutations scattered throughout its whole genomic sequence, suggesting that SDHA genetic analysis must be performed entirely and by qualified high-volume molecular diagnostic centers, as already suggested in other settings (19).

Lastly, all the described patients carried two mutational events at the *SDHA* locus, either the loss of the wild-type allele or a second somatic event in compound heterozygosity, in full agreement with the two-hit hypothesis of tumor suppressor genes inactivation. Therefore, this suggests the possibility of a germline first mutation also in the two cases of our series for which the matched normal DNA counterpart was not available. Interestingly, an in-depth biochemical study of the functional effects of *SDHA* variants of unknown significance supported the pathogenicity of two somatic mutations (p.Arg451Cys e p.Gly419Arg) classified as uncertain by ACMG recommendations, while it did not support the loss of function of the Arg171His variant (20).

Furthermore, our study definitely confirms the highly relevant association between germline *SDHA* pathogenic variants and GIST onset, which is supported by many previous studies reporting the development of GIST as the only cancer

Pts N°		Normal	Counterpart Germ		Tumor Tissue Som	atic Mutation				
	Sample	Mutation	Variant Classification	Exon	Status	GnomAD Frequency	Sample	Mutation	Variant Classification	Exon
#1	PB	c.1151C>G; p.Ser384*	Р	9	Hetero	7.96x10 ⁻⁶	FF	LOH	Р	9
#2	PB	c.91C>T; p.Arg31*	Р	2	Hetero	2.09x10 ⁻⁴	FF	c.1765C>T; p.Arg589Trp	LP	13
#3	PB	c.1151C>G; p.Ser384*	Р	9	Hetero	7.96x10 ⁻⁶	FF/ FFPE	LOH	Р	9
#4	PB	c.698G>T; p.Gly233Val	UNC	6	Hetero	ND	FFPE	c.1351C>T; p.Arg451Cys	UNC	10
#5	PB	c.512G>A; p.Arg171His	UNC	5	Hetero	1.41x10 ⁻⁵	FFPE	LOH	UNC	5
#6	PB	c.1766G>A; p.ArgR589Gln	LP	13	Hetero	ND	FF	c.511C>T; p.Arg171Cys	UNC	5
#7	PB	c.457-2_457del	Р	5	Hetero	ND	FFPE	LOH	Р	5
#8	PB	c.770G>C; p.Gly257Ala	Р	6	Hetero	ND	FFPE	c.1754G>A; p.Arg585Gln	UNC	13
#9	FFPE	c.356G>A; p.Trp119*	Р	4	Hetero	ND	FF	LOH	Р	4
#10	FFPE	c.1799G>A; p.Arg600Gln	UNC	14	Hetero	2.09x10 ⁻⁵	FFPE	LOH	UNC	14
#11	PB	c.1663+3G>C	UNC	12	Hetero	1.59x10 ⁻⁵	FFPE	LOH	UNC	12
#12	FFPE	c.1799G>A; p.Arg600Gln	UNC	14	Hetero	2.09x10 ⁻⁵	FFPE	c.923C>T; p.Thr308Met	UNC	8
#13	FFPE	c.1754G>A; p.Arg585Gln	UNC	13	Hetero	7.97x10 ⁻⁶	FFPE	LOH	UNC	13
#14	FFPE	c.628C>T; p.Arg210*	Р	6	Hetero	3.98x10 ⁻⁶	FFPE	c.526C>T; p.Gln176*	Р	5
#15	NA	NA	NA	NA	NA	NA	FFPE	c.923C>T; p.Thr308Met	UNC +	8+13
								+ c.1741G>A; p.Gly581Arg	LP	
#16	NA	NA	NA	NA	NA	NA	FFPE	c.1255G>A; p.Gly419Arg + c.1690G>A; p.Glu564Lys	UNC + UNC	9+13

The cDNA and protein mutation in the normal germline and the second somatic mutation in GIST is reported, along with the variant classification following ACMG recommendations, the allelic status, and the allelic frequency in the general population (gnomAD database). PB, peripheral blood; FF, fresh-frozen; NA, not available; ND, not detected; P, pathogenic; LP, likely pathogenic; UNC, uncertain significance.

disease in SDHA germline-variant carrier population (3, 7, 13, 14, 18). In our series, only one case reported having been affected by paraganglioma more than 10 years before, but the diagnosis was not now revised. Recently, a large case-control study has estimated the penetrance of SDHx variants for PPGL, indicating a 1.7% lifetime disease penetrance for SDHA pathogenic variants (21). Up to now, the onset of other tumor types in the context of SDHA-mutant GIST appears to be very rare and limited to the concurrent description of paraganglioma in one case, of pulmonary chondroma in two cases, and of the full Carney triad in one patient (8, 14). Indeed, it was shown that mutations in SDHA account for around 30% of all KIT/PDGFRA-WT GIST (10), while being responsible for <1% of all pheochromocytoma and paraganglioma cases (PPGL), as opposed for example to SDHB that accounts for about 10% of all PPGL (22, 23). The only study that up to now suggesting the possible role of SDHA as a predisposing factor for other tumor types reported the development of neuroblastoma in one SDHA-mutation carrier showing the inactivation of the second allele (17). Therefore, our result, coupled with previous data and with the identification of a

higher-than-expected frequency of *SDHA* germline pathogenic variants in a cohort of cancer patients with respect to healthy individuals from the Exome Aggregation Consortium (ExAC) database (17), further endorses the view of *SDHA* as a cancer gene mainly predisposing to GIST development. This aggregate result underscores the different tumor spectrum of *SDHA*-germline variant carriers and Carney Stratakis Syndrome patients, which are known to harbor *SDHB*, *SDHC*, or *SDHD* mutations, and to develop invariably the association of GIST and paraganglioma during their life course (24).

Actually, in literature and in our experience, a clear syndrome has not been clearly defined since all these cases appear to be sporadic without other neoplasms except from GIST, and therefore it still remains unclear if *SDHA* mutations should be accounted in the CSS. CSS by definition is due to *SDHB*, *SDHC*, and *SDHD* mutations and is characterized by the combination of GIST and paraganglioma inherited in an apparently autosomal dominant manner and with incomplete penetrance. Paragangliomas were described to be multicentric and GIST multifocal. Both these clinical presentations support the

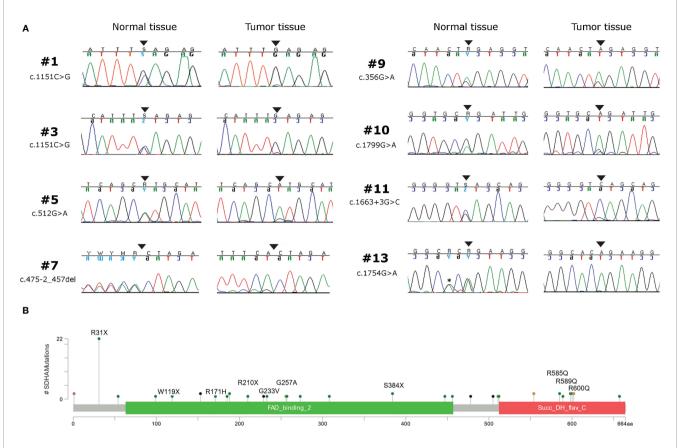


FIGURE 1 | (A) Chromatograms showing the loss of heterozygosity of *SDHA* germline variants in tumor tissue of eight GIST patients. Black arrows indicate the position of the mutation; the asterisk indicates a silent single nucleotide polymorphism. (B) *SDHA* germline variant frequency of GIST patients coming from this series (written protein mutation) and from previous reports (3, 7, 8, 11–17), shown as lollipop plot.

inherited nature of this tumor predisposition (4). Currently, germline testing is recommended for all SDH-*deficient* GIST including *SDHA* in some clinical practice guidelines (9, 25), but no clear protocol for genetic counseling and follow-up of *SDHx* variant carriers and relatives has been released, especially for those carrying a germline *SDHA* pathogenic variant not yet linked to well-defined hereditary syndrome.

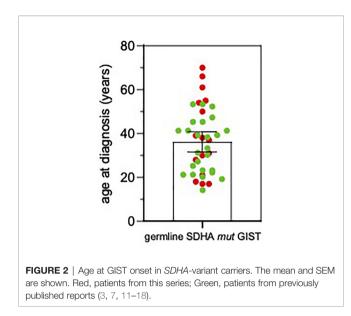
Regarding the clinical findings in GIST patients with SDHA germline mutation, our series confirms the stomach as unique site of GIST onset but underlines a higher heterogeneity for the multifocality development and also for the metastatic presentation at diagnosis. Larger series and international efforts should be required to better define a phenotype/genotype correlation in this subset of disease between GIST features, other tumor development, and SDH genotyping in tumor/ germline. No conclusive consideration can be made regarding the survival rates because of the heterogeneity of clinical presentations and the multiple treatments received in cases with metastatic disease. After all, our series confirms the indolent clinical course and long survival expectations since half of population presents a follow-up higher than 10 years, and four of these patients present a metastatic disease.

This study therefore shows that, differently from previously stated, the *SDHA*-mutant GIST patient is almost exclusively a

germline *SDHA*-variant carrier that is prone to develop the tumor throughout his entire life and not just in his early adulthood (7, 10– 12, 14). In fact, 38% of the patients from our series were older than 50 years, with peaks up to 70 years old. Actually, this result is supported by previous studies that occasionally reported age ranges up to 70 years old and subsets of *SDHA*-mutant GIST patients of more than 50 years old (3, 7, 13, 14, 16). This fact has very important implications supporting the need for genetic counseling, since even though complete pedigree analysis of *SDHA* mutation carrier families is still lacking and the clinical insights suggest a low penetrance of *SDHA* mutation in affected families (12, 26), the notion of a lifelong risk for GIST development urges the need for genetic testing and permanent clinical surveillance of *SDHA*-variant carriers.

As the process of referral to genetic counseling of the patients included in the study is still ongoing, conclusive information on documented history of cancer in relatives, segregation of the variant in the family, and clinical assessment of carriers, who will be included in management programs according to updated recommendations, are not yet available. Now, this may represent a formal limitation of the study, however not modifying its substantial conclusions.

In conclusion, germline pathogenic variants in SDHA are highly frequent in SDHA-deficient GIST in young and adult patients, and



the disease may occur also in older adulthood. Genetic counseling of *SDHA*-variant carriers and relatives should be planned, and their clinical follow-up should be accurately defined.

DATA AVAILABILITY STATEMENT

The datasets presented in this study was submitted to ClinVar (SUB10460221). Variants identified are available under accession numbers from SCV002026125 to SCV002026144.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Ethical Committe of Pol. Sant'Orsola Malpighi,

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AUTHOR CONTRIBUTIONS

Conceptualization: MP, MU, AAs, and VI. Data curation: VI, AAs, MN, and MSa. Formal Analysis; MU, VI, AS, AAs, and GR. Funding acquisition: MP and MN. Investigation: MU, VI, AS, AAs, BV, AB, GG, MC, EF, EC, MSa, AL, and GT. Methodology: MU, VI, AS, AAs, EG, AAl, and GR. Project administration: MP. Software: VI, GR, and MU. Supervision: MP, AAs, AAr, and PS. Validation: MU, VI, AS, AA, GR, AL, EG, AAl, and DB. Visualization: DT, MSe, and GT. Writing—original draft: MP, AAs, and VI. Writing—review and editing: MP, AAs, MU, and VI. All authors contributed to the article and approved the submitted version.

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