

Microplastic ingestion affects the lateralised processing of predator stimuli in fish

Georgiana Andrei¹  | Alessandro Colombani¹ | Elia Gatto^{1,2} | Marco Scoconi³ | Luigi Abelli¹ | Annalaura Mancia^{1,4} | Cristiano Bertolucci¹ | Tyrone Lucon-Xiccato¹

¹Department of Life Sciences and Biotechnology, University of Ferrara, Ferrara, Italy

²Department of Chemical, Pharmaceutical and Agricultural Sciences, University of Ferrara, Ferrara, Italy

³Istituto sulla Sintesi Organica e Fotoreattività del CNR, Research unit at the Department of Life Sciences and Biotechnology, University of Ferrara, Ferrara, Italy

⁴Department of Biology, University of North Florida, Florida, USA

Correspondence

Georgiana Andrei, Department of Life Sciences and Biotechnology, University of Ferrara, Ferrara, via Luigi Borsari, 46, 44121, Italy. Email: georgiana.andrei@unife.it

Funding information

Regione Emilia Romagna - PR FSE+ 2021/2027, Grant/Award Number: PA 2023-19070/RER

Abstract

Microplastic ingestion affects fish brains at the molecular level, but its impact on cognitive phenotype remains unclear. We fed zebrafish (*Danio rerio*) food containing either polyethylene or poly(butylene-adipate-co-terephthalate) microplastics for 20 days and assessed their lateralisation, which reflects how information processing is split between brain hemispheres. No changes appeared in rotational or mirror tests, but lateralisation was disrupted in the detour test when facing a predator model. These results suggest microplastic ingestion can impair specific cognitive traits at the phenotypic level.

KEYWORDS

cognitive ecology, cognitive plasticity, environmental pollution, laterality

Microplastics, plastic fragments smaller than 5 mm, have become one of the most pervasive pollutants in natural habitats (Rillig et al., 2017; Scheurer & Bigalke, 2018), including aquatic ecosystems (Li et al., 2020; Lindeque et al., 2020). When ingested by fish, they induce several toxic effects, such as inflammation (Kim et al., 2021; Lu et al., 2016; Qiao et al., 2019) and alterations in physiology (Sharifinia et al., 2020; Wang et al., 2019), metabolism (Boopathi et al., 2023; Cedervall et al., 2012) and other related biological processes (Huang et al., 2022). Although the typical size of microplastics may prevent direct penetration into the brain, the resulting physiological disruptions are expected to indirectly affect brain function (Maille & Schradin, 2017). Moreover, it has recently been shown that microplastics may affect blood flow to the brain (Huang et al., 2025). Accordingly, various studies have reported changes in gene expression and neurotransmitter levels in the brains of fish exposed to microplastics (Barboza et al., 2018, 2023; Ding et al., 2023; Hoyo-Alvarez

et al., 2022; Huang et al., 2023). However, studies investigating the cognitive phenotype found no significant effects on learning, cognitive flexibility and inhibitory control (Irwin et al., 2024; Lucon-Xiccato et al., 2025). In this study, we investigated whether microplastics could lead to phenotypic effects on a fundamental cognitive function: lateralisation, defined as the division of information processing between the two cerebral hemispheres (Bisazza et al., 1998).

Following the treatment procedure by Lucon-Xiccato et al. (2025), two groups of zebrafish (*Danio rerio*) were fed commercial dry food mixed with polyethylene (PE; $N = 35$) or poly(butylene-adipate-co-terephthalate) (PBAT; $N = 35$) microplastics (0.01 g of food and 100 μg of microplastics per fish/day) for 20 days. PE was selected as a common conventional plastic and PBAT as a common biodegradable plastic. A control group ($N = 35$) received only dry food. Fish were kept in groups of five in $40 \times 30 \times 22$ cm tanks under standard conditions ($28 \pm 1^\circ\text{C}$, 14 h:10 h light–dark cycle).

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Journal of Fish Biology* published by John Wiley & Sons Ltd on behalf of Fisheries Society of the British Isles.

After the treatment, subjects' lateralisation was assessed using a battery of three standard tests based on asymmetric behaviours: the rotational test, the mirror test and the detour test. In the rotational test (De Russi et al., 2025; Rovegno et al., 2025), each subject was individually tested in a circular white tank to observe its spontaneous swimming behaviour. Immediately after this, the subject was transferred to an octagonal tank with mirrored walls to assess visual lateralisation in response to their own reflection, which simulates a conspecific (Lucon-Xiccato et al., 2020; Rovegno et al., 2025). Both the rotational and mirror tests lasted 20 min. The trials were recorded using a Sony Handycam HDR-CX405 (resolution camera 1920×1080 , 25 frames per second) located approximately 50 cm above the tanks. Using the recordings and a custom stopwatch software (Ciclic Timer, written in Delphi 5 Borland), we calculated the time each subject spent swimming both clockwise and counterclockwise close to the tank edge (within two body lengths, 3 cm) in the rotational test as a measure of lateralisation in motor functions. In the mirror test, we similarly measured the time spent swimming clockwise and counterclockwise near the mirror (within one body length). Clockwise swimming in the mirror test is assumed to reflect left-eye use and, due to the decussation of the optic nerves, right-hemisphere processing of the social stimulus, and vice versa (Sovrano et al., 1999; De Santi et al., 2001). Finally, subjects were tested in a detour test to evaluate lateralisation in eye preference when inspecting a predator model. We used the predator model because a recent study suggested that, under certain conditions, the detour test using non-biological stimuli may not be reliable in fish (Roche et al., 2020). However, research conducted on our study species using our specific stimulus has not raised such concerns and has demonstrated substantial within- and between-study reliability (Bisazza et al., 2007; Facchin et al., 2009). Following Rovegno et al. (2025), each subject was gently guided through a corridor toward a plastic predator dummy placed behind a barrier made of vertical sticks. The dummy-predator consisted of a fish lure (18 cm) resembling a natural predator of zebrafish, the Indian leaf fish, *Nandus nandus* (Parichy & Postlethwait, 2020). The direction of the turn made in front of the barrier (left or right) was used to infer hemisphere use and thus lateralisation in response to the predator stimulus (Sovrano et al., 1999; De Santi et al., 2001): a right turn corresponded to left-eye use (right hemisphere processing) and a left turn indicated right-eye use (left hemisphere processing). We collected 10 consecutive detour trials for each subject.

For all the tests, we computed two lateralisation indices. The relative lateralisation index indicated the average directionality of swimming or turning preference of each subject. It ranged from -1 to $+1$, where -1 indicates a counterclockwise swimming preference (in the rotational and mirror tests) or a left turning preference (in the detour test), and $+1$ indicates a clockwise swimming preference (rotational and mirror tests) or a right turning preference (detour test). For the rotational and the mirror test, the relative lateralisation index was computed as: (time spent swimming in the right direction – time spent swimming in the left direction)/(time spent swimming in the right direction + time spent swimming in the left direction). For the detour test, the relative lateralisation index was computed as:

(number of turns right – numbers of turns left)/total number of turns. The second index is the absolute lateralisation index, which is the absolute value of the relative lateralisation index and describes the strength of lateralisation, regardless of its direction (De Russi et al., 2025).

A one-way analysis of variance (ANOVA) with treatment as fixed effect revealed no significant effect of microplastics on the relative ($F_{2,99} = 1.160$, $p = 0.566$) and the absolute ($F_{2,99} = 1.387$, $p = 0.255$) lateralisation index of the rotational test (Figure 1d). Also in the mirror test, the ANOVA did not show a significant effect of the treatment (relative index: $F_{2,88} = 0.015$, $p = 0.985$; absolute index: $F_{2,88} = 0.755$, $p = 0.473$; Figure 1e).

In the detour test, a generalized linear-mixed effects model on the outcome of each trial (0 = left turn, 1 = right turn) did not show a significant effect of the trial number ($\chi^2_1 = 0.929$, interaction trial \times treatment $\chi^2_2 = 0.914$), suggesting that fish did not alter their behaviour across the 10 trials. An ANOVA revealed a significant effect of treatment (Figure 1f) on the relative lateralisation index ($F_{2,99} = 3.321$, $p = 0.040$), but not on the absolute lateralisation index ($F_{2,99} = 0.545$, $p = 0.582$). The post hoc tests showed a significant difference in the relative lateralisation index between the control and the PE group ($p = 0.048$), but there was no significant difference between the control and the PBAT group ($p = 0.122$) and between the PE and PBAT groups ($p = 0.912$). A one-sample *t*-test revealed that the relative index of the control group was significantly greater than zero, indicating a preference to observe the dummy predator with the left eye, thus processing it with the right hemisphere ($t_{34} = 2.111$, $p = 0.042$). Conversely, the subjects exposed to microplastics did not show a viewing preference for the predator stimulus and thus no lateralisation (PE: $t_{32} = -1.139$, $p = 0.263$; PBAT: $t_{32} = -0.641$, $p = 0.526$).

Our findings suggest that microplastic ingestion does not affect the lateralisation of cognitive functions involved in the rotational and mirror tests. These functions likely play a role in determining spontaneous locomotion direction and the visual processing of conspecifics. However, we cannot exclude the possibility that the absence of an effect was due to these functions not being lateralised in the studied population. Indeed, we did not observe any significant population bias in the control group. In contrast, the processing of a predator stimulus in the detour test was lateralised in the control population. For this function, our analysis revealed a significant disruption due to the microplastic treatment. Specifically, fish in both the PE- and PBAT-treated groups were not lateralised, unlike those in the control group.

Considering also previous studies, we conclude that although microplastic ingestion appears not to affect learning, cognitive flexibility or inhibitory control (Irwin et al., 2024; Lucon-Xiccato et al., 2025), it does influence certain aspects of cerebral lateralisation. This may be because lateralisation represents a more fundamental aspect of brain function and may therefore be more susceptible to molecular alterations in the brain. Notably, the effects of microplastics were similar regardless of their origin. This raises concerns about biodegradable plastics (e.g. PBAT), which may generate low-molecular-weight polymer chains more rapidly than conventional plastics, potentially leading to greater impacts on aquatic organisms.

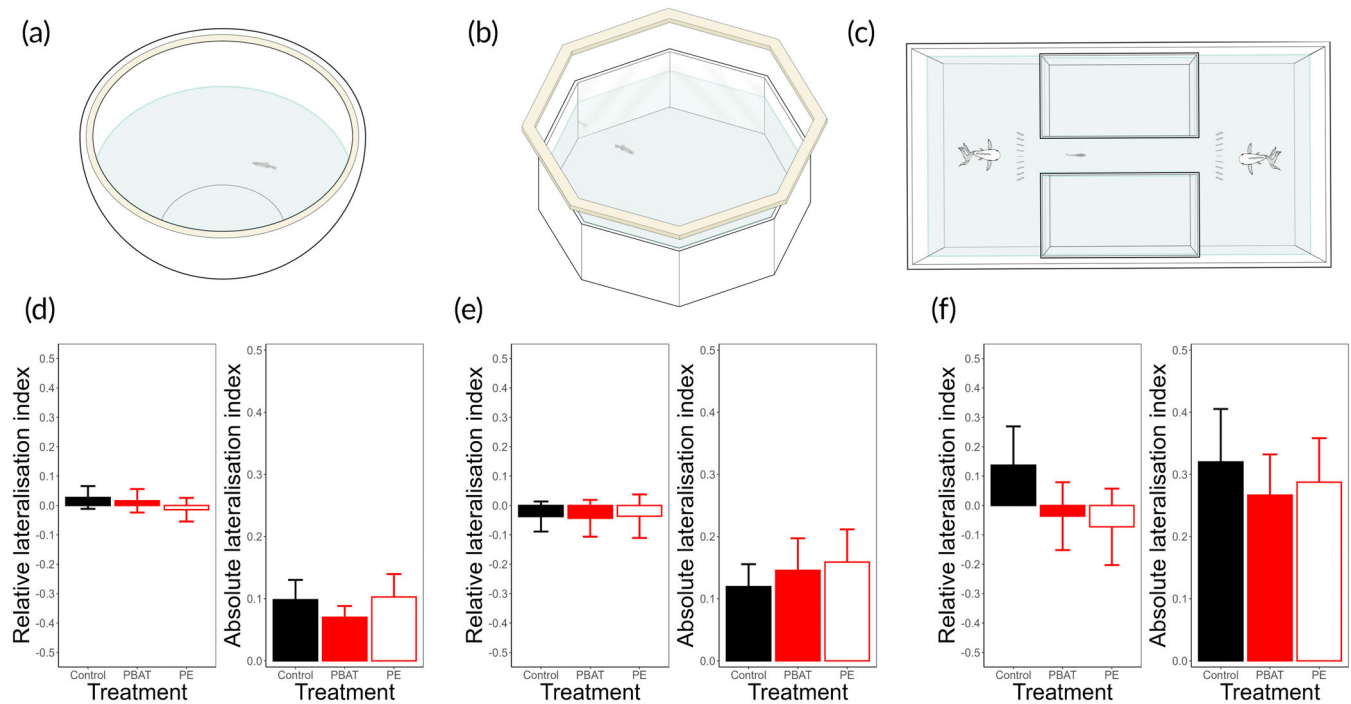


FIGURE 1 Apparatuses used in the (a) rotational test, (b) mirror test and (c) detour test. Means and standard errors of the relative lateralisation index and the absolute lateralisation index of the subjects assessed in the (d) rotational test, (e) mirror test and (f) detour test. PBAT, poly(butylene-adipate-co-terephthalate); PE, polyethylene.

What does altered lateralisation mean for fish exposed to microplastics in natural environments? It is generally believed that lateralisation in a population results from both genetic and plasticity adaptations (reviewed in Bisazza & Lucon-Xiccato, 2025). In particular, fish from high-predator environments tend to exhibit stronger lateralisation, which enhances predator avoidance and survival (Brown et al., 2004; Ferrari et al., 2015). Disruptions in lateralisation due to microplastic exposure could lead to a phenotype-environment mismatch, potentially reducing fitness. Additionally, microplastics may not be the only anthropogenic factor affecting lateralisation in fish populations. Water acidification (Domenici et al., 2012), hypoxia (Lucon-Xiccato et al., 2014), rising temperatures (Domenici et al., 2014), artificial illumination (De Russi et al., 2025) and various chemical pollutants (Merola et al., 2024; Suriyampola et al., 2024) have all been observed to alter lateralisation. Consequently, the effects due to microplastics could be additive to those caused by other environmental stressors, potentially leading to even more pronounced fitness consequences.

AUTHOR CONTRIBUTIONS

G.A.: Investigation, formal analysis, data curation, writing – original draft, visualization. A.C.: Investigation. E.G.: Methodology, writing – review and editing; M.S.: Resources, writing – review & editing. L.A.: Writing – review and editing. A.M.: Writing – review and editing. C.B.: Conceptualization, writing – review and editing, funding acquisition. T.L.-X.: Conceptualization, methodology, writing – original draft, supervision, funding acquisition.

ACKNOWLEDGEMENTS

We have no competing interests. We are thankful to Anna Chiofalo. The G.A. Doctoral scholarships were funded by Regione Emilia Romagna – PR FSE+ 2021/2027 (PA 2023-19070/RER). The project was approved by the Ethical Committee of the University of Ferrara Organismo Preposto al Benessere degli Animali and by the Italian Ministry of University and Research (protocol n. 446/2021-PR). Open access publishing facilitated by Università degli Studi di Ferrara, as part of the Wiley - CRUI-CARE agreement.

ORCID

Georgiana Andrei  <https://orcid.org/0009-0000-4316-701X>

REFERENCES

- Barboza, L. G. A., Otero, X. L., Fernández, E. V., Vieira, L. R., Fernandes, J. O., Cunha, S. C., & Guilhermino, L. (2023). Are microplastics contributing to pollution-induced neurotoxicity? A pilot study with wild fish in a real scenario. *Heliyon*, 9(1), e13070.
- Barboza, L. G. A., Vieira, L. R., Branco, V., Figueiredo, N., Carvalho, F., Carvalho, C., & Guilhermino, L. (2018). Microplastics cause neurotoxicity, oxidative damage and energy-related changes and interact with the bioaccumulation of mercury in the European seabass, *Dicentrarchus labrax* (Linnaeus, 1758). *Aquatic Toxicology*, 195, 49–57.
- Bisazza, A., & Lucon-Xiccato, T. (2025). Individual differences in vertebrate behavioural lateralisation: The role of genes and environment. *Symmetry*, 17, 527.
- Bisazza, A., Dadda, M., Facchin, L., & Vigo, F. (2007). Artificial selection on laterality in the teleost fish *Girardinus falcatus*. *Behavioural Brain Research*, 178, 29–38.

- Bisazza, A., Rogers, L. J., & Vallortigara, G. (1998). The origins of cerebral asymmetry: A review of evidence of behavioural and brain lateralization in fishes, reptiles and amphibians. *Neuroscience & Biobehavioral Reviews*, 22, 411–426.
- Boopathi, S., Haridevamuthu, B., Mendonca, E., Gandhi, A., Priya, P. S., Alkahtani, S., & Malafaia, G. (2023). Combined effects of a high-fat diet and polyethylene microplastic exposure induce impaired lipid metabolism and locomotor behavior in larvae and adult zebrafish. *Science of the Total Environment*, 902, 165988.
- Brown, C., Gardner, C., & Braithwaite, V. A. (2004). Population variation in lateralized eye use in the poeciliid *Brachyraphis episcopi*. *Proceedings. Biological sciences*, 271(suppl_6), S455–S457.
- Cedervall, T., Hansson, L. A., Lard, M., Frohm, B., & Linse, S. (2012). Food chain transport of nanoparticles affects behaviour and fat metabolism in fish. *PLoS One*, 7, e32254.
- de Russi, G., Bertolucci, C., & Lucon-Xiccato, T. (2025). Artificial light at night impairs visual lateralisation in a fish. *Journal of Experimental Biology*, 228, JEB249272.
- De Santi, A., Sovrano, V. A., Bisazza, A., & Vallortigara, G. (2001). Mosquitofish display differential left-and right-eye use during mirror image scrutiny and predator inspection responses. *Animal Behaviour*, 61(2), 305–310.
- Ding, P., Xiang, C., Li, X., Chen, H., Shi, X., Li, X., & Hu, G. (2023). Photoaged microplastics induce neurotoxicity via oxidative stress and abnormal neurotransmission in zebrafish larvae (*Danio rerio*). *Science of the Total Environment*, 881, 163480.
- Domenici, P., Allan, B. J., Watson, S. A., McCormick, M. I., & Munday, P. L. (2014). Shifting from right to left: The combined effect of elevated CO₂ and temperature on behavioural lateralization in a coral reef fish. *PLoS One*, 9, e87969.
- Domenici, P., Allan, B., McCormick, M. I., & Munday, P. L. (2012). Elevated carbon dioxide affects behavioural lateralization in a coral reef fish. *Biology Letters*, 8, 78–81.
- Facchin, L., Argenton, F., & Bisazza, A. (2009). Lines of *Danio rerio* selected for opposite behavioural lateralization show differences in anatomical left–right asymmetries. *Behavioural Brain Research*, 197, 157–165.
- Ferrari, M. C., McCormick, M. I., Allan, B. J., Choi, R. B., Ramasamy, R. A., & Chivers, D. P. (2015). The effects of background risk on behavioural lateralization in a coral reef fish. *Functional Ecology*, 29, 1553–1559.
- Hoyo-Alvarez, E., Arechavala-Lopez, P., Jiménez-García, M., Solomando, A., Alomar, C., Sureda, A., & Deudero, S. (2022). Effects of pollutants and microplastics ingestion on oxidative stress and monoaminergic activity of seabream brains. *Aquatic Toxicology*, 242, 106048.
- Huang, H., Hou, J., Li, M., Wei, F., Liao, Y., & Xi, B. (2025). Microplastics in the bloodstream can induce cerebral thrombosis by causing cell obstruction and lead to neurobehavioral abnormalities. *Science Advances*, 11, eadr8243.
- Huang, J. N., Wen, B., Xu, L., Ma, H. C., Li, X. X., Gao, J. Z., & Chen, Z. Z. (2022). Micro/nano-plastics cause neurobehavioral toxicity in discus fish (*Symphysodon aequifasciatus*): Insight from brain-gut-microbiota axis. *Journal of Hazardous Materials*, 421, 126830.
- Irwin, K., Hathorn, G., & Gabor, C. R. (2024). Cognitive and behavioral response of mosquitofish (*Gambusia affinis*) to environmental factors: Microplastics, predator cues, and detour design methods. *Journal of Fish Biology*, 106, 1–10. <https://doi.org/10.1111/jfb.15998>
- Kim, J. H., Yu, Y. B., & Choi, J. H. (2021). Toxic effects on bioaccumulation, hematological parameters, oxidative stress, immune responses and neurotoxicity in fish exposed to microplastics: A review. *Journal of Hazardous Materials*, 413, 125423.
- Li, C., Busquets, R., & Campos, L. C. (2020). Assessment of microplastics in freshwater systems: A review. *Science of the Total Environment*, 707, 135578.
- Lindeque, P. K., Cole, M., Coppock, R. L., Lewis, C. N., Miller, R. Z., Watts, A. J., Watts, A. J. R., Wilson-McNeal, A., Wright, S. L., & Galloway, T. S. (2020). Are we underestimating microplastic abundance in the marine environment? A comparison of microplastic capture with nets of different mesh-size. *Environmental Pollution*, 265, 114721.
- Lu, Y., Zhang, Y., Deng, Y., Jiang, W., Zhao, Y., Geng, J., & Ren, H. (2016). Uptake and accumulation of polystyrene microplastics in zebrafish (*Danio rerio*) and toxic effects in liver. *Environmental Science & Technology*, 50, 4054–4060.
- Lucon-Xiccato, T., Montalbano, G., Andrei, G., Frigato, E., Scoponi, M., Abelli, L., Mancía, A., & Bertolucci, C. (2025). Does microplastic ingestion affect fish cognition? Under review.
- Lucon-Xiccato, T., Montalbano, G., Dadda, M., & Bertolucci, C. (2020). Lateralization correlates with individual differences in inhibitory control in zebrafish. *Biology Letters*, 16, 20200296.
- Lucon-Xiccato, T., Nati, J. J., Blasco, F. R., Johansen, J. L., Steffensen, J. F., & Domenici, P. (2014). Severe hypoxia impairs lateralization in a marine teleost fish. *Journal of Experimental Biology*, 217, 4115–4118.
- Maille, A., & Schradin, C. (2017). Ecophysiology of cognition: How do environmentally induced changes in physiology affect cognitive performance? *Biological Reviews*, 92, 1101–1112.
- Merola, C., Caioni, G., Bertolucci, C., Lucon-Xiccato, T., Savaşçı, B. B., Tait, S., & Perugini, M. (2024). Embryonic and larval exposure to propylparaben induces developmental and long-term neurotoxicity in zebrafish model. *Science of the Total Environment*, 912, 168925.
- Parichy, D. M., & Postlethwait, J. H. (2020). The biotic and abiotic environment of zebrafish. In D. M. Parichy & J. H. Postlethwait (Eds.), *Behavioral and neural genetics of zebrafish* (pp. 3–16). Academic Press.
- Qiao, R., Deng, Y., Zhang, S., Wolosker, M. B., Zhu, Q., Ren, H., & Zhang, Y. (2019). Accumulation of different shapes of microplastics initiates intestinal injury and gut microbiota dysbiosis in the gut of zebrafish. *Chemosphere*, 236, 124334.
- Rillig, M. C., Ingrassia, R., & de Souza Machado, A. A. (2017). Microplastic incorporation into soil in agroecosystems. *Frontiers in Plant Science*, 8, 1805.
- Roche, D. G., Amcoff, M., Morgan, R., Sundin, J., Andreassen, A. H., Finnøen, M. H., & Binning, S. A. (2020). Behavioural lateralization in a detour test is not repeatable in fishes. *Animal Behaviour*, 167, 55–64.
- Rovigno, E., Frigato, E., Dalla Valle, L., Bertolucci, C., & Lucon-Xiccato, T. (2025). Expression of glucocorticoid-receptor covaries with individual differences in visual lateralisation in zebrafish. *Animal Cognition*, 28, 21.
- Scheurer, M., & Bigalke, M. (2018). Microplastics in Swiss floodplain soils. *Environmental Science & Technology*, 52, 3591–3598.
- Sharifinia, M., Bahmanbeigloo, Z. A., Keshavarzifard, M., Khanjani, M. H., & Lyons, B. P. (2020). Microplastic pollution as a grand challenge in marine research: A closer look at their adverse impacts on the immune and reproductive systems. *Ecotoxicology and Environmental Safety*, 204, 111109.
- Sovrano, V. A., Rainoldi, C., Bisazza, A., & Vallortigara, G. (1999). Roots of brain specializations: preferential left-eye use during mirror-image inspection in six species of teleost fish. *Behavioural Brain Research*, 106(1-2), 175–180.
- Suriyampola, P. S., Huang, A. J., Lopez, M., Conroy-Ben, O., & Martins, E. P. (2024). Exposure to environmentally relevant concentrations of Bisphenol-a linked to loss of visual lateralization in adult zebrafish (*Danio rerio*). *Aquatic Toxicology*, 268, 106862.
- Wang, J., Li, Y., Lu, L., Zheng, M., Zhang, X., Tian, H., & Ru, S. (2019). Polystyrene microplastics cause tissue damages, sex-specific reproductive disruption and transgenerational effects in marine medaka (*Oryzias melastigma*). *Environmental Pollution*, 254, 113024.

How to cite this article: Andrei, G., Colombani, A., Gatto, E., Scoponi, M., Abelli, L., Mancía, A., Bertolucci, C., & Lucon-Xiccato, T. (2025). Microplastic ingestion affects the lateralised processing of predator stimuli in fish. *Journal of Fish Biology*, 1–4. <https://doi.org/10.1111/jfb.70169>