Table 1. Essential agreement (%) between M.I.C.E. or Etest[™] strips and CLSI BMD (±1 log₂ dilution)

	Enterobacteriaceae ^a (n=40)		Non-fermenters ^b (n=40)		Staphylococci ^c (n=40)		Enterococci ^d (n=40)	
	M.I.C.E.	Etest	M.I.C.E.	Etest	M.I.C.E.	Etest	M.I.C.E.	Etest
Ampicillin	100.0	100.0					95.0	100.0
Amoxicillin/clavulanic acid	90.0	97.5						
Ciprofloxacin	100.0	97.5	97.5	97.5	85.0	95.0		
Cefotaxime	85.0	82.5						
Erythromycin					100.0	100.0		
Gentamicin	100.0	97.5	95.0	92.5	95.0	97.5		
Gentamicin HL							100.0	100.0
Imipenem	97.5	100.0	100.0	100.0				
Linezolid					97.5	100.0	100.0	100.0
Oxacillin					90.0	95.0		
Vancomycin					72.5	90.0	80.0	87.5
Overall	95.4	95.8	97.5	96.7	90.0	96.3	93.8	96.9

^aK. pneumoniae (n=20) and E. coli (n=20).

^bP. aeruginosa (n=20) and Acinetobacter spp. (n=20).

^cS. aureus (n=20) and coagulase-negative Staphylococcus (n=20).

^dE. faecalis (n=20) and E. faecium (n=20).

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Transparency declarations

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References

1 Mushtaq S, Warner M, Cloke J *et al.* Performance of the Oxoid M.I.C.EvaluatorTM Strips compared with the Etest[®] assay and BSAC agar dilution. J Antimicrob Chemother 2010; **65**: 1702–11.

2 Clinical and Laboratory Standards Institute. *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically— Eighth Edition: Approved Standard M07-A8.* CLSI, Wayne, PA, USA, 2009.

3 Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing: Twentieth Informational Supplement M100-S20. CLSI, Wayne, PA, USA, 2010.

4 National Committee for Clinical Laboratory Standards. *Development of In Vitro Susceptibility Testing Criteria and Quality Control Parameters— Second Edition: Approved Guideline M23-A2.* NCCLS, Wayne, PA, USA, 2001. **5** Mason EO, Lamberth LB, Hammeman WA *et al.* Vancomycin MICs for *Staphylococcus aureus* vary by detection method and have increased in a pediatric population since 2005. *J Clin Microbiol* 2009; **47**: 1628–30.

6 Sader HS, Rhomberg PR, Jones RN. Nine-hospital study comparing broth microdilution and Etest method results for vancomycin and daptomycin against methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2009; **53**: 3162–5.

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Comment on: Rifaximin modulates the colonic microbiota of patients with Crohn's disease: an *in vitro* approach using a continuous culture colonic model system

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Keywords: immunity, gastroenterology, CD4+/CD103+, gut homing, uncomplicated diverticular disease

Sir,

We read with interest the paper by Maccaferri et al.¹ We appreciated their reference to our previous work.² However, their reference misrepresents our data. In fact, in our paper we did not show that rifaximin reduced release of inflammatory cvtokines. Rather, we evaluated T cell subpopulations in uncomplicated diverticular disease (UDD) before and after a short course of rifaximin treatment. In the peripheral blood we demonstrated that CD103+ (gut homing T cells) were significantly increased at baseline and decreased after antibiotic treatment, suggesting a role for central immunity in the pathogenesis of the disease. More important and novel were our findings at the tissue level where CD4+/CD62L+ cells were increased before treatment, while CD4+/CD103+ cells increased after treatment. Our data strongly suggest that antibiotic treatment modified T cell subpopulation distributions both in the peripheral blood and at the tissue level and that the study of tissue infiltrating lymphocytes (TILs)^{3,4} is an interesting new tool in the study of gut homing, especially when combined with the characterization of T cell subpopulations in the peripheral blood. We have previously suggested that the study of TILs, as compared with peripheral blood, better reflects in more detail the pathological process at the site of infection.² Accordingly, in our previous study of immunological modifications induced by rifaximin, we demonstrated that in UDD antibiotic treatment reduced the recruitment of CD4+/ CD103+ cells in the peripheral blood and induced a significant increase in CD4+/CD103+ subpopulations at the tissue level, resulting in a redistribution of CD4+/CD103+. Although it is possible that these events may lead to altered cytokine production as suggested by Maccaferri et al.,¹ unfortunately this was not part of our study. Heterogeneity of CD4 cells has been shown in the past,⁵ but only recently novel T cell subpopulations, such as T regulatory cells (Tregs), Th17 cells and CD4+/ CD103+ gut homing cells, have been described in detail (as reviewed by Pandolfi et $al.^6$).

We conclude that the study of CD4 subpopulations in the gastrointestinal mucosa is a valuable new tool to understand the effects of treatments for diseases causing inflammation. Our results were not acknowledged by Maccaferri's citation of our paper, while they incorrectly reported that our work showed a modification in cytokine production.

Transparency declarations

None to declare.

References

1 Maccaferri S, Vitali B, Klinder A *et al.* Rifaximin modulates the colonic microbiota of patients with Crohn's disease: an *in vitro* approach using a continuous culture colonic model system. J Antimicrob Chemother 2010; **65**: 2556–65.

2 Cianci R, Iacopini F, Petruzziello L *et al.* Involvement of central immunity in uncomplicated diverticular disease. *Scand J Gastroenterol* 2009; **44**: 108–15.

3 Kurnick JT, Ramirez-Montagut T, Boyle LA *et al.* A novel autocrine pathway of tumor escape from immune recognition: melanoma cell lines produce a soluble protein that diminishes expression of the gene encoding the melanocyte lineage melan-A/MART-1 antigen through down-modulation of its promoter. *J Immunol* 2001; **167**: 1204–11.

4 Bennett WT, Pandolfi F, Grove BH *et al.* Dominant rearrangements among human tumor-infiltrating lymphocytes. Analysis of T-cells derived from 32 patients with melanoma, lung, and renal cell carcinoma. *Cancer* 1992; **69**: 2379–84.

5 Pandolfi F, Corte G, Quinti I *et al.* Defect of T helper lymphocytes, as identified by the 5/9 monoclonal antibody, in patients with common variable hypogammaglobulinaemia. *Clin Exp Immunol* 1983; **51**: 470–4.

6 Pandolfi F, Cianci R, Pagliari D *et al.* Cellular mediators of inflammation: tregs and TH17 cells in gastrointestinal diseases. *Mediators Inflamm* 2009; **2009**: 132028 (doi:10.1155/2009/132028).

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Rifaximin modulates the colonic microbiota of patients with Crohn's disease: an *in vitro* approach using a continuous culture colonic model system—authors' response

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Keywords: inflammatory bowel disease, immune modulation, immune response

Sir,

We read with interest the comment by Cianci *et al.*,¹ referring to our recent publication in JAC.²

Cianci *et al.*¹ state that we cited their recent work³ incorrectly. We appreciate the importance of the research of Cianci *et al.*³ on gut-homing T cells for the understanding of inflammatory conditions in the bowel. Further, we regret that in our sentence concerning 'alternative mechanisms of action' of rifaximin, including '(iii) reduction of inflammatory cytokine release', we did not expand on the complexity of the involvement of the immune system. We are grateful for the detailed explanation and clarification by Cianci *et al.*¹ regarding their article³ that we referred to.

We would like to emphasize that it was our intention to discuss how the therapeutic efficacy of the antibiotic rifaximin can be