

Chemosphere

Elsevier Editorial System(tm) for

Manuscript Draft

Manuscript Number: CHEM53564

Title: Soil health, tomato yield and quality under organic vs. conventional management in a geogenic arsenic rich area

Article Type: Research paper

Section/Category: Environmental Chemistry (including Persistent Organic Pollutants and Dioxins)

Keywords: arsenic uptake; arsenic mobility; organic farming; microbial community; human risk.

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Abstract: The research studied the effects of organic vs. conventional management on soil health and tomato yield and quality cultivated in a geogenic arsenic contaminated soil. It was analyzed chemical and biochemical properties to evaluate soil health, arsenic mobility and its phyto-availability, as well arsenic accumulation in the tomato plant tissues and if tomato cultivated in arsenic rich soil represents a risk for human health. A general improvement of tomato growth and soil health was observed in the organic system, where soil organic carbon increased from 1.24 to 1.48% and total nitrogen content from 0.11 to 0.22%. Arsenic content in the organic system increased from 57.0 to 65.3 mg kg⁻¹, probably due to a greater content of organic matter that allow the soil to retain the arsenic naturally present in irrigation water. The increase of the total arsenic concentration in the organic system did not represent a stress factor for soil microbial biomass carbon (C_{mic}), which was higher in the organic system than in the conventional one (267 vs. 132 µg C_{mic} g⁻¹). A shift of microbial population from bacteria to fungi and protozoa was observed in the organic system; probably due to soil chemical properties changes such as the lower C/N ratio, the slight increase of pH (6.78 vs. 6.63) and the lower concentration of bioavailable arsenic. In the organic soil arsenic uptake resulted toned down compared to the conventional one. In both managements the concentration of arsenic in tomato fruits did not reach the threshold for human health risk.

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Prof. Roberto Mancinelli, PhD

Viterbo, April 11th, 2018

Prot. _____

To the Editors in Chief
GEODERMA Journal

Dear Editors,

Please, find here enclosed the Manuscript titled “Soil health, tomato yield and quality under organic vs. conventional management in a geogenic arsenic rich area” to be considered for publication in Geoderma journal.

This study was aimed to assess the impact of agricultural management, organic vs. conventional, on soil and tomato health in an As rich area irrigated with contaminated water. In particular, the attention has been posed on the tomato crops grown in a geogenic contaminated soil (As ranging from 56 to 75 mg kg⁻¹), irrigated with As rich water (35 µg L⁻¹ of As).

The very interesting aspect is that the concentration of As in the tomato fruits, which resulted not significantly different between the two systems, was not affected by the concentration of the As in soil. In addition, in both management systems, the tomato fruits analysed in this study have not exceeded the thresholds for As provided by the limits suggested by the European Food Safety Authority (EFSA).

To date there aren't scientific work aimed to compare these two managements for As behavior in soil-plant system, for this reason we think that this manuscript is of interest for Geoderma readers.

Best regards,

Roberto Mancinelli, on behalf of all the authors

1 **Soil health, tomato yield and quality under organic vs. conventional management in a**
2 **geogenic arsenic rich area**

3 Silvia Rita Stazi^{a*§}, Roberto Mancinelli^{b*§}, Rosita Marabottini^a, Enrica Allevato^a, Emanuele
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22 The authors of this article declared they have no conflicts of interest

23

Highlights

- An improvement of tomato and soil health was observed in the organic system.
- 56-75 mg As kg⁻¹ in soil didn't represent a stress for the microbial community
- The organic system mitigate As accumulation in plant tissues.
- Tomato fruits grown in As rich soil do not represent a risk for human health.

1 **Soil health, tomato yield and quality under organic vs. conventional management in a**
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24 **Abstract**

25 The research studied the effects of organic vs. conventional management on soil health and
26 tomato yield and quality cultivated in a geogenic arsenic contaminated soil. It was analyzed
27 chemical and biochemical properties to evaluate soil health, arsenic mobility and its phyto-
28 availability, as well arsenic accumulation in the tomato plant tissues and if tomato cultivated
29 in arsenic rich soil represents a risk for human health. A general improvement of tomato
30 growth and soil health was observed in the organic system, where soil organic carbon
31 increased from 1.24 to 1.48% and total nitrogen content from 0.11 to 0.22%. Arsenic content
32 in the organic system increased from 57.0 to 65.3 mg kg⁻¹, probably due to a greater content
33 of organic matter that allow the soil to retain the arsenic naturally present in irrigation water.
34 The increase of the total arsenic concentration in the organic system did not represent a stress
35 factor for soil microbial biomass carbon (C_{mic}), which was higher in the organic system than
36 in the conventional one (267 vs. 132 µg C_{mic} g⁻¹). A shift of microbial population from
37 bacteria to fungi and protozoa was observed in the organic system; probably due to soil
38 chemical properties changes such as the lower C/N ratio, the slight increase of pH (6.78 vs.
39 6.63) and the lower concentration of bioavailable arsenic. In the organic soil arsenic uptake
40 resulted toned down compared to the conventional one. In both managements the
41 concentration of arsenic in tomato fruits did not reach the threshold for human health risk.

42

43 **Keywords:** arsenic uptake; arsenic mobility; organic farming; microbial community; human
44 risk.

45

46

47 **1. Introduction**

48 Soil is a complex system with a high variability in terms of mineral and organic matter
49 composition. Some elements in agricultural soil, such as heavy metals, can be an undesirable
50 presence to ensure the quality of crop yield since those elements may represent risks for
51 human health. Arsenic (As) in its inorganic form can interact in various ways with the soil's
52 chemical and biological components that can affect its mobility, bioavailability and
53 translocation in plants tissues and finally to food chains. Arsenic concentration, in
54 uncontaminated soils, rarely exceeds 10 mg kg^{-1} ; conversely, depending on the nature of the
55 source (geogenic or anthropogenic), in polluted soils As concentration ranges from <1 to
56 $250,000 \text{ mg kg}^{-1}$.

57 A deeper knowledge of soil characteristics could allow to foresee As absorption and
58 desorption mechanisms, which would enable a proper use of soils in agricultural cropping
59 system to minimize the risks of the food chain contamination.

60 The As bioavailability does not correspond to the phyto-availability; the plants uptake of As
61 vary among species and it depends on As form and oxidation state. The management of
62 agroecosystems plays a key role on soil health, since the agricultural practices influence the
63 elements uptake by crops. Therefore their concentration in plant tissues, such as food or feed,
64 produces potential effects on human and animal health. Soil microorganisms and enzyme
65 activities are sensitive indicators of soil functionality when investigating the impacts of
66 metals/metalloids (e.g. As) (He et al., 2005) and/or agricultural management, such as organic
67 or conventional systems on environmental quality (Marinari et al., 2006). In particular, it is
68 known that the organic management improves soil nutritional and microbiological conditions;
69 with increased levels of available nutrients, microbial biomass content, and its activity
70 (Marinari et al., 2006; Reganold, 1988). Organic management is often considered as an
71 effective agricultural system producing healthy food (Reeve et al, 2016). However, very few

72 studies investigated the effect of agricultural management system, such as the organic
73 farming, on the behavior of heavy metals in naturally contaminated soils. Gousul Azam et al.
74 (2017) have studied the effect of organic vs. conventional farming on the As bioavailability
75 only for cereal crops, but no other study has been carried out on different crops. For this
76 reason, this study focused on the effect of agricultural management (organic vs. conventional)
77 on soil health and tomato yield quality in an As naturally contaminated area. In particular, the
78 attention has been posed on the tomato crops grown on geogenic contaminated soils (As
79 ranging from 56 to 75 mg kg⁻¹), irrigated with As rich water (35 µg L⁻¹ of As). The main
80 questions posed were: (i) Does agricultural management influence the As mobility and soil
81 health in a geogenic As rich area? (ii) Does the agricultural management system (ORG or
82 CONV) affect As accumulation in plant tissues? In particular, does the ORG system mitigate
83 the As plant uptake with respect to the CONV one? (iii) Does the tomato grown in As rich
84 soil represent a risk for human health?

85

86 **2. Materials and Methods**

87 2.1 The study area

88 The study area is located within the Cimino and Vico volcanic complexes (Figure 1), which
89 formed in the Pleistocene after post-orogenic extensional and local subsidence processes. The
90 region is characterized by substantial CO₂ emissions, which control the genesis of the
91 travertine deposits that outcrop throughout the Viterbo thermal area (Minissale and Duchi,
92 1988). The natural occurrence of arsenic in soil and water in the study area is related to the
93 presence of geothermal systems (Webster and Nordstrom, 2003; Ballantyne and Moore, 1988)
94 or to water–rock interactions that lead to arsenic mobilization from the aquifer (Charlet and
95 Polya, 2006; Smedley and Kinniburgh, 2002). In the Italian volcanic areas, high arsenic

96 concentrations have been related to the deep-rising fluids of the active geothermal systems
97 (Aiuppa et al., 2003).

98

99 **2.2 Experimental site**

100 The study was carried out in 2013 as part of a long-term field experiment established in
101 October 2001 at the Experimental Farm of the University of Tuscia (Viterbo, Italy) (Figure 1).

102 The soil was volcanic and classified in the textural class (0-20 cm depth) as clay loam with
103 45% sand, 38% clay and 17% silt, and a pH (H₂O) of 6.79.

104 The experimental field was arranged in a randomized block design with three replications,
105 where conventional (CONV) and organic (ORG) cropping systems were compared. In the
106 CONV system, the traditional agricultural practices (e.g. pesticides, chemical fertilizers, etc.)
107 were adopted, while in the ORG system the operations were carried out according to the
108 Council Regulation N. 834/2007 (EC, 2007) concerning organic production and the labeling
109 of organic products. A three-year crop rotation was carried out for both cropping systems
110 (CONV and ORG), including durum wheat (*Triticum durum* Desf.), processing tomato
111 (*Lycopersicon esculentum* Mill.), and chickpea (*Cicer arietinum* L.). In the organically
112 managed cropping system, the crop rotation was implemented with common vetch (*Vicia*
113 *sativa* L.) and oilseed (*Brassica napus* L.) cover crops, which were green manured about 10
114 days before tomato transplanting and pea planting, respectively. The three main crops were
115 grown each year in the experimental field consisting in 18 plots (2 cropping systems x 3
116 crops x 3 replicates). In both cropping systems (CONV and ORG) the maximum tillage depth
117 was 20 cm. Soil tillage was carried out before tomato transplanting in May and at the same
118 time the soil was fertilized with 80 kg ha⁻¹ P₂O₅ (using perphosphate in CONV and rock
119 phosphate in ORG system). The tomato crops were drip irrigated in order to reintegrate the
120 90% of water lost through evapotranspiration estimated by a class A pan and adjusted by crop

121 coefficients during the growth cycle. This study focused on the tomato crop inserted in the
122 organically and conventionally managed cropping systems in the long-term field experiment,
123 described above.

124

125 **2.3 Soil and plant sampling**

126 Soil sampling was performed in summertime (August 2013) at the end of the vegetative cycle
127 of the tomato crop. After removing the litter layer two soil cores (0-20 cm depth) were taken
128 from each plot and then pooled together, for a total of 12 soil samples. The soil samples were
129 air dried, sieved (<2 mm) prior to analyses. Two plant samples at their physiological maturity
130 were taken from the central area of each plot. The various parts of each plant (fruits, leaves,
131 stems and roots) were separated and transferred in laboratory for further analyses.

132

133 **2.4 Soil health**

134 Soil health in the ORG and CONV systems was assessed according to chemical and
135 biological properties, such as nutrients content, microbial community structure, its activity
136 and diversity.

137

138 *2.4.1 Soil nutrient content*

139 Soil total organic carbon (TOC) and total nitrogen (TN) were determined by dry combustion
140 using an elemental analyzer (Thermo Soil NC – Flash EA1112, USA), while Carbon and
141 Nitrogen labile pools were extracted using K₂SO₄ 0.5 M (1:4 w:v) and determined with the
142 TOC-V CSN and TNM-1 analyzer (Shimadzu, Japan). Moreover, soil available P was
143 determined following the colorimetric method (Bray and Kurtz, 1945).

144

145 *2.4.2 Microbial biomass, community profiles and its activity*

146 The total amount of microbial biomass C (MBC) and N (MBN) were analyzed using the
147 fumigation extraction (FE) method (Vance et al., 1987). Microbial biomass C was calculated
148 as follows: $C_{mic} = EC:k_{EC}$, where EC is the difference between organic C extracted from
149 fumigated soils and organic C extracted from non-fumigated soils and $k_{EC} = 2.64$. Microbial
150 biomass in terms of N was calculated as: $N_{mic} = EN:k_{EN}$, where EN is the difference between
151 organic N extracted from fumigated soils and organic N extracted from non-fumigated soils
152 and $k_{EN} = 2.22$.

153 The Ester linked-Fatty Acid Methyl Ester (EI-FAME) microbial community profiles were
154 determined (Stazi et al., 2017), quantified and converted to $\mu\text{mol}\cdot\text{g}^{-1}$ using peak areas from
155 internal standard (methylnonadecanoate, C19:0), used at known concentrations. Three
156 analytical replicates of each sample were extracted.

157 The sum of the EI-FAMEs considered to be predominantly of bacterial origin (i14:0, 14:0,
158 i15:0, a15:0, 15:0, i16:0, 16:0 16:1 ω 9; 16:1 ω 7t, i17:0, a17:0, 17:0, cy17:0, 18:0 and cy19:0)
159 was used as an index of the bacterial biomass (B), and the quantity of the FAs 18:2 ω 6,9 and
160 18:1 ω 9c were used as an index of the fungal biomass (F) (de Dato et al., 2017). The fungal
161 bacterial ratio (F/B ratio) was calculated as an index of soil microbial community change.

162 The sum of EI-FAMEs characteristic of general bacteria, G+ bacteria, G- bacteria,
163 actinomycetes, fungi, were used as broad taxonomic microbial groupings (Table 1). Lastly,
164 the ratio of monounsaturated (MONO) and saturated (SAT) EI-FAMEs were used as
165 indicators of physiological or nutritional stress in bacterial communities (Bossio and Scow
166 1998). Finally, the EI-FAME obtained were identified and organized in biomarker groups (de
167 Dato et al., 2017).

168 Soil microbial activity was measured in terms of both respiration and enzyme activities. For
169 measuring microbial respiration 20 g of soil (oven-dry basis) at 60 % of water holding

170 capacity (WHC) were placed in 1L hermetically sealed glass jars. The CO₂ released was
171 trapped, after 1, 3, 7, 10, 14, 21, 28 35, 42 days of incubation, in 2 ml 1N NaOH and
172 determined by titrating the excess NaOH with 0.1N HCl as reported in **Marabottini** et al.
173 (2013). Ecophysiological indices such as microbial quotient (q_{mic}, the percentage of
174 microbial carbon to total organic carbon) and metabolic quotient (qCO₂, the basal respiration
175 of microbial biomass unit) were both calculated according to Moscatelli et al. (2005). The
176 soils were analyzed for eight enzyme activities involved in the major nutrient cycles (C, N, P,
177 S-(**Marinari** et al., 2013).

178 The enzyme activities were determined with the microplate assay using fluorogenic
179 methylumbelliferyl (MUF)-substrates (Marx et al., 2001). The Synthetic Enzymatic Index
180 (SEI) was calculated as the sum of all enzyme activities which release the same reaction
181 product (MUF) (Moscatelli et al. 2018). Substrate induced respiration (SIR) was determined
182 using the Microresp© soil respiration system with 15 carbon substrates (Lagomarsino et al.,
183 2007). Finally, the microbial structural and functional diversity were assessed by the
184 Shannon's Weiner Index calculated using data of PLFA and microbial activity [enzyme
185 activities (H'Enz) and substrate induced respiration (H'SIR)], respectively (Zak et al., 1994).

186

187 **2.5 Arsenic in soil, plant tissues and irrigation water**

188 The total amount of As in soil (As_{tot}) and in the tomato plant components (roots, stem, leaves,
189 berries) was determined after mineralization of the samples with concentrated hydrochloric
190 acid (36% HCl), nitric acid (69% HNO₃) and hydrogen peroxide (30% H₂O₂) (Merck,
191 Darmstadt, Germany) with a microwave assisted digestion, using a commercial laboratory
192 microwave oven (Mars plus CEM, Italy). The procedure was reported in **Stazi** et al 2016 (a).
193 For the As quantification in soil and plant, was used an ICP-OES (8000 DV, PerkinElmer,
194 Shelton, CT, USA) equipped with: an ultrasonic nebulizer for plant samples, and a Scott

195 nebulizer for soil samples. The accuracy of the measurements was assessed using standard
196 reference materials trace metals: Loamy Sand 3 (CRM034-Fluka) and tomato leaves (SRM
197 1573a). The quantity of inorganic As in irrigation water was determined with hydride
198 generation method (HG-ICP-OES).

199 The mobility of As in soil was determined by sequential extraction procedure according to
200 **Marabottini** et al. (2013) with minor modifications. Briefly, three soil fractions were studied:
201 Water Soluble (WS), not Specifically Sorbed (NSS) and Specifically Sorbed (SS). The As of
202 the three fractions is considered chemically labile and available for plants. The sum of As
203 measured in the three fractions is named as $A_{S_{bio}}$. $A_{S_{tot}}$ and $A_{S_{bio}}$ together with soil available
204 phosphorous were used to assess the influence of the agricultural management systems on the
205 mobility of the toxic element in soil.

206 The plant/soil bioaccumulation factor of As (BAF) was calculated as the ratio between the
207 concentration of As in root and the $A_{S_{bio}}$ in soil. Arsenic translocation, from roots to the
208 aboveground part of the plant, was calculated as the ratio of the amount of As in shoots or
209 berries and the that in roots (Translocation Factor-TF) (Stazi et al., 2016b).

210

211 **2.6 Statistical analysis**

212 All data were subjected to analysis of variance (ANOVA) using JMP statistical software
213 package 4.0 (JMP, 2000). To homogenize the variance, after the Bartlett test, if necessary, the
214 data were subjected to angular transformation before the variance analysis. A randomized
215 block experimental design was adopted in the field located in the geogenic As rich area.
216 Treatment mean values were compared with Fisher's protected least significant difference
217 (LSD) test at the 0.05 probability level.

218

219 **3. Results**

220 **3.1 Soil health and arsenic availability**

221 In this work, soil health in ORG and CONV agricultural systems was assessed in terms of soil
222 chemical and biochemical properties. The organically managed soil showed a slight but
223 significant increase of pH (6.78 vs. 6.63), of total organic carbon (+19%) and a major increase
224 of total nitrogen (+100%). Similar trends were observed for the extractable fractions of C and
225 N. Conversely, even if the difference was not statistically significant, available P tended to be
226 higher in CONV with respect to ORG soil (+5%) (Table 1). The ORG system showed a
227 general soil health improvement, also by the increase of microbial activity (SEI and SIR)
228 (Table 1). Moreover, the Shannon-Weiner indices showed different patterns of microbial
229 functional diversity between the two systems, in ORG the index was lower in terms of
230 enzymatic hydrolytic activities (H'Enz) but higher in terms of catabolic functions (H'SIR)
231 than in CONV system (Table 1). Similarly, the soil microbial community composition
232 measured by EL-FAME analysis differed between the two systems. In particular, the ORG
233 system, microbial biomass and its diversity showed a shift of population from bacteria to
234 fungi and protozoa (Figure 2).

235 As for the concentration of arsenic in the ORG soil, the total As resulted significantly higher
236 than in the CONV one (65.3 vs. 57.5 mg kg⁻¹) (P= 0.002). Conversely, according to the
237 chemical fractionation, the bioavailable As_{bio} was significantly higher in CONV than in ORG
238 soils (7.05 vs 6.18 mg kg⁻¹) (P = 0.009). Moreover, negative correlations between As_{bio} and
239 TOC, As_{SS} and TOC (P<0.0001) were found in CONV soil, while between the same soil
240 properties the correlation coefficients were positive in the ORG soils (Table 2).

241 The concentration of 18:1 ω 9c was well correlated to 18:2 ω 6,9 and both FAs increased in
242 ORG soils, attesting an proliferation of fungal biomass. EL-FAMES markers of Gram (-)
243 bacteria were lower in ORG soil than in CONV ones. The ORG system showed the lowest

244 amount of saturated EL-FAMES typical markers of Gram (+) bacteria, while specific protozoa
245 and actinomycetes EI-FAMES, increased in the ORG system. Moreover, the sum of methyl-
246 branched FAs representing actinomycetes was slightly lower in CONV than in ORC system.
247 In addition, the G+/G- ratio was lower in ORG than in CONV, suggesting a relative
248 dominance of G+, compared with G-. An opposite response was obtained by the ratio of
249 monounsaturated and saturated EI-FAMES: the value for the ratio of Mono/Sat fatty acids was
250 0.8 in ORG and 0.7 in CONV which is consistent with the lower C and higher As_{bio} contents
251 in CONV. In Table 1 is also reported the F/B ratio which was significantly higher in ORG
252 than in CONV.

253

254 **3.2 Arsenic uptake and partitioning in plant organs**

255 In Table 3 is reported the partitioning of As in plant organs (concentration of As is referred to
256 the dry matter). The results show a higher quantity of phytoavailable As in roots grown in the
257 CONV than those grown in ORG system displayed by the significantly difference of BAF
258 (0.66 vs. 0.46). The bioavailable toxic element is easily accessible for the tomato plant uptake
259 by the root system. Tomato plants grown in As rich soil are able to take up As and accumulate
260 it in all the plant organs in both management systems. The As partitioning followed the trend
261 $As_{root} > As_{leaf} > As_{stem} > As_{fruit}$. The concentration of As accumulated in roots grown in CONV
262 is about two fold higher than As measured in roots from ORG. Even in the aerial parts, the
263 concentration of As in CONV was significantly higher than those in ORG, except the As in
264 berries where difference between the plants from the two managements was not statistically
265 significant. In both management systems, the plants were able to accumulate the toxic
266 element in the roots and only a little As was translocated in apogee. The ability of a plant to
267 translocate an element was measured by TF, this was higher in plants grown in ORG system
268 (0.035) with respect to those grown in the CONV system (0.019). This was an expected

269 result: the plant moves the toxic element through non-specific transporters (thanks to
270 structural affinity). The plant response to an increase in As concentration was in limiting the
271 toxic element's movement by rising transporters specificity and reducing its translocation
272 from hypogeous to the epigeous plant components.

273

274 **3.3 Tomato yield**

275 Thirteen years after the experimental field establishment, the organic cropping system
276 management significantly affected the tomato productions in a positive way (Table 3). Fruits,
277 leaves, stems and roots of tomato crop showed a significant increase of both fresh and dry
278 weight of biomass in the ORG management compared with the CONV one. In particular, the
279 aboveground biomass (fruits, leaves and stems) resulted higher in the organic management
280 than in the conventional one (+58%, +56% and +40% of dry weight fruits, leaves and stems,
281 respectively), while the roots biomass increased only by 27% of dry weight in the organic
282 management. Considering the whole biomass production of the tomato crop, the organic
283 management determined an increase of dry weight (+53%).

284

285 **4. Discussion**

286 Previous study conducted in a Mediterranean environment (Marinari et al., 2006; Mancinelli
287 et al., 2010) attested a general improvement of soil quality under organic management with
288 respect to the conventional one. In this study, conducted in a geogenic As rich area, although
289 an active metabolic response of microorganisms was observed under both managements, the
290 ORG system showed a general improvement of soil health, in terms of microbial activity (SEI
291 and SIR) and microbial diversity, with respect to CONV system. In fact, in the ORG system,
292 the increase of soil organic matter with lower C/N ratio, may enhance microbial biomass and
293 its diversity showing a shift of population from bacteria to fungi and protozoa, as resulted by

294 El-FAME analysis. Moreover, the increase of readily available nutrients, such as extractable
295 C and N, may have originated from a more intense rhizodeposition or turnover of roots in the
296 agricultural system, where cover crops are included in the crop rotation (Kong & Six, 2012).
297 By means of an ecological point of view, As labile form represents the bioavailable and
298 phytoavailable forms of the toxic element (Marabottini et al., 2013). Therefore, the CONV
299 managed soils could display an increased risk for soil health and tomato quality, because the
300 percentage of bioavailable As was 14% higher than in the ORG system. The management
301 seems to induce change of As and P concentration in soil; in fact the available P in CONV
302 system slightly increased (+5%). Therefore, as documented in previous works, the increased
303 available P may promote As mobility in soil through competition for the anionic sorption
304 sites, very affine to available P (Pigna et al., 2015; Caporale et al., 2018).

305 The highest concentration of As_{bio} in CONV could be one of the soil environmental factor
306 affecting microbial groups relative abundance. Therefore, the higher content of organic matter
307 and a lower concentration of As_{bio} in the ORG system, may affect soil microbial community
308 composition showing a more abundant fungal biomass with a reduction of bacteria (G+ and
309 G-). In this study, Protozoa represents the most sensible microbial group to the As_{bio}
310 concentrations in soil; this negative effect of heavy metals on Protozoa was previously
311 reported by Ekelund et al. (2003). Moreover, the sum of methyl-branched FAs representing
312 actinomycetes was slightly lower in the CONV. This result turns out to be in agreement with
313 those reported by other authors, as actinomycetes FAs are usually found in the soil where low
314 quantities of metal are detected (Kelly et al., 2003; He et al., 2005). The elevated
315 concentrations of the metalloid in organic matter enriched soil may contribute to a selection or
316 a shift of the microbial communities towards more metal-tolerant or metal-resistant species
317 (Moreno et al., 2012).

318 The ratio Mono/Sat used to evaluate the soil microbial communities status under various
319 environmental conditions, should approximately be less than 1 for soils receiving low organic
320 inputs (Zelles et al., 1995). In this study, the ORG management promoted an increase of the
321 Mono/Sat ratio, even if it was still lower than 1. This ratio, also used as an indicator of
322 microbial nutritional status, suggested the maintenance of nutrient bioavailability in the
323 contaminated soils, especially in the ORG system (Ellis et al., 2001). Moreover, since the
324 relative proportion of F/B biomass provides an indication of soil microbial community
325 stability, ecosystem self-regulation (Zeller et al., 2001) and soil quality change in
326 contaminated area (Mummey et al., 2002), the results obtained in this study proved that the
327 ORG management enhanced soil health. The ORG management, compared to the CONV one,
328 revealed a higher soil organic carbon content due to the higher plant residues soil input
329 (Mancinelli et al., 2013). At the same time, the soil As_{tot} resulted higher in the ORG than in
330 the CONV system probably because the increase of soil organic matter promoted the
331 adsorption of As added by irrigation water as a common scenario occurring in As-affected
332 Italian areas (Baiocchi et al. 2013). Conversely the bioavailable As form resulted higher in
333 the CONV system probably because the mineral fertilization, adopted in this system,
334 enhanced the mobility of As in soil (Caporale et al., 2018). Therefore, the As form in soil may
335 depend on applied agronomic practices since the crop residues management, addressed to
336 increase the soil organic matter in the ORG system, could contribute to link As, justifying the
337 highest total concentration and the lower mobility of this element in soil.

338

339 **5. Conclusions**

340 A general improvement of tomato growth and soil health was obtained by the organic
341 management system in a geogenic As rich area in Mediterranean environment. The
342 concentration of As in the tomato fruits was not significantly affected by the management

343 systems. In addition, in both CONV and ORG management, the tomato fruits have not
344 exceeded the thresholds for As provided by the limits suggested by the European Food Safety
345 Authority (EFSA, 2009).

346 According to the main questions posed, it can be concluded that: (1) the soil As concentration
347 ranging from 56-75 mg kg⁻¹ does not represent a stress factor for soil microbial community;
348 (2) the ORG management caused an increase of total As concentration in soil through the
349 enhanced organic matter content, retaining As from irrigation water; (3) the mobility of As
350 increases under CONV system, where more phosphorus is available. Moreover, the ORG
351 management mitigate the As uptake by tomato plant. Finally, even if the agricultural
352 management affects As translocation in plant tissues, (4) the tomato produced under both
353 ORG and CONV management in As rich soil does not represent a risk for human health.

354

355 **Acknowledgements**

356 The authors are grateful for the technical support in the experimental fields of Claudio
357 Stefanoni and Fulvia Gatti. The research activity was co-funded by the Italian Ministry of
358 University and Research (PRIN project "Health of agroecosystems: chemical, biochemical
359 and biological processes that regulate the mobility of As in the soil-water-plant
360 compartments" code 2010JBNLJ7_006).

361

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471 biomass and community structures in soils. *Biol. Fertil. Soils* 19, 115–123.

472

473 **Table 1** Soil quality in organic (ORG) and conventional (CONV) agricultural systems:
 474 chemical - biochemical properties and microbial diversity. Values belonging to the same
 475 parameter with different letters are statistically different according to LSD (0.05). In brackets
 476 the standard errors are reported (n=6).

	ORG	CONV
<i>Soil chemical properties</i>		
pH (H ₂ O)	6.78 a	6.63 b
TOC (%)	1.48 a	1.24 b
TN (%)	0.22 a	0.11 b
C/N	6.58 b	10.98 a
Available P (µg P g ⁻¹)	13.75 a	14.44 a
Extractable C (µg C g ⁻¹)	47.29 a	42.25 b
Extractable N (µg N g ⁻¹)	9.99 a	8.81 b
<i>Microbial biomass and diversity</i>		
Microbial Biomass (µg C-mic g ⁻¹)	267 a	132 b
H' Enz	1.21 b	1.38 a
H' SIR	2.62 a	2.54 b
H' EI-FAME	2.71 a	2.56 b
<i>Microbial activity</i>		
SEI (nmol MUF g ⁻¹ h ⁻¹)	1293 a	945 b
SIR (mg CO ₂ g ⁻¹)	26.64 a	25.01 b
<i>EI-FAME</i>		
Total Bacteria (µmol g ⁻¹)	6.56 b	7.76 a
G(+) (µmol g ⁻¹)	2.31 a	2.85 a
G(-) (µmol g ⁻¹)	1.27 a	1.36 a
Actynomicetes (µmol g ⁻¹)	0.70 a	0.66 a
Fungi (µmol g ⁻¹)	1.75 a	1.48 b
Protozoa (µmol g ⁻¹)	0.08 a	0.04 b
<i>Microbial indicators of stress</i>		
G(+)/G(-)	1.82 a	2.10 a
Mono/Sat	0.80 a	0.72 b
F/B	0.29 a	0.18 b

477 TOC: total organic carbon, TN: total nitrogen, H'Enz: Shannon-Weaver Index on enzyme
 478 activities base, H'SIR: Shannon's Index on substrate induced respiration base, SEI: Synthetic
 479 Enzyme Activity. H' EI-FAME : Shannon-Weaver Index from EI-FAMEs data.
 480

481

482 **Table 2** Pearson-correlation analysis results between studied parameter TOC-organic carbon
 483 content, TN-total nitrogen content; C/N ratio; As_{total} total As; WS-water soluble As; SS-No
 484 Specifically Sorbed; As; SS-Specifically Sorbed As; As_{bioav} -bioavailable As. n.s. = not
 485 significant; *, **, *** indicate the significance level at P≤0.05, P≤0.01, P≤0.001, respectively.

BIO	TOC	TN	C/N	As _{total}	WS	NSS	SS	As _{bioav}
CONV								
TOC		0.82 *		0.79 *			0.90**	0.74 *
TN	1.00 ***			0.99 ***			0.98 ***	0.99 ***
C/N	1.00 ***	1.00***			-0.96 ***	-0.82 *		
As _{total}	-0.75 *	-0.75 *	-0.76 *				0.97 ***	0.99 ***
WS	0.71 *	0.71 *				0.94 ***		
NSS				0.85 **				
SS	-1.00 ***	-1.00 ***	-1.00 ***	0.75 *	-0.71 *			0.95 ***
As _{bioav}	-0.99 ***	-0.99 ***	-0.99***	0.79 *			0.99 ***	

486

487

488 **Table 3** Arsenic partitioning , fresh and dry of biomass fractions and yield of the tomato
 489 production in organic (ORG) and conventional (CONV) management system. Values between
 490 systems for plant components with different letters are statistically different according to
 491 LSD (0.05).

492

Plant components	System	As mg kg⁻¹ D.W.	Fresh weight (t ha⁻¹)	Dry weight (t ha⁻¹)
fruit	ORG	0.09 a	50.60 a	4.05 a
	CONV	0.08 a	32.10 b	2.57 b
leaf	ORG	1.24 b	4.73 a	0.89 a
	CONV	1.30 a	2.98 b	0.57 b
steam	ORG	0.60 b	7.37 a	1.12 a
	CONV	0.69 a	5.07 b	0.80 b
root	ORG	2.85 b	0.88 a	0.19 a
	CONV	4.64 a	0.59 a	0.15 a
whole plant			63.58 a	6.25 a
			40.74 b	4.09 b

493

494

495 **Figure captions**

496

497 Figure 1 - Experimental sites in the thermal area of Bullicame (Baiocchi et al. 2013,
498 modified)

499

500 Figure 2 - Microbial population based on EL-FAME profiles under the two different
501 managements. G+= positive bacteria; G- = negative bacteria; Bacterial General = non-specific
502 bacteria; Fungi; Act = Actinomycetes; Protozoa. Microbial groups are assigned to ester linked
503 fatty acids as reported in de Dato et al. (2017). Values belonging to the same parameter
504 between systems with different letters are statistically different according to LSD (0.05).

505

506

Figure 1

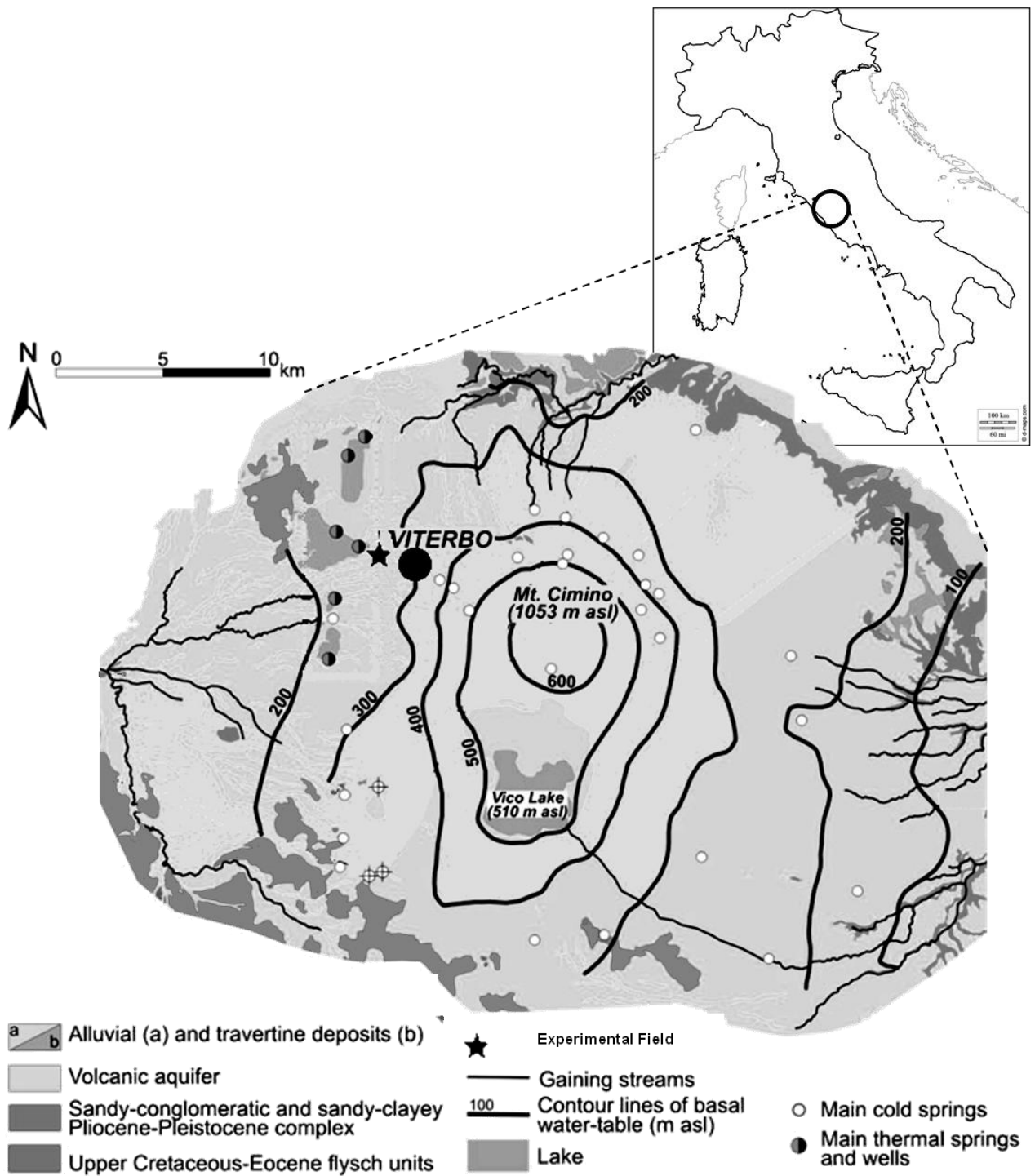


Figure 2

