

1 **High resolution short-term investigation of soil CO₂, N₂O, NO_x and NH₃ emissions after different**
2 **chabazite zeolite amendments**

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32 **Abstract**

33 Reducing the effects of climate change is a key-point to achieve sustainable development. It is known that fertilizer
34 application stimulates soil gaseous N losses, especially immediately after their application. In this short-term incubation
35 experiment, the effects of different chabazite zeolite amendments on soil gaseous emissions (CO₂, N₂O, NO_x and NH₃)
36 were evaluated in high resolution as potential mitigation strategy for agricultural soils. Different soil-zeolite mixtures,
37 including both natural and NH₄⁺-enriched chabazite zeolites, were incubated for 24 h both immediately after the
38 application of urea fertilizer and without a further N input in order to carry out a high resolution investigation of soil
39 CO₂, N₂O, NO_x and NH₃ fluxes, in comparison to an unamended soil. Immediate CO₂, N₂O, NO_x and especially NH₃
40 emissions after fertilizer application were generally reduced in soils amended with zeolites at natural state, indicating a
41 potential valuable material for reducing soil C-N gaseous losses. On the other hand, the application of NH₄⁺-enriched
42 zeolites lowered CO₂ and N₂O emissions, but very high NO_x fluxes occurred during the first 24 h even without applying
43 any further N input. NH₃ emissions were higher in NH₄⁺-enriched zeolites amended soil, but if the amendment is
44 performed without further N inputs, the emissions can be significantly lowered with respect to a conventional urea
45 fertilization.

46

47 **Keywords:** *natural zeolites; NH₃; CO₂; N₂O; NO_x; soil emissions.*

48

49 **1.0 Introduction**

50 Notwithstanding the increasing number of climate change mitigation policies, the annual emissions of greenhouse gases
51 (GHGs) keeps rising, leading to high costs in terms of environmental quality (Edenhofer et al., 2014). In particular, the
52 agricultural sector accounts for an estimated emission of 10-12 % of the total global anthropogenic GHGs production
53 (Smith et al., 2007).

54 CO₂ and N₂O are potent GHGs whereby N₂O has a 298-fold global warming potential “GWP” of CO₂ (Edenhofer et al.,
55 2014). In addition, nitrogen oxides (NO_x) and ammonia (NH₃), even if not considered GHGs, can cause indirect N₂O
56 emissions, increase acidity of precipitations and strongly influence tropospheric O₃ formation (Akiyama et al., 2004;
57 Schindlbacher and Zechmeister-Boltenstern, 2004). Moreover, all N gaseous losses cause remarkable reductions in
58 fertilization efficiency, representing thus an economic loss for the farmers (Smil, 1999).

59 It is known that the great majority of gaseous N losses occurs during the first days after fertilizer application and these
60 “immediate” direct flux rates are not completely known, in particular, NH₃ emission have been found to account up to
61 75 % of the applied N within the first 24 h (Stevens and Logan, 1987). For this reason, finding new methodologies that

62 can delay immediate gaseous N-losses may lead to an increase of fertilizer use efficiency and thus to a reduction of their
63 environmental impacts.

64 Since many years, natural zeolites have been recognized as valuable soil conditioners that can positively affect the soil-
65 plant system. These aluminosilicates are characterized by a high and selective cation exchange capacity (CEC),
66 reversible dehydration and molecular sieving that can strongly influence soil physico-chemical characteristics and N
67 cycling by reducing N leaching, increasing nitrogen use efficiency and crop yield (Colombani et al., 2014; De Campos
68 Bernardi et al., 2013; Gholamhoseini et al., 2013; Malferrari et al., 2013; Reháková et al., 2004). Zeolites can be easily
69 modified from their natural state by enrichment processes which cause the adsorption of specific cations (i.e., NH_4^+ ,
70 Na^+) (Barbarick et al., 1990; Campisi et al., 2016; Dwairi, 1998; Leggo, 2000), for this reason, the use of NH_4^+ -enriched
71 zeolite has been proposed as “slow release fertilizer” (Barbarick and Pirela 1984; Dwairi, 1998; Faccini et al., 2015;
72 Lewis et al. 1984). Up to now, however, very few studies exist on the effects of zeolite amendments on soil gaseous
73 emission especially concerning natural zeolite of the chabazite type. Natural clinoptilolite zeolite as amendment to soils
74 have been described to reduce respiratory activity probably because of the adsorption capacity of zeolites for CO_2
75 (Mühlbachová and Šimon, 2003). Kučić et al. (2013) showed that natural clinoptilolite zeolite adsorption potential for
76 CO_2 and NH_3 evolved during co-composting of grape and tobacco waste was 31% and 100%, respectively. In addition,
77 pasture and wetland soils amended with clinoptilolite zeolite showed a reduction in daily and total N_2O emissions
78 during a 90-day incubation compared to unamended soils (Zaman et al., 2007). These reductions are due to the sorption
79 of NH_4^+ by zeolite, lowering the available substrate for microbial nitrification and hence potential losses of N_2O from
80 nitrification and denitrification. The literature about the mitigation potential of zeolite application for NH_3 emissions is
81 more consistent (Bundan et al., 2011; De Campos Bernardi et al., 2013; Haruna Ahmed et al., 2008; He et al., 2002;
82 Kučić et al., 2013), however, the majority of these studies has been conducted on surface applied zeolite in combination
83 with fertilizer and not as amendment well homogenized in the soil profile. Yet, to our best knowledge, the effects of
84 zeolite as soil amendments on NO_x emissions have not been reported. Besides, the majority of studies were conducted
85 employing clinoptilolite zeolites, with so far unknown effects of the less abundant chabazite zeolite, the main natural
86 zeolite occurring in the Italian peninsula (Malferrari et al., 2013). Moreover, the effects of NH_4^+ -enriched zeolites on
87 soil gaseous emissions are completely unknown and usually the N input added with this material is very high (Ferretti et
88 al., 2017). Finally, there is a lack of studies applying different type of zeolites as soil amendment that simultaneously
89 investigate a wide range of gaseous species focusing on a high resolution investigation of the immediate emission after
90 fertilizer applications.

91 To this end, the present study aims at elucidating the effects of different chabazite zeolites amendments on soil CO₂,
92 N₂O, NO_x and NH₃ emissions evolved during the first 24 h after fertilizer application from an agricultural soil in high
93 resolution. The experimentation was designed in order to mimic the same treatment and condition of the ZeoLIFE
94 experimental field, which consist in an EU funded project that is testing different chabazite zeolite amendment at the
95 field scale in an Italian nitrate vulnerable zone (NVZ) (Nitrate Directive 91/676/EEC).

96 For this reason, we decided to test chabazite zeolites amendments both at natural and NH₄⁺-enriched state in comparison
97 to an unamended soil both under an N input from urea of 170 kg N ha⁻¹ (maximum amount of N applicable in a NVZ)
98 and without any further N input (unfertilized conditions). Considering the actual knowledge about zeolite amendments,
99 we builded our experiment in order to test the following hypotheses:

- 100 1) The zeolite-derived effects on the dynamics of NH₄⁺ ions (Leyva-Ramos et al., 2010) are expected to decrease
101 short-term N₂O, NO_x and NH₃ emissions after fertilizer application especially in soil amended with zeolites at
102 natural states;
- 103 2) The controlled slow NH₄⁺ release behavior of NH₄⁺-enriched chabazite zeolites is expected to not influence
104 significantly soil N fluxes in the first 24 h, if no further fertilizer is applied notwithstanding its high N content;
- 105 3) The possibility of a CO₂ sorption and the probable influence on microbial activity (Bonenfant et al., 2008;
106 Kučić et al., 2013; Mühlbachová and Šimon, 2003; Reháková et al., 2004; Ridha and Webley, 2009) are
107 expected to decrease the short-term soil CO₂ emissions.

108

109

110 **2.0 Material and Methods**

111 Soil samples (Clayey-Silt Calcaric Gleyic Cambisol) were taken from the ZeoLIFE project experimental field (Ferrara
112 province, Italy, 44°50'33" N and 12°05'40"E) where both natural chabazite zeolites (NZ) and NH₄⁺-enriched chabazite
113 zeolites (CZ) (created following the method proposed by Faccini et al., 2015) were added to the soil as amendments at
114 the field scale (www.zeolife.it) (see Electronic Supplementary Material "ESM1" for the main soil properties and
115 additional information on zeolites characteristics). Soil samples were taken from an unamended plot and zeolites were
116 added in the laboratory immediately before the begin of the experiment, in order to reproduce the same conditions
117 immediately after the amendment. NZ and CZ were added to air dry sieved soil (≤ 5 mm) in different proportions, in
118 order to obtain various soil-zeolite mixtures (see Table 1 for the experimental set up) as in the ZeoLIFE experimental
119 field.

120 60 g of each treatment were incubated at 20°C (well representative of average field temperatures) for 24 h in three
121 replicates. Water filled pore space (WFPS) level was adjusted according to Haney and Haney (2010) to 65 %
122 accounting for changes in bulk density after the amendment. This WFPS level is considered a threshold between the
123 relative prevalence of nitrification/denitrification processes (Bateman and Baggs, 2005). In order to check the moisture
124 level reached, soil water content was also measured by oven-drying soils at 105°C overnight and the difference between
125 the adjusted and measured WFPS was < 5 %. No rewetting operations were performed during the incubation.

126 The experiment was conducted with and without urea input equivalent to 170 kg-N ha⁻¹, (in liquid form, maintaining 65
127 % WFPS) representing fertilized and unfertilized conditions, respectively. The employed fertilizer rate was the same as
128 in the ZeoLIFE experimental field. These high application rates are common in countries such as Italy (in which
129 fertilization can reach more than 340 kg ha⁻¹ in non-nitrate vulnerable zones) or even lower in comparison to some
130 studies available in the scientific literature (e.g. Min et al., 2011; Ju et al., 2006) in which more than 600 kg N ha⁻¹ have
131 been applied. We choose to reproduce the maximum admissible rates of the nitrate vulnerable zone where the ZeoLIFE
132 experimental field site is located in order to get insights on the amounts of N losses immediately after the application of
133 chemical fertilizers like urea with and without the use of zeolites as soil amendment in this particular area.

134 The 10CZ treatment that was applied in the ZeoLIFE field experiment was also reproduced in the present study
135 providing an N input equivalent to approximately 497 kg N ha⁻¹. Although, the applied N is unbalanced if compared to
136 the other treatments, this allowed to test the behaviour in presence of an excess of N above the recommendation.

137 This N input from CZ was mainly constituted by exchangeable NH₄⁺-N and by a little fraction of residual NO₃⁻-N from
138 the enrichment process (ESM 1). The 10CZ treatment was tested with and without urea fertilizer, consequently it is
139 further termed as “unfertilized” when no urea was added, while it is further termed “fertilized” when it received the
140 additional 170 kg N ha⁻¹ from urea as all the other treatments.

141 CO₂ and NO_x (NO + NO₂) were measured in a fully automated continuous flow laboratory system, well described in the
142 work of Schindlbacher and Zechmeister-Boltenstern (2004), with doubled acquisition time of each chamber. N₂O was
143 sampled by closing the chambers and taking samples with a gas tight syringe after 0, 15, 30 and 45 minutes of the
144 closure of the chamber. Samples were injected in a pre-evacuated 10 ml glass vials (Agilent Technologies) and then
145 analysed with a GC-System (Agilent Technologies) equipped with a headspace auto-sampler and an electron capture
146 detector (ECD), for more information see also Leitner et al. (2016). N₂O measurements were performed once after
147 approx. 4 hours of incubations and fluxes were calculated according to Metcalfe et al. (2007).

148 NH₃ measurements were conducted with a fully automated device composed of an incubation system with 6 chambers
149 (ø 7.8 cm and 7.8 cm height) connected to a Cavity Ring-Down Spectrometer (CRDS, Picarro G2103). Chambers and

150 tubes of the incubation device are built of Polytetrafluoroethylene (PTFE) in order to avoid any NH₃ retention. Dried
151 compressed air was used as carrier gas with a constant flow of 1 l min⁻¹. One chamber was used as blank in order to
152 measure NH₃ background air concentrations (Haller, 2015).

153 CO₂, NO_x and NH₃ fluxes were calculated according to Schindlbacher and Zechmeister-Boltenstern (2004) and
154 represented as cumulative emissions by integrating over the 24 h of incubation. Moreover, we considered also the
155 emissions per unit of N added by calculating the ratio between the amount of gas emitted divided by the total N added.

156 NO₃⁻-N was extracted with Milly-Q (Millipore USA) water in a 1:10 (w/v) ratio, the solution was shaken for 1 h and
157 then filtered. NO₃⁻-N was determined by ion chromatography with an isocratic dual pump (ICS-1000 Dionex), equipped
158 with an AS9-HC 4 × 250 mm high-capacity column and an ASRS-Ultra 4-mm self-suppressor. An AS-40 Dionex auto-
159 sampler was employed to run the analysis; quality control (QC) samples were run every ten samples. pH was measured
160 on the same extracts with an Orion 9102BNWP pH-meter connected to an Orion 4star pH – ISE benchtop (Thermo
161 Fisher).

162 Exchangeable NH₄⁺-N was extracted with 1 M KCl in a 1:10 (w/v) ratio, the solution was shaken for 1 h and then
163 filtered. The solution was diluted and analysed with an Ion Selective Electrode (ISE) Orion 95-12 connected to an Orion
164 4star pH – ISE benchtop (Thermo Fisher).

165 Shapiro-Wilk and Levene's Test were performed for testing data normality and homogeneity of variance. One-way
166 ANOVA (or Kruskal-Wallis test if ANOVA assumption were not met) and Fisher (LSD) tests were then employed for
167 evaluating significant differences between the treatments at p=0.05 level. SigmaPlot 12.0 was used for statistical
168 analysis.

169

170 **3.0 Results and discussions**

171 Cumulative CO₂, NO_x and NH₃ emissions over the 24 h are presented in Fig 1. Soil pH, exchangeable NH₄⁺-N, NO₃⁻-N
172 mean values and N₂O-N fluxes are visible in Table 2, while the total amounts of volatilized gaseous species (expressed
173 in kg N ha⁻¹), the reduction/increment % with respect to the CNTR and the effects of the fertilization rates on
174 cumulative gaseous losses are shown in Table 3. The total amounts of volatilized gaseous species (expressed in kg N ha⁻¹
175 ¹), the reduction/increment % with respect to the CNTR of the treatments with no N added from fertilization (or from
176 CZ) are visible in the Electronic Supplementary Material "ESM2".

177 All zeolite amended soils (see Table 1 for treatments description) resulted in significantly lower (p<0.05) CO₂
178 emissions compared to the unamended soil (CNTR) under fertilized conditions (Fig 1a). These preliminary results show
179 a reduction in CO₂ emissions with the addition of zeolites regardless of zeolite type and application rate, and may

180 indicate a CO₂ sorption effect by zeolites after the fertilization (Kučić et al., 2013; Mühlbachová and Šimon, 2003) or,
181 alternatively, to a higher Carbon Use Efficiency (CUE) (Blagodatskaya et al., 2014; Keiblinger et al., 2010). On the
182 other hand, regarding CO₂ emissions under unfertilized conditions (Fig 1b), the fluxes were not significantly different
183 between the various treatments ($p>0.05$).

184 It is possible that, without the addition of urea, the substrate C:N ratio was higher, resulting in a higher amount of
185 respired CO₂ to meet metabolic requirements (Russell and Cook, 1995; Schimel and Weintraub, 2003) and hence a
186 lower CUE (Keiblinger et al., 2010). In the present work, it has to be noticed that cumulative CO₂ emissions were
187 similar between fertilized and unfertilized conditions, highlighting that microbial activity was high in this agricultural
188 soil even without adding a fresh N-C input. Concerning this aspect, contrasting effects were reported in the scientific
189 literature; while urea fertilization was found to increase soil CO₂ emissions because of increased mineralization of soil
190 organic carbon by heterotrophic bacteria (Choi et al., 2011), the opposite effect was reported by Wilson and Al-Kaisi
191 (2008) after fertilization with NH₄NO₃. However, the reason why no significant differences in CO₂ emissions were
192 observed under unfertilized conditions is hard to explain without a more detailed investigation on treatment specific
193 microbial CUE and CO₂ adsorption mechanisms. Concomitantly, also the cation exchange processes between the soil
194 solution and zeolite minerals were probably different without and with the addition of fertilizers. It is in fact widely
195 known that the CO₂ adsorption capacity of zeolites depends on many factors including also the distribution and the
196 number of exchangeable cations presents in their cavities (Bonenfant et al., 2008).

197 The cumulative soil CO₂ emissions evolved in 24 h have been reduced by ~21 % to approximately ~30 % with respect
198 to the CNTR soil in presence of urea fertilizer, coherently, the CNTR showed the highest emissions in relation to the
199 total amount of N added (Table 3), even if compared to the very high amount of N supplied by CZ.

200 N₂O fluxes of fertilized soils were significantly lower ($p<0.05$) in both 5NZ (soil plus 5% in weight of NZ) and 10CZ
201 (soil plus 10 % in weight of CZ) treatments with respect to CNTR, while 15NZ treatment (soil plus 15% in weight of
202 NZ) tended to be slightly lower albeit not statistically different (Table 2). N₂O is produced by both nitrification and
203 denitrification processes, where the main substrate for nitrification is NH₄⁺ while for denitrification is NO₃⁻. However,
204 these processes are strongly influenced by WFPS level, substrate and oxygen availability (Akiyama et al., 2004;
205 Schindlbacher and Zechmeister-Boltenstern, 2004; Stevens et al., 1997). The WFPS used in this experiment should be
206 the critical threshold between these two processes (Bateman and Baggs, 2005). Since WFPS and oxygen availability are
207 closely related, the observed differences regarding N₂O emissions can be mainly attributed to a change in soil physical
208 properties operated by zeolite amendments. Zeolite amendment (in a 3-5 mm size) has probably increased soil aeration
209 and thus reduced the presence of anaerobic microsites in which denitrification can occur at 65 % WFPS, consequently

210 reducing total N₂O emissions with respect to CNTR. Another explanation can be that the NH₄⁺ adsorption from soil
211 solution, managed especially by the initially N-devoid NZ, might have reduced the immediate availability of the
212 substrate required for nitrification, thus reducing N₂O emissions (Zaman et al., 2007). The latter hypothesis is supported
213 by our results from NZ amendments where in fact NO₃⁻-N was generally lower, but not for CZ amendment, where on the
214 contrary NO₃⁻-N was strongly higher than in CNTR (Table 2).

215 NO_x are intermediate reactive volatile compounds of both nitrification and denitrification processes and mainly
216 produced after the addition of fertilizers (Skiba et al., 1997). Coherently, the amount of N lost as NO_x was considerably
217 higher after urea application compared to unfertilized conditions (Fig 1c,d). Cumulative NO_x emissions after
218 fertilization were significantly lower (p<0.05) in 5NZ and 15NZ treatments compared to CNTR (Fig 1c). This evidence
219 can be partially explained by the lower NO₃⁻-N contents in both 5NZ and 15NZ treatments, which suggests that some
220 NH₄⁺ ions were probably protected from nitrification (Table 2) in this time frame (Zaman et al., 2007). The entity of the
221 reduction of the 5NZ and 15NZ with respect to the CNTR soil under fertilized conditions ranged from ~ 25 - 31 %
222 (Table 3).

223 Generally, urea fertilization caused an equal increase of the NO₃⁻-N level of around 40 mg kg⁻¹ in all the treatments.
224 However, the NO₃⁻-N content of 10CZ soil was extremely high both with and without the addition of urea fertilizer,
225 meaning that probably part of the N supplied with CZ has been subjected to immediate nitrification after the application
226 to the soil. These high NO₃⁻-N levels in the short-term period needs to be taken into account for potential immediate
227 effects on N leaching from the soil system. However, concerning NO_x emissions, it has to be noticed that
228 notwithstanding the excessive N level of this treatment (CZ + urea), the cumulative emissions were not significantly
229 different from CNTR and both NZ treatments (p>0.05), thus the total amounts of NO_x emitted per unit of N added
230 result considerably lower (Table 3). On the other hand, 10CZ NO_x emissions of the unfertilized soils (Fig 1d) were
231 comparable to the fertilized one and not statistically different from the fertilized CNTR although a 3-fold N amount was
232 applied to the same soil (Table 3). While for the other unfertilized treatments, NO_x emissions were very low as
233 expected. These higher emissions in 10CZ can be attributed to an immediate transformation of the N supplied by the
234 already N-enriched CZ. In fact, the extremely high NO₃⁻-N contents suggests strongly enhanced nitrification (biotic or
235 abiotic) in this particular treatment. Commonly, NH₄-enriched zeolites are considered as “slow release” fertilizers
236 (Barbarick and Pirela 1984; Dwairi, 1998; Lewis et al. 1984) but Leggo (2000) found a strong increase in NO₃⁻-N
237 content immediately after the amendment with NH₄-enriched zeolites (starting from the first week). In that work, the
238 increase in NO₃⁻-N was attributed to an exchange reaction with soil cations (which have substituted the NH₄⁺ inside the
239 mineral lattice) and subsequently NH₄⁺ oxidation by nitrifying bacteria. This may have occurred also in the present

240 study since the soil is characterized by a high natural salinity (Di Giuseppe et al., 2014) and a high CEC, which has
241 been further increased by the zeolite amendment (Ferretti et al., 2017; Gholamhoseini et al., 2013).

242 Considering the fact that without addition of urea NO_3^- -N levels and NO_x emissions increased significantly with CZ
243 application to the soil with respect to the CNTR, this material likely supplied an immediate available N source so that
244 CZ should be considered as a fertilizer. However, considering the extremely higher N input from CZ (~ 3 times more
245 than the N added from urea), the emission evolved from CZ treated soil in relation to the amount of N added were
246 always lower with respect to an untreated soil (Table 3).

247 In the present study, NH_3 emissions evolved within the first 24 h were comparable with those observed by other authors
248 in the literature (Bundan et al., 2011). Soil NH_3 emissions are known to be highly variable and dependent by many
249 factors such as soil pH, wind speed, soil moisture and CEC (Bundan et al., 2011). Notwithstanding the soil pH was sub-
250 alkaline in all the treatments (Table 2) and thus favourable to NH_3 volatilization, the relatively low NH_3 losses occurred
251 in all the incubated soils may be due to the high soil CEC that, even without the addition of zeolites, can act a good
252 retention of NH_4^+ ions reducing NH_3 volatilization phenomena (Bundan et al., 2011).

253 In both fertilized and unfertilized conditions, a clear reduction in NH_3 emissions was found (Fig. 1e-f) in NZ treatments
254 with respect to CNTR ($p < 0.05$). The entity of reduction after fertilization was of -24 and -55% in 5NZ and 15NZ,
255 respectively, while in unfertilized conditions the reduction was of -40 and -60 % respectively (Table 3). This behaviour
256 is attributable to the NH_4^+ ions adsorption inside NZ, which can physically protect NH_4^+ from NH_3 volatilization (De
257 Campos Bernardi et al., 2013) and it is related to the amount of NZ added in the substrate. However, 10CZ treatment
258 showed high NH_3 emissions in both fertilized and unfertilized conditions, suggesting that some residual NH_4^+ ions from
259 the enrichment process were available for volatilization. It is interesting to note that for all the other gaseous species,
260 10CZ (without further N input from urea), showed higher or similar emissions with respect to the urea-fertilized CNTR
261 (Fig 1a,c), while regarding cumulative NH_3 emissions, those of urea-fertilized CNTR were about twice of unfertilized
262 10CZ (Fig 1e-f, Table 3) notwithstanding a three times lower application rate. This means that urea-N contribute much
263 more to NH_3 emissions than CZ-N and that even applying a considerably higher N input, the losses by NH_3
264 volatilization can be reduced by the half.

265 Finally, it has to be noticed that generally, the ratio between total emissions of each gaseous species and the amounts of
266 N added was always higher in the CNTR soil, meaning that zeolite addition (even in presence of a great excess of N
267 from fertilization) may have important positive effects on gaseous N (and C) losses and consequently also on
268 fertilization efficiency.

269

270 **4.0 Conclusions**

271 The outcomes of this high-resolution short-term incubation experiment highlighted the potential of zeolite application
272 as soil amendment for the reduction of immediate gaseous N (and C) emissions. Results revealed that after 24 h of
273 monitoring, NZ amendments seemed to reduce the immediate production of CO₂, N₂O, NO_x, and especially NH₃
274 emissions after the application of urea fertilizer, constituting thus a potential valuable material for reducing short-term
275 soil gaseous C and N losses. The amount of NZ applied (5 % or 15 % in weight) had no influences on CO₂, N₂O and
276 NO_x emissions, while the cumulative emissions of NH₃ were inversely related to the amount of NZ in the substrate.

277 On the other hand, the outcomes of this study revealed that NH₄⁺-enriched zeolites, created following the ZeoLIFE
278 project protocol, supplied a fraction of N that was immediately subjected to gaseous losses. CZ amendment lowered
279 CO₂ and N₂O emissions evolved during the first 24 h after fertilizer application, however, nitrification was stimulated
280 and NO_x emission were significantly high even without the addition of urea fertilizer, indicating that part of the N
281 supplied with CZ was immediately transformed biotically or abiotically. The emissions per unit of N added were
282 generally higher in the CNTR soil, meaning that zeolite addition (even in presence of a great excess of N from
283 fertilization) may play an important role in GHG and non-GHG mitigation in agricultural context and consequently also
284 on fertilization efficiency.

285 We are aware of the fact that our short-term, high resolution experiment is limiting the potential to draw conclusions on
286 long-term mitigation strategies, in terms of immediate gaseous N losses, for this reason, further long-term investigations
287 are required.

288

289 **Acknowledgements**

290 The authors are grateful to Helmut Haller and Dr. Christian Holtermann for their help during laboratory procedures and
291 analysis. Special thanks to the anonymous reviewer for very helpful comments to improve this manuscript. This work
292 has been co-funded by the EU-funded ZeoLIFE project (LIFE+10/ENV/IT/000321) and by UNIFE young researchers
293 grant 2015. It is related to the EU-funded Eclairé project (FP7-Env.2011.1.1.2-1 282910) and to the project NitroAustria
294 KR14AC7K11916.

295

296

297 **References**

298 Akiyama, H., McTaggart, I.P., Ball, B.C., Scott, A., 2004. N₂O, NO, and NH₃ emissions from soil after the application
299 of organic fertilizers, urea and water. *Water Air Soil Pollut.* 156, 113–129.

300 Barbarick, K.A., Lai, T.M., Eberl, D.D., 1990. Exchange Fertilizer (Phosphate Rock plus Ammonium-Zeolite) Effects
301 on Sorghum-Sudangrass. *Soil Sci. Soc. Am. J.*

302 Barbarick, K.A., Pirela, H.J., 1984. Agronomic and horticultural uses of zeolites: a review. In: Pond WG and Mumpton
303 FA (eds). *Zeo-Agriculture: Use of Natural Zeolites in Agriculture and Aquaculture*, International Committee on
304 Natural Zeolites, pp 93-103.

305 Bateman, E.J., Baggs, E.M., 2005. Contributions of nitrification and denitrification to N₂O emissions from soils at
306 different water-filled pore space. *Biol. Fertil. Soils* 41, 379–388.

307 Blagodatskaya, E., Blagodatsky, S., Anderson, T.H., Kuzyakov, Y., 2014. Microbial growth and carbon use efficiency
308 in the rhizosphere and root-free soil. *PLoS One* 9.

309 Bonenfant, D., Kharoune, M., Niquette, P., Mimeault, M., Hausler, R., 2008. Advances in principal factors influencing
310 carbon dioxide adsorption on zeolites. *Sci. Technol. Adv. Mater.* 9, 1468–6996.

311 Bundan, L., Majid, N.M.A., Ahmed, O.H., Jiwan, M., Kundat, F.R., 2011. Ammonia volatilization from urea at
312 different levels of zeolite. *Int. J. Phys. Sci.* 6, 7717–7720.

313 Campisi, T., Abbondanzi, F., Faccini, B., Di Giuseppe, D., Malferrari, D., Coltorti, M., Laurora, A., Passaglia, E., 2016.
314 Ammonium-charged zeolite effects on crop growth and nutrient leaching: Greenhouse experiments on maize (*Zea*
315 *mays*). *Catena* 140, 66–76. d

316 Choi, W.-J., Matushima, M., Ro, H.-M., 2011. Sensitivity of soil CO₂ emissions to fertilizer nitrogen species: urea,
317 ammonium sulfate, potassium nitrate, and ammonium nitrate. *J. Korean Soc. Appl. Biol. Chem.* 54, 1004–1007.

318 Colombani, N., Mastrocicco, M., Di Giuseppe, D., Faccini, B., Coltorti, M., 2014. Variation of the hydraulic properties
319 and solute transport mechanisms in a silty-clay soil amended with natural zeolites. *Catena* 123, 195–204.

320 De Campos Bernardi, A.C., Anchão Oliviera, P.P., De Melo Monte, M.B., Souza-Barros, F., 2013. Brazilian
321 sedimentary zeolite use in agriculture. *Microporous Mesoporous Mater.* 167, 16–21.

322 Di Giuseppe, D., Faccini, B., Mastrocicco, M., Colombani, N., Coltorti, M., 2014. Reclamation influence and
323 background geochemistry of neutral saline soils in the Po River Delta Plain (Northern Italy). *Environ. Earth Sci.* 72,
324 2457–2473.

325 Dwairi, I.M., 1998. Evaluation of jordanian zeolite tuff as a controlled slow-release fertilizer for NH₄⁺. *Environ. Geol.*
326 34, 1–4.

327 Edenhofer, O., Pichs-Madruga, R., Sokona, Y., Minx, J.C., Farahani, E., Susanne, K., Seyboth, K., Adler, A., Baum, I.,
328 Brunner, S., Eickemeier, P., Kriemann, B., Savolainen, J., Schlomer, S., von Stechow, C., Zwickel, T., 2014.

329 Climate Change 2014: Mitigation of Climate Change, Working Group III Contribution to the Fifth Assessment
330 Report of the Intergovernmental Panel on Climate Change.

331 Faccini, B., Di Giuseppe, D., Malferrari, D., Coltorti, M., Abbondanzi, F., Campisi, T., Laurora, A., Passaglia, E., 2015.
332 Ammonium-exchanged zeolite preparation for agricultural uses: From laboratory tests to large-scale application in
333 ZeoLIFE project prototype. *Period. di Mineral.* 84, 303–321.

334 Ferretti, G., Di Giuseppe, D., Natali, C., Faccini, B., Bianchini, G., Coltorti, M. 2017. C-N elemental and isotopic
335 investigation in agricultural soils: Insights on the effects of zeolite amendments. *Chem Erde-Geochem.*
336 <http://dx.doi.org/10.1016/j.chemer.2017.02.002>

337 Gholamhoseini, M., Ghalavand, A., Khodaei-Joghan, A., Dolatabadian, A., Zakikhani, H., Farmanbar, E., 2013.
338 Zeolite-amended cattle manure effects on sunflower yield, seed quality, water use efficiency and nutrient leaching.
339 *Soil Tillage Res.* 126, 193–202.

340 Haller, H., 2015. Ammonia emissions from beech litter as source of reactive nitrogen gas in deciduous forests. Master
341 thesis, Universität für Bodenkultur, Wien.

342 Haney, R.L., Haney, E.B., 2010. Simple and Rapid Laboratory Method for Rewetting Dry Soil for Incubations.
343 *Commun. Soil Sci. Plant Anal.* 41, 1493–1501.

344 Haruna Ahmed, O., Husin, A., Husni Mohd Hanif, A., 2008. Ammonia volatilization and ammonium accumulation
345 from urea mixed with zeolite and triple superphosphate. *Acta Agric. Scand. Sect. B - Plant Soil Sci.* 58, 182–186.

346 He, Z.L., Calvert, D. V., Alva, A.K., Li, Y.C., Banks, D.J., 2002. Clinoptilolite zeolite and cellulose amendments to
347 reduce ammonia volatilization in a calcareous sandy soil. *Plant Soil* 247, 253–260.

348 Ju, X.T., Kou, C.L., Zhang, F.S., Christie, P., 2006. Nitrogen balance and groundwater nitrate contamination:
349 comparison among three intensive cropping systems on the North China Plain. *Environmental Pollution*, 143(1),
350 117-125.

351 Keiblinger, K.M., Hall, E.K., Wanek, W., Szukics, U., Hämmerle, I., Ellersdorfer, G., Böck, S., Strauss, J., Sterflinger,
352 K., Richter, A., Zechmeister-Boltenstern, S., 2010. The effect of resource quantity and resource stoichiometry on
353 microbial carbon-use-efficiency. *FEMS Microbiol. Ecol.* 73, 430–440.

354 Kučić, D., Kopčić, N., Briški, F., 2013. Zeolite and potting soil sorption of CO₂ and NH₃ evolved during co-
355 composting of grape and tobacco waste. *Chem. Pap.* 67, 1172–1180.

356 Leggo, P.J., 2000. An investigation of plant growth in an organo-zeolitic substrate and its ecological significance. *Plant*
357 *Soil* 219, 135–146.

358 Leitner, S., Sae-Tun, O., Kranzinger, L., Zechmeister-Boltenstern, S., Zimmermann, M., 2016. Contribution of litter
359 layer to soil greenhouse gas emissions in a temperate beech forest. *Plant Soil* 1–15.

360 Lewis, M.D., Moore, F.D., Goldsberry, K.L., 1985. Ammonium-exchanged clinoptilolite and granulated clinoptilolite
361 with urea as nitrogen fertilizers. In: Pond WG and Mumpton FA (eds), *Zeo-Agriculture: Use of Natural Zeolites in*
362 *Agriculture and Aquaculture*; International Committee on Natural Zeolites 105-111.

363 Leyva-Ramos, R., Monsivais-Rocha, J.E., Aragon-Piña, A., Berber-Mendoza, M.S., Guerrero-Coronado, R.M.,
364 Alonso-Davila, P., Mendoza-Barron, J., 2010. Removal of ammonium from aqueous solution by ion exchange on
365 natural and modified chabazite. *J. Environ. Manage.* 91, 2662–2668.

366 Malferrari, D., Laurora, A., Brigatti, M.F., Coltorti, M., Di Giuseppe, D., Faccini, B., Passaglia, E., Vezzalini, M.G.,
367 2013. Open-field experimentation of an innovative and integrated zeolitite cycle: Project definition and material
368 characterization. *Rend. Lincei* 24, 141–150.

369 Metcalfe, D.B., Meir, P., Aragão, L.E.O.C., Malhi, Y., da Costa, A.C.L., Braga, A., Gonçalves, P.H.L., de Athaydes,
370 J., de Almeida, S.S., Williams, M., 2007. Factors controlling spatio-temporal variation in carbon dioxide efflux from
371 surface litter, roots, and soil organic matter at four rain forest sites in the eastern Amazon. *J. Geophys. Res.*
372 *Biogeosciences* 112, 1–9.

373 Min, C., Xuecheng, S., Chengxiao, H., Hong J.D., Qiling, T., Changsheng Z., 2011. Effective mitigation of nitrate
374 leaching and nitrous oxide emissions in intensive vegetable production systems using a nitrification inhibitor,
375 dicyandiamide. *J Soil Sediments.* 11, 722:730.

376 Mühlbachová, G., Šimon, T., 2003. Effects of zeolite amendment on microbial biomass and respiratory activity in
377 heavy metal contaminated soils. *Plant, Soil Environ.* 49, 536–541.

378 Reháková, M., Čuvanová, S., Dzivák, M., Rimár, J., Gaval'ová, Z., 2004. Agricultural and agrochemical uses of
379 natural zeolite of the clinoptilolite type. *Curr. Opin. Solid State Mater. Sci.* 8, 397–404.

380 Ridha, F.N., Webley, P.A., 2009. Anomalous Henry's law behavior of nitrogen and carbon dioxide adsorption on alkali-
381 exchanged chabazite zeolites. *Sep. Purif. Technol.* 67, 336–343.

382 Russell, J.B., Cook, G.M., 1995. Energetics of bacterial growth: balance of anabolic and catabolic reactions. *Microbiol.*
383 *Rev.* 59, 48–62.

384 Schimel, J.P., Weintraub, M.N., 2003. The implications of exoenzyme activity on microbial carbon and nitrogen
385 limitation in soil: A theoretical model. *Soil Biol. Biochem.* 35, 549–563.

386 Schindlbacher, A., Zechmeister-Boltenstern, S., 2004. Effects of soil moisture and temperature on NO, NO₂, and N₂O
387 emissions from European forest soils. *J. Geophys. Res.* 109, 1–12.

- 388 Skiba, U., Fowler, D., Smith, K. a, 1997. Nitric oxide emissions from agricultural soils in temperate and tropical
389 climates: sources, controls and mitigation options. *Nutr. Cycl. Agroecosystems* 48, 139–153.
- 390 Smil, V., 1999. Nitrogen in crop production: An account of global flows. *Global Biogeochem. Cycles* 13, 647–662.
- 391 Smith, P., Martino, D., Cai, Z., Gwary, D., Janzen, H., Kumar, P., McCarl, B., Ogle, S., O’Mara, F., Rice, C., Scholes,
392 B., Sirotenko, O., 2007. Agriculture. In *Climate Change 2007: Mitigation, Contribution of Working Group III to the*
393 *Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, in: *Climate Change 2007:*
394 *Mitigation. Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on*
395 *Climate Change*. pp. 497–540.
- 396 Stevens, R.J., Laughlin, R.J., Burns, L.C., Arah, J.R.M., Hood, R.C., 1997. Measuring the contributions of nitrification
397 and denitrification to the flux of nitrous oxide from soil. *Soil Biol. Biochem.* 29, 139–151.
- 398 Stevens, R.J., Logan, H.J., 1987. Determination of the volatilization of ammonia from surface-applied cattle slurry by
399 the micrometeorological mass balance method. *J. Agric. Sci.* 109, 205.
- 400 Wilson, H.M., Al-Kaisi, M.M., 2008. Crop rotation and nitrogen fertilization effect on soil CO₂ emissions in central
401 Iowa. *Appl. Soil Ecol.* 39, 264–270.
- 402 Zaman, M., Nguyen, M.L., Matheson, F., Blennerhassett, J.D., Quin, B.F., 2007. Can soil amendments (zeolite or lime)
403 shift the balance between nitrous oxide and dinitrogen emissions from pasture and wetland soils receiving urine or
404 urea-N? *Aust. J. Soil Res.* 45, 543–553.

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409 **Figure Captions** (*created with SigmaPlot 12.0 and Canvas 11*) **ONLINE COLOR ONLY**

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411 **Fig 1** Cumulative emissions with standard error (error bars) over the incubation of both fertilized and unfertilized
412 treatments. Graphs “a” and “b” refers to soil CO₂ emissions, graphs “c” and “d” to soil NO_x emissions and graphs “e”
413 and “f” to soil NH₃ emissions. Different lowercase letters next to the legend indicate significant differences between the
414 treatments while different capital letters indicate significant differences between fertilized and unfertilized conditions of
415 each gas species, as results of statistical analysis (ANOVA and multi-comparison Fisher LSD test at p=0.05)

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418 **Table 1:** Experimental set-up. CNTR refer to the unamended soil, 5NZ mixture is created by adding 5% in weight of
 419 natural zeolite (NZ) to the untreated soil, 15NZ mixture is created by adding 15% in weight of NZ to the untreated soil
 420 while 10CZ mixture is created adding 10% in weight of NH₄⁺-enriched zeolites (CZ) to the untreated soil.
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	Treatments	Zeolite amount (wt%)	Zeolite type	N input from zeolites (kg N ha⁻¹)	N input from urea (kg N ha⁻¹)	Total N input to soil (kg N ha⁻¹)
	CNTR	0	/	/	170	170
Fertilized with Urea	5NZ	5	NZ	0	170	170
	15NZ	15	NZ	0	170	170
	10CZ	10	CZ	497	170	667
Unfertilized (no Urea added)	CNTR	0	/	/	0	0
	5NZ	5	NZ	0	0	0
	15NZ	15	NZ	0	0	0
	10CZ	10	CZ	497	0	497

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441 **Table 2:** Soil pH, exchangeable $\text{NH}_4^+\text{-N}$, $\text{NO}_3^-\text{-N}$ mean values (measured at the end of the incubation) and $\text{N}_2\text{O-N}$
 442 fluxes (measured after 4 h of incubation) with the associated standard errors between brackets. Different upper case
 443 letters indicate significant differences ($p < 0.05$) as results of statistical analysis (ANOVA and multi-comparison Fisher
 444 LSD test at $p = 0.05$). The presence of “n.m” indicate not measured values.

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	Treatments	pH	$\text{NH}_4^+\text{-N}$ (mg kg^{-1})	$\text{NO}_3^-\text{-N}$ (mg kg^{-1})	$\text{N}_2\text{O-N}$ ($\mu\text{g m}^{-2} \text{h}^{-1}$)
Unfertilized	CNTR	7.73 (0.04) ^a	10.1 (0.4) ^a	54.3 (0.7) ^b	n.m
	5NZ	7.80 (0.04) ^a	9.7 (0.3) ^a	47.9 (0.4) ^a	n.m
	15NZ	7.93 (0.06) ^a	9.5 (0.2) ^a	53.7 (0.9) ^b	n.m
	10CZ	7.79 (0.04) ^a	79.6 (6.9) ^b	309 (6.4) ^c	n.m
Fertilized with Urea (170 kg-N ha ⁻¹)	CNTR	7.94 (0.02) ^a	9.7 (0.3) ^a	97.4 (0.9) ^b	358 (69) ^b
	5NZ	7.99 (0.06) ^a	10.1 (0.2) ^a	84.7 (0.5) ^a	165 (37) ^a
	15NZ	8.12 (0.09) ^a	9.9 (0.4) ^a	83.6 (1.0) ^a	281 (21) ^{ab}
	10CZ	8.11 (0.10) ^a	81.6 (8.3) ^b	347 (12) ^c	155 (11) ^a

446 **Table 3:** Cumulative NO_x, NH₃ and CO₂ emissions over 24 h, % of reduction or increment with respect to the CNTR soil and ratio between cumulative emissions and N added.
 447 For treatments with no N added see ESM 2. Different upper case letters indicate significant differences (p<0.05) as results of statistical analysis (ANOVA and multi-comparison
 448 Fisher LSD test at p=0.05).

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	N input from zeolites (kg N ha ⁻¹)	N input from Urea (kg N ha ⁻¹)	24 h cumul. CO₂-C (kg C ha ⁻¹)	Reduction %	CO₂-C/N added	24 h cumul. NO_x-N (kg N ha ⁻¹)	Reduction %	NO_x-N/N added	24 h cumul. NH₃-N (kg N ha ⁻¹)	Reduction %	NH₃-N/N added
CNTR	0	170	425 (60) ^b	-	2.5	1.11 (0.06) ^b	-	0.007	0.42 (0.05) ^c	-	0.003
5NZ	0	170	334 (10) ^a	-21	2.0	0.77 (0.06) ^a	-31	0.005	0.32 (0.02) ^b	-24	0.002
15NZ	0	170	323 (51) ^a	-24	1.9	0.83 (0.17) ^a	-25	0.006	0.19 (0.01) ^a	-55	0.001
10CZ	497	0	457 (85) ^b	-	0.9	1.29 (0.15) ^b	-	0.003	0.18 (0.04) ^a	-57	0.0004
10CZ	497	170	298 (34) ^a	-30	0.4	1.01 (0.16) ^{ab}	-	0.002	0.52 (0.06) ^b	+24	0.001

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