

Review

# Aged Garlic Extract and Its Bioactive Molecules S-Allyl-Cysteine and S1-Propenyl-Cysteine: A Review Focusing on Evidences Supporting Their Use for Mitigating the Effects of Cigarette Smoking

Roberto Gambari \*  and Alessia Finotti \* 

Department of Life Sciences and Biotechnology, Ferrara University, I-44121 Ferrara, Italy

\* Correspondence: gam@unife.it (R.G.); alessia.finotti@unife.it (A.F.)

## Abstract

One of the major social issues worldwide is tobacco dependency and cigarette smoking (CS) abuse. Given the significant impact of cigarette smoking on human health and diseases, extensive tobacco use and cigarette smoking abuse are certainly a form of drug addiction and should be considered a serious threat to human health. Notably, healthcare spending attributable to cigarette smoking is very high. In this regard, a significant number of biomolecules of natural origin have been described as capable of mitigating the adverse effects of cigarette smoking. In this review, (a) we discuss the impact that the habit of smoking tobacco has on human health and (b) we describe products of natural origin capable of mitigating the effects of cigarette smoke. The conclusion of this review article is that the available information strongly indicates a possible use of the anti-inflammatory aged garlic extract (AGE) and its bioactive components for mitigating the detrimental effects of cigarette smoke on human tissues. The key reasons for proposing this application are that AGE and its key components S-allyl-cysteine and S1-propenyl-cysteine are potent anti-inflammatory agents, bind to Toll-like Receptor-4, inhibit Nuclear Factor- $\kappa$ B, inhibit the expression of pro-inflammatory genes, revert apoptosis induced by cigarette smoke in several cellular model systems and are strong inhibitors of Reactive Oxygen Species (ROS) formation.

**Keywords:** natural products; cigarette smoke; inflammation; aged garlic extract



Academic Editor: Hyun-Ock Pae

Received: 23 July 2025

Revised: 21 August 2025

Accepted: 23 August 2025

Published: 26 August 2025

**Citation:** Gambari, R.; Finotti, A. Aged Garlic Extract and Its Bioactive Molecules S-Allyl-Cysteine and S1-Propenyl-Cysteine: A Review Focusing on Evidences Supporting Their Use for Mitigating the Effects of Cigarette Smoking. *Molecules* **2025**, *30*, 3496. <https://doi.org/10.3390/molecules30173496>

**Copyright:** © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

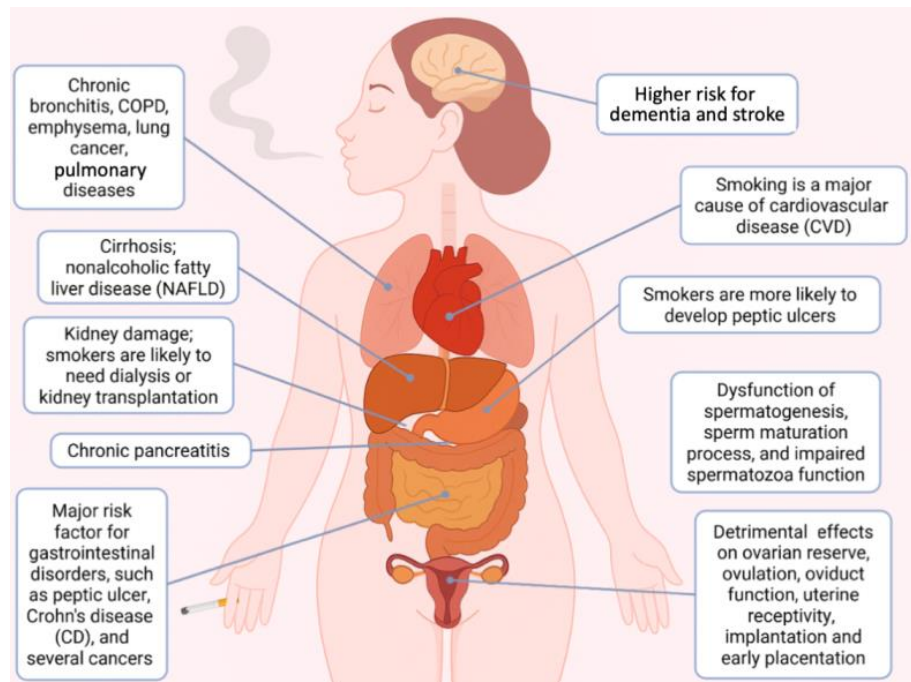
One of the major social issues worldwide is tobacco dependency and cigarette smoking (CS) abuse [1]. According to the publication *WHO global report on trends in prevalence of tobacco use 2000–2030* (16 January 2024, ISBN: 978-92-4-008828-3) [1], although the total number of tobacco users has declined steadily over the period 2000–2022, this number is still expected to be very high (around 1.20 billion) by 2030 [1,2]. Given the significant impact of cigarette smoking on human health and diseases [3–9], extensive tobacco use and cigarette smoking abuse are certainly a form of drug addiction and should be considered a serious threat to human health [1]. Notably, healthcare spending attributable to cigarette smoking is very high [10,11]. In order to limit tobacco use, several actions have been considered to help eliminate smoking [12–15], such as bans of tobacco advertising [16] and the introduction of taxes as a share of cigarette prices [17,18]. Despite these initiatives, the habit of smoking tobacco on a consistent basis is still a very significant social problem. In

this regard, a significant number of biomolecules of natural origin have been described as capable of mitigating the adverse effects of cigarette smoking "in vitro" on cells and tissues and "in vivo" on complex organisms [19–21]. In this review, (a) we discuss the impact that the habit of smoking tobacco has on health and costs for national health systems, (b) we describe products of natural origin capable of mitigating the adverse effects of cigarette smoking and (c) we focus on the possible use of aged garlic extract (AGE) and its bioactive components for mitigating the adverse effects of cigarette smoking.

## 2. Impact of Cigarette Smoke on Human Health

### 2.1. Smoking and Human Diseases

Smoking causes cancer [22], heart disease [23], stroke [24], lung diseases [9,25], diabetes [26], chronic obstructive pulmonary disease (COPD) [27] and pancreatic diseases [28], as shown in Figure 1. Smoking is a particularly large problem in high-income countries, where cigarette smoking is the most important cause of preventable disease and death [29]. The impact of smoking is devastating on the individual level, considering that the life expectancy of those who smoke regularly is about 10 years lower than that of non-smokers [29]. The decline of cigarette smoking might be achieved through successful global health campaigns, including bans on tobacco advertising, the introduction of taxes on cigarettes and the development of plans to help people quit smoking. All these issues are discussed by Roser M (<https://ourworldindata.org/smoking-big-problem-in-brief#>) [accessed on 22 May 2025] [29]. For example, by taxing cigarettes very heavily, many governments made cigarettes much more expensive. Of course, reducing the affordability of cigarettes is one of the most important—and cost-effective—ways to reduce smoking and increase public health [29].



**Figure 1.** Human pathologies associated with cigarette smoking abuse. Picture created using Bio-Render.com (accessed on 16 July 2025).

### 2.2. Smoking and Cancer

Smoking (and indirect smoking) causes or increases the risk for many types of cancer [30–32], including acute myeloid leukemia [33], bladder cancer [34], cervical cancer [35], colorectal cancer [36], esophageal cancer [37], prostate cancer [38], kidney cancer [39], laryn-

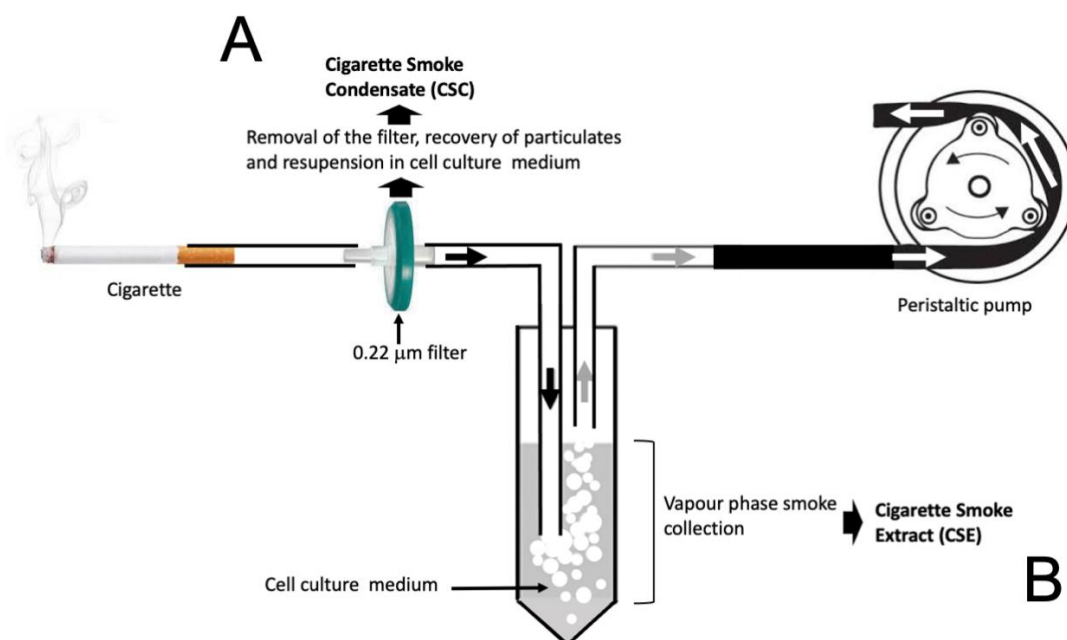
geal cancer and other throat cancers [40], liver cancer [41], lung cancer [42], oral cancer [43], pancreatic cancer [44] and stomach cancer [45].

In this respect, it should be underlined that tobacco smoking is associated in many cases with reduced efficacy or even failure of first-line cancer treatments; this causes incremental costs for the management of cancer patients [46]. In this respect, it is generally accepted that smoking seriously impacts health system costs, including those regarding cancer patients [47–51]. Accordingly, it is imperative that more stringent steps are taken to reduce the huge economic burden of human pathologies (including cancer) linked to smoking.

### 3. Mechanism(s) of Action of Cigarette Smoking: Inflammation

The cellular and molecular mechanisms responsible for the solid interplay between cigarette smoke (CS) and inflammation have been reviewed by Lee et al. [52]. In this respect, it should be underlined that the identification of cellular, biochemical and molecular effects of CS is a key step for the identification of molecular targets for medical interventions. As a first consideration, we should mention that the several toxins and trace amounts of microbial cell components present in CS induce chronic inflammation [53–55]. In the CS-dependent activation of pro-inflammatory genes, several proteins play a crucial role and should be considered as possible biochemical targets for therapeutic intervention, among which is the Nuclear Factor- $\kappa$ B (NF- $\kappa$ B) pathway [56,57], associated with the activation of Toll-like Receptor-4 (TLR4) [58–63].

Several experimental model systems are available to characterize the effects of cigarette smoke on cultured cell lines and the mitigation of these detrimental effects using natural products. Two are based on the production of “Cigarette Smoke Condensates” (CSCs) [64–66] and “Cigarette Smoke Extracts” (CSEs) [67–69]. Figure 2 reports a pictorial representation of the production of CSCs and CSEs starting from cigarette burning.



**Figure 2.** Scheme outlining the preparation of “Cigarette Smoke Concentrates” (CSCs) and “Cigarette Smoke Extracts” (CSEs), using information taken from Kim and Kim (2023) [70], Higashi et al. (2014) [71] and Wright (2015) [72]. In the representation here depicted, CSC is derived from the particulates trapped on the filter (A), whereas CSE is the resulting aqueous solution after bubbling the smoke through the medium (B).

The key step of CSC preparation is the trapping of the cigarette condensate in a 0.22  $\mu\text{m}$  filter pad; then the cigarette smoke particulates are eluted using solvents, such as methanol, dimethyl sulfoxide (DMSO) or ethanol, recovered and transferred to tissue culture medium (CSC) for testing the CSC effects on cultured cells. Description of CSC preparation methods can be found in Kim and Kim [70] and in Mathewson [73]. CSE is an aqueous solution that contains toxic compounds produced by cigarette smoke. Therefore, CSE is useful to determine the effects of cigarette smoke on in vitro cultured cell lines. CSE can be prepared by collecting the smoke from a cigarette as shown in Figure 2. The cigarette smoke is “bubbled” in cell culture medium under a negative pressure generated by a peristaltic pump. The aqueous components are therefore diluted in the cell culture medium which, at the end of the procedure, is referred as “Cigarette Smoke Extract” (CSE). The parameters to be considered are the following: (a) number of the cigarettes; (b) volume of the cell culture medium and (c) flow rate generated by the peristaltic pump. Description of CSE preparation methods can be found in Amel Al-Hashimi et al. [69], Higashi et al. [71], Wight [72] and Agraval et al. [74]. A detailed protocol is available (<https://dx.doi.org/10.17504/protocols.io.bnymmfu6>; accessed on 22 May 2025).

In addition to using CSC and CSE, the effect of cigarette smoke in vitro can be assessed by direct exposure of cells or cellular tissues to cigarette smoke based on the air–liquid interface exposure [75]. In this respect, Singh et al. presented a perspective view of the challenges and opportunities of “Lung-on-Chip” technologies in studies focusing on cigarette smoking related in vitro inhalation toxicology [76]. With respect to the chemical composition of CSC and CSE, several studies are available [77–81]. In this respect, Kim et al. compared the volatile organic compounds (VOCs) of cigarette smoke condensate (CSC) and extract (CSE) samples [82]. The CSC sample mainly contained nicotine, nicotyrine and a lower relative amount of 1,2,3-propanetriol, triacetate, ethyl chloride and phenol [82]. The main composition of the CSE sample was different and contained acetonitrile, acetone, 2-hydroxy-2-methyl-propanenitrile and lower amounts of nicotine and nicotyrine [82]. Therefore, considering that the compounds in CSC and CSE are different, the effects (including toxicity) determined using CSC and CSE might differ. The following sections summarize the effects of CSC and CSE on biological functions, most of which are related to inflammation.

### 3.1. Cigarette Smoking and Nuclear Factor- $\kappa\text{B}$ (NF- $\kappa\text{B}$ )

Concerning the effects of cigarette smoking on the NF- $\kappa\text{B}$  pathway, Anto et al. found that the CSC-mediated induction of cyclooxygenase-2 was associated with activation of NF- $\kappa\text{B}$  through phosphorylation and degradation of I $\kappa\text{B}$ ( $\alpha$ ) [56]. The proteasome-linked degradation of I $\kappa\text{B}$ ( $\alpha$ ) causes the translocation of NF- $\kappa\text{B}$  to the nucleus and the transcriptional activation of NF- $\kappa\text{B}$ -dependent genes [83–87]. Activation of NF- $\kappa\text{B}$  by cigarette smoke was also reported by Zhang et al. [88] and by Wang et al. [89]. Accordingly, products from the natural world targeting the NF- $\kappa\text{B}$  signaling pathway are of great interest and should be considered as potential anti-inflammatory agents for mitigating the effects of cigarette smoking [90–93]. For instance, Wang et al. reported that ghrelin inhibits interleukin-6 production induced by cigarette smoke extract (CSE) and this inhibition is based on targeting the NF- $\kappa\text{B}$  pathway [90]. In our own laboratory, we found that the NF- $\kappa\text{B}$  inhibitor corilagin attenuates the loss of cellular junctions induced by cigarette smoke in epithelial lung cells [93].

### 3.2. Cigarette Smoke and Toll-like Receptor-4 (TLR4)

Nadigel et al. have reported that cigarette smoke increases TLR4 and TLR9 expression, thereby inducing increased cytokine production [61]. Interestingly, increased TLR4

expression was found in tissues of mice exposed to acute levels of cigarette smoke, and this was associated with lung inflammation [91,94]. Notably, elevated TLR4 and MMP-1 levels were found in lungs from smokers [94]. In conclusion, there is a general agreement on the fact that cigarette-smoking-related effects are mediated by activation of TLR-4 [58–63,94]. Accordingly, TLR4 inhibitors are expected to attenuate the acute cigarette-smoke-induced pulmonary inflammation [94,95]. As a representative and informative example, the TLR4 inhibitor TAK-242 (resatorvid) was administered by Wang et al. to mice exposed to cigarette smoke [91]. TAK-242 is a cyclohexane selected for inhibition of TLR4 [96]. It binds to the cysteine residue 747, preventing TLR4 binding with the toll-interleukin-1 receptor (TIR) domain-containing adaptor protein (TIRAP) [97] and downstream signal transduction. In the study by Wang et al., it was found to be very effective in mitigating the effects of exposure of mice to cigarette smoking. In fact, TAK-242 significantly decreased the accumulation of macrophages, neutrophils, lymphocytes and dendritic cells and the upregulation of IL-6, IL-8 and TNF- $\alpha$  in BAL fluid and lungs of the cigarette-smoke-exposed mice [91]. The results of this study demonstrated that the release of various inflammatory mediators is inhibited by TAK-242; notably, TAK-242 suppressed in lungs the expression of TLR4 and MyD88 as well as the activation of NF- $\kappa$ B [91]. These findings support the concept that TAK-242-mediated inhibition of cigarette smoke effects is associated with alterations of the TLR4/NF- $\kappa$ B signal pathway. Accordingly, TAK-242 can be proposed as a potent therapeutic agent in the treatment of cigarette-smoke-induced pulmonary inflammation.

### 3.3. Cigarette Smoke and Increased Release of Pro-Inflammatory Proteins

Fully in agreement with the effects of cigarette smoke on the TLR/NF- $\kappa$ B axis (see Sections 3.1 and 3.2), cigarette smoke regulates the production of pro-inflammatory cytokines and chemokines by several in vitro cellular model systems [53,60,98–103]. Induced pro-inflammatory proteins include IL-6, TNF- $\alpha$ , IL-1 $\beta$ , IL-8, G-CSF, GM-CSF and MCP-1. For instance, Mio et al. reported that cigarette smoke induces IL-8 release from human bronchial epithelial cells [98]. Remarkably, cigarette smoke induced IL-8, but inhibits eotaxin and RANTES release from airway smooth muscle [104].

### 3.4. Cigarette Smoke and Apoptosis

Several reports are available on the induction of apoptosis with tobacco smoke and related products. Ramage et al. studied the induction of apoptosis using A549 lung epithelial cells as an in vitro model system [105]. In their study, A549 cells were treated with tobacco smoke condensate and apoptosis was measured morphologically following staining of cells with DAPI. In addition, activation of Bax-alpha, an early event in the apoptotic process, was measured; the results demonstrated that tobacco smoke was able to initiate apoptosis in A549 airway epithelial cells and this resulted in a cell detachment and full apoptosis. Cigarette-smoke-induced apoptosis was also demonstrated in alveolar epithelial cells [106], endothelial cells [107,108] and Raw264.7 cells [109]. Concerning cigarette-smoke-induced apoptosis, Banerjee et al. reported the very interesting observation that it was prevented by black tea in a guinea pig “in vivo” model system, associated with prevention of lung damage [110].

### 3.5. Cigarette-Smoke-Induced Formation of Reactive Oxygen Species (ROS)

Cigarette smoke (CS) promotes ROS formation in different ways [111,112]. First of all, ROS, as well as radicals, are intrinsically present in CS [113–115]. In addition, CS constituents generate ROS through chemical reactions with biomolecules (quinones, redox-active metals, peroxy acids). For example, benzosemiquinones can penetrate the blood–air barrier and gain access to the blood circulation, thereby consistently producing superoxide through quinone redox cycling and forming adducts with biomolecules, such

as hemoglobin and albumin [116,117]. Furthermore, CS stimulates cellular ROS sources (NOX, mitochondria, uncoupled eNOS) to enhance ROS production [112,118]. Finally, CS components (such as ethyl vinyl chetone, chrotonaldehyde, acrolein) disrupts the antioxidant system, aggravating ROS generation and functions [112,119,120].

#### 4. Natural Products for the Mitigation of Toxic Biological Effects of Cigarette Smoke

The impact of natural products in preventing some of the more common detrimental effects of cigarette smoke is very high due to the low cost of these medical interventions, thereby allowing their use in developing low-income countries. A comprehensive review focusing on the protective effects of medicinal plants against cigarette smoke has been published by Tabeshpour et al. [19]. In this respect, Oriola and Oyedeji reviewed plant-derived natural products as useful agents against common respiratory diseases caused by cigarette smoke [121] (see Figure 1).

In this section, we will discuss some of the available examples showing the validated use of natural products for protecting cells or tissue against cigarette smoking and supporting the use of garlic-derived products (such as Aged Garlic Extract, S-allyl-cysteine and S1-propenyl-cysteine) for mitigating the effects of cigarette smoking both “in vitro” and “in vivo”.

##### 4.1. Silymarin

Silymarin is a flavonolignan extracted from *Silybum marianum* (milk thistle seeds) reported to exhibit a broad spectrum of biological and pharmacological properties, including antioxidant, antiviral, anticancer and immunomodulatory activities [122]. Li et al. have reported that silymarin attenuates cigarette-smoke-extract-induced inflammation via simultaneous inhibition of autophagy and the ERK/p38 MAPK pathway in human bronchial epithelial cells “in vitro” [122]. In another study, the effects of silymarin were analyzed “in vivo”, demonstrating silymarin as a powerful inhibitor of airway inflammation induced by cigarette smoke in mice [123]. Silymarin pretreatment dampened the secretion of TNF- $\alpha$ , IL-1 $\beta$  and IL-8 in BALF. These results suggest that silymarin attenuated inflammation and oxidative stress induced by cigarette smoke.

##### 4.2. Eucalyptol

1,8-cineole (Eucalyptol), a naturally occurring compound derived from botanical sources such as *Eucalyptus globulus*, *Rosmarinus officinalis* and Camphor laurel (*Cinnamomum camphora*), has a long history of use in traditional medicine and exhibits an array of biological properties, including anti-inflammatory, antioxidant, antimicrobial, bronchodilatory and analgesic effects [124]. Recent evidence has also indicated its potential role in managing conditions such as Alzheimer’s disease, neuropathic pain and cancer [125]. Eucalyptol suppresses lipopolysaccharide (LPS)-induced production of proinflammatory cytokines through an action on NF- $\kappa$ B, TNF- $\alpha$ , IL-1 $\beta$  and IL-6 as well as the extracellular signal-regulated kinase (ERK) pathway [125]. Eucalyptol was found to modulate CSE-induced human bronchial epithelial cell damage [126]. Accordingly, Yu et al. reported that treatment of rats exposed to cigarette smoke (CS) with eucalyptol mitigates CS-induced lung injury by suppressing ICAM-1 gene expression [127]. In addition, Kennedy-Feitosa et al. reported that eucalyptol inhibits lung inflammation and oxidative stress and promotes lung repair in mice following cigarette-smoke-induced emphysema [21,128].

##### 4.3. Curcumin

Curcumin is a constituent (up to ~5%) of the traditional medicine known as turmeric [129,130]. Interest in the therapeutic use of turmeric and the relative ease of

isolation of curcuminoids has led to their extensive investigation [130]. A comprehensive review on the protective effects of curcumin against cigarette-smoke-induced toxicity is available [131], and research articles reported that curcumin and liposomal curcumin inhibit cigarette-smoke-induced senescence and inflammation in human bronchial epithelial cells [132]. This effect is associated with a reduction in the expression of cigarette-smoke-extract-induced inflammatory markers IL-8 and IL-24 in vitro [133] through the modulation of the PPAR $\gamma$ -NF- $\kappa$ B signaling pathway [134].

#### 4.4. Taraxasterol

Taraxasterol is a pentacyclic-triterpene extracted from *Taraxacum officinalis* exhibiting anti-inflammatory properties [135]. Using lipopolysaccharide (LPS)-stimulated RAW264.7 cell as experimental model system, taraxasterol was reported as suppressing inflammatory cytokines, COX-2 and iNOS expression [136]. Xueshibojie et al. reported that taraxasterol inhibits CS-induced lung inflammation, ROS generation, IL-8 production, NF- $\kappa$ B activation, and TLR4 recruitment into lipid rafts [137].

#### 4.5. Sulforaphane

The isothiocyanate sulforaphane (SFN) is one of the most abundant bioactive components of Brassicaceae (for example, broccoli) [138]. As extensively reported in previous studies, SFN exhibits a wide range of biological effects including anticancer, antioxidant, antimicrobial, neuroprotective, cardioprotective and anti-inflammatory activities [139]. As demonstrated by several studies, the anti-inflammatory activity of SFN is mediated by NF- $\kappa$ B inhibition [140,141]. Published research results are available demonstrating that sulforaphane protects alveolar epithelial cells against injury caused by cigarette smoke extract (CSE). In a first report, SFN was demonstrated to inhibit de novo synthesis of IL-8 and MCP-1 induced in human epithelial cells by CSE [142]. In another study, SFN was found to exhibit a protective role on CSE-exposed alveolar epithelial cells through an increase in Nrf2 expression [143,144].

#### 4.6. Corilagin

The polyphenol corilagin is extracted from different plants, including *Phyllanthus urinaria* [145], *Dimocarpus longan* [146] and *Geranium thunbergii* [147]. The beneficial effects of this natural compound in cardiovascular disorders, hypertension, thrombosis and atherosclerosis have been reported [145]. Zhao et al. have demonstrated that the anti-inflammatory properties of corilagin are based on a block of NF- $\kappa$ B activation and its nuclear translocation [148]. In agreement, corilagin decreases the production of pro-inflammatory proteins, such as TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IL-8, iNOS and COX-2 [148]. In addition, corilagin inhibits ROS production from leukocytes as well as the formation of free radicals and lipid peroxidation in mitochondria [149,150]. In the study by Muresan et al., corilagin was found to mitigate the loss of cellular junctions induced in epithelial lung cells by cigarette smoke [93]. The results of this study demonstrated that CS induced the loss of cellular junctions in lung epithelium, possibly as a consequence of Cx-4HNE adduct formation, and corilagin was shown to be able to abolish these CS-induced alterations [93].

#### 4.7. Trans-4,4'-dihydroxystilbene

Trans-4,4'-dihydroxystilbene (DHS) is an analogue of the naturally occurring hydroxystilbene, resveratrol (3,4',5-trihydroxystilbene, Resv), present in grape skins, red wines and grape juices. These molecules are widely accepted as very interesting because of their diverse pharmacological attributes [151]. Wang et al. found that 4,4'-dihydroxystilbene ameliorates cigarette-smoke-induced progression of chronic obstructive pulmonary disease via inhibiting oxidative stress and inflammatory response [152]. This study demonstrated

that DHS attenuates the CS-induced pulmonary impairments through inhibition of oxidative stress and inflammatory responses targeting Nrf2 and NF- $\kappa$ B “in vitro” and “in vivo”, and could be developed into a preventive agent against pulmonary impairments induced by CS [152].

#### 4.8. Other Example of Natural Products Against CS Effects

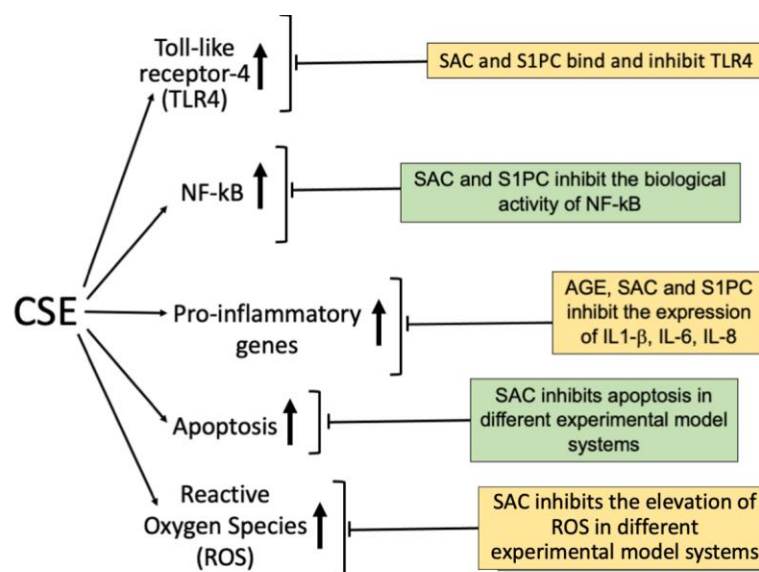
Several studies support the concept that natural products from medicinal plants alleviate cigarette-smoke-induced acute lung injury. Here are some examples. Liaqat et al. demonstrated that *Lavandula stoechas* significantly alleviates cigarette-smoke-induced acute lung injury via modulation of oxidative stress and the NF- $\kappa$ B pathway [153]. Similarly, Husain et al. found that *Cichorium intybus* L. significantly alleviates cigarette-smoke-induced effects by lowering NF- $\kappa$ B pathway activation and inflammatory mediators [154]. Inhibition of the NF- $\kappa$ B pathway was also demonstrated as the mechanism of action explaining the anti-inflammatory and anti-oxidant properties of *Ipomoea nil* (Linn.) Roth [155]. Furthermore, examples of reversion of the detrimental effects of cigarette smoke were found using propolis [156], mate tea [157] and grape skin extracts [158].

### 5. Aged Garlic Extract and Its Bioactive Components: Candidates for Mitigating the Cigarette Smoking Effects

Among a large variety of natural products of biomedical relevance, garlic-based products have recently gained great attention [159,160]. Among these products, AGE (aged garlic extract) is well known and has been studied in detail [161]. AGE is a commercially available odorless preparation obtained by immersing fresh garlic in 15% aqueous ethanol solution over a prolonged period of time (up to 20 months) at room temperature [161–165]. This natural product has been shown to possess immunomodulatory and anticancer properties [160,161].

The chemical composition of garlic and AGE has been described by Kodera et al. [166], Borek [167], Ryu et al. [168] and El-Saadony et al. [169]. In particular, Kodera et al. focused on the number of the compounds present in AGE, their changes in content during the aging process, and their production mechanisms involving various chemical and enzymatic reactions [166]. The beneficial effects of garlic have been attributed to several bioactive compounds, including lipid-soluble allyl sulfur compounds (e.g., diallyl sulfide, diallyl disulfide and diallyl trisulfide) and water-soluble compounds such as S-allyl-cysteine (SAC), S-allylmercaptocysteine (SAMC) and S1-propenyl-cysteine (S1PC) [162–166]. In particular, water-soluble compounds (such as SAC and S1PC) are of interest, considering their high oral bioavailability, favorable pharmacokinetics and tissue distribution, which facilitate their clinical applications [170]. In this review, among the variety of chemical components [166], we focused on SAC and S1PC. These bioactive compounds might be extracted from AGE by unique manufacturing processes [165].

The anti-inflammatory Aged Garlic Extract (AGE) and its major bioactive components might be of great interest for mitigating the effects of cigarette smoking. The key reasons for proposing this application are summarized in Figure 3.



**Figure 3.** Biological features of AGE and AGE constituents SAC and S1PC supporting their use for mitigating the effects of cigarette smoke.

Notably, CS has been shown to induce a chronic inflammation. In this respect, several studies have revealed that AGE and its key components are potent anti-inflammatory agents, both “in vitro” and “in vivo” [171]. Furthermore, CS induced the TLR4/NF-κB pathway (see Sections 3.1 and 3.2). In this respect, the AGE component S-allyl-cysteine (SAC) and S1-propenyl-cysteine bind to TLR4 [172–174] and inhibit NF-κB [171,175]. These findings should be further confirmed, since the cited studies have been performed using different methodological approaches and different cellular model systems, such as bronchial epithelial cells [171–173], chondrocytes [174] and T lymphoid cells [175].

A further consideration concerns the effects of CS on the expression of pro-inflammatory genes. CS induces IL-6, IL-8, IL-1 $\beta$  and several pro-inflammatory genes [60,98–104], and this effect appears to be selective. For instance, Oltmanns et al. reported that cigarette smoke induces IL-8, but inhibits eotaxin and RANTES release from airway smooth muscle [104].

We and several other research groups have clearly shown that AGE and the AGE components SAC and S1PC inhibit the expression of pro-inflammatory genes (such as IL-1 $\beta$ , IL-6, IL-8 and G-CSF) by targeting the TLR4 receptor [172–174] and the NF-κB pathway [171,175]. A consideration should also be made concerning the CS-mediated induction of apoptosis [105–110], as outlined in Section 3.4. Notably, Ramage et al. reported induction of apoptosis with tobacco smoke and related products in A549 lung epithelial cells in vitro [105]. In this respect, reports underlining the effects of garlic compounds on induced apoptosis in several cellular model systems are available [176–179]. Finally, CS induces Reactive Oxygen Species (ROS) [112–115], and this is strongly associated with oxidative stress and human diseases [180,181]. In this respect, S-allyl-cysteine is a strong inhibitor of ROS formation [182–185].

In this respect, we have to underline that few studies are available regarding the effects of garlic compounds on the biological effects caused by cigarette smoking. One of these studies has been reported by Hudlikar et al. in 2023 [64]. In this important study, the authors analyzed the effects of garlic compounds on transcriptomic changes induced in normal human lung epithelial Beas-2b cells by long-term exposure to cigarette smoke condensate (CSC). The effects of the organosulfur garlic compounds diallyl sulfide (DAS) and diallyl disulfide (DADS) were studied by Next Generation Sequencing (NGS) transcriptomic analysis. It was found that CSC regulated 1077 genes, including 36 genes

modulated by DAS and 101 genes modulated by DADS [64]. The conclusion of this study was that CSC induces global gene expression changes which can be delayed with DS and DADS dietary phytochemicals [64]. This study therefore supports the concept that garlic compounds, including aged garlic extract, should be carefully analyzed for mitigation of the effects of cigarette smoke.

The industrial interest in AGE and AGE-related products is documented by the fact that AGE is proposed and commercialized by several pharmaceutical companies, including for example Wakunaga Pharmaceuticals, Ltd (Hiroshima, Japan) (Kyolic® Aged Garlic Extract), Evergreen Health Foods, Galway, Ireland (Quest Kyolic Aged Garlic Extract), Shaanxi Tianrun Phytochemical Co., Ltd, Xi'an, China (Garlic Extract, Allicin), Best Pharmacy.gr, Crete, Greece (Quest Kyolic Garlic) and Bizen Chemical Co., Ltd, Okayama, Japan (High SAC-Content Garlic). Notably, a trademark for S1-propenylcysteine (S1PC™) has been recently obtained by Wakunaga Pharmaceuticals (registered on 9 July 2024; <https://branddb.wipo.int/>; accessed on 7 May 2025).

The industrial impact of AGE and AGE-related products is demonstrated by patents and patent applications focusing on these products. For instance, US8187654B2 (Title: Process for preparing aged garlic; Assignee: Blackgarlic Inc., Hayward, CA, USA) concerns a method of producing aged garlic in which its antioxidative capability is significantly increased as compared to that of raw garlic, which is used as a raw material. Methods for preparing aged garlic are described also in US20110293803, CN110623255A and EP1752051A1, as reported by Agostinelli et al. [171].

The possible transfer of the results concerning AGE and AGE-related products from bench to the bedside is supported by the growing number of clinical trials. For instance, NCT1950646 (The Effect of AGE on the Immune System -EAGESIS II; sponsor University of Florida; last updated 26 February 2016) demonstrated that AGE consumption modulated immune cell distribution, prevented the increase in serum TNF- $\alpha$  and IL-6 concentrations and reduced blood LDL concentration in adults with obesity [186]. A further example is NCT03860350 (Aged Garlic Extract Study – AGE; sponsor Lund University Hospital; last updated 11 June 2019) demonstrating that AGE, supplemented with B vitamins, folic acid and L-arginine retards the progression of subclinical atherosclerosis [187]. Moreover, the same NCT03860350 trial found that AGE reduced IL-6 in females with a low risk of cardiovascular diseases [188]. Relevant to this review, NCT02019368 (A Randomized, Double-blind, Placebo Controlled, Crossover Study to Evaluate the Antioxidant Effect of Aged Garlic Extract in Heavy Smokers; sponsor Hiroshima University; last updated 19 August 2015) compared the oxidative status of heavy smokers with that of non-smokers and determined the antioxidant effect of aged garlic extract (<https://clinicaltrials.gov>; accessed on 18 July 2025). Inclusion criteria were smoking (at least 20 cigarettes per day) or non-smoking (more than 20 years). In this study, smoker subjects were programmed to take 1.5 g of the dietary supplement aged garlic extract in six capsules once a day for 4 weeks. The primary outcome measure was urinary 8-hydroxydeoxyguanosine (8-OHdG), taken every 4 weeks (overall 12 weeks). Despite the fact that information on the results obtained is still not available, the activation of this clinical trial (60 subjects were enrolled according to the 17 August 2015 status report) demonstrates the interest in studying AGE in clinical settings for heavy cigarette smokers. Based on the little information discussed in the present review, further pre-clinical studies and clinical trials are highly warranted.

A final comment concerns the very interesting possibility that the best effects on CS-induced alterations occur when natural products are employed in combination. In the study performed by Reis et al., eucalyptol and curcumin used in combination exhibited the highest efficiency in modulating cigarette-smoke-extract-induced human bronchial epithelial damage [126]. Therefore, combined use of eucalyptol and curcumin might be

a potential therapeutic against smoking-induced lung diseases through antioxidant and inflammatory pathways [126]. Moreover, possible combinations using RNA/DNA-based drugs and natural products should be considered in the future. In this respect, aged garlic extract was recently proposed in combined treatments with microRNA miR-93-5p, previously demonstrated to inhibit TLR4, NF- $\kappa$ B and IL-8 gene expression [189]. This study provided preliminary evidence suggesting that the miR-93-5p-based miRNA therapeutics could be combined with the anti-inflammatory aged garlic extract (AGE) to more effectively inhibit IL-8 gene expression [189].

## 6. Conclusions

The conclusion of this review article is that the available information strongly indicates a possible use of the anti-inflammatory aged garlic extract (AGE) and its bioactive components S-allyl-cysteine (SAC) and S1-propenyl-cysteine (S1PC) for mitigating the detrimental effects of cigarette smoke on human tissues. Notably, the reported bioactive concentrations of AGE and AGE components are highly variable both “in vitro” and “in vivo”, depending on the biomarker analyzed, the methods employed for the analysis and the model system and administration protocol used. For instance, bioactive SAC concentrations were reported to vary between 10–200  $\mu$ M [172–174,182] and 2–20 mM [164,165,179]. Therefore, preliminary studies are necessary to determine the optimal concentration to be used. In the representative clinical trials cited in this review, effective AGE concentrations employed varied between 250 mg daily [187] and 2400–3600 mg daily [186,188].

The key reasons for proposing AGE, S-allyl-cysteine (SAC) and S1-propenyl-cysteine (S1PC) for mitigating cigarette smoke effects are the following (summarized in Figure 3). First of all, AGE and its key components are potent anti-inflammatory agents, both “in vitro” and “in vivo”. Second, “in silico” and bio-molecular analyses indicate that the AGE bioactive components SAC and S1PC bind to TLR4, inhibit NF- $\kappa$ B and induce a decrease in the expression of pro-inflammatory genes. Furthermore, AGE and AGE components revert apoptosis induced by cigarette smoke in several cellular model systems. Finally, S-allyl-cysteine is a strong inhibitor of ROS formation. All the biological pathways mentioned are strongly induced by cigarette smoke in several cellular model systems (Figure 3). Experimental projects to verify this very interesting possibility are highly warranted, considering the impact of tobacco smoke on the health system (see Figure 1) [3–9]. It should be considered that healthcare spending attributable to cigarette smoking is very high [10,11] and several actions have been considered to help eliminate smoking [12–15], such as bans of tobacco advertising [16] and introduction of taxes as a share of cigarette price [17,18]. These smoking cessation interventions are important [14–16,190], even if difficulty in quitting smoking might be encountered [15,16]. In this context, strategies in preventing or mitigating the effects of tobacco abuse (such as those based on natural products, including aged garlic extracts and AGE components) are of great interest, considering the world-wide distribution of tobacco abuse [1].

**Author Contributions:** Conceptualization, formal analysis, resources, writing—original draft preparation, writing—review and editing, project administration: A.F. and R.G. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** No new data were created or analyzed in this study; additional information will be shared upon request to the corresponding authors.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## Abbreviations

The following abbreviations are used in this manuscript:

CS	Cigarette Smoke
CSC	Cigarette Smoke Condensate
CSE	Cigarette Smoke Extract
VOC	Volatile Organic Compound
NF- $\kappa$ B	Nuclear Factor-kappa-B
TLR4	Toll-like Receptor-4
Nrf2	Nuclear Factor Erythroid 2-related factor 2
IL	Interleukin
ROS	Reactive Oxygen Species
AGE	Aged Garlic Extract
SAC	S-allyl-cysteine
SIPC	S1-propenyl-cysteine
SFN	Sulforaphane
DHS	Trans-4,4'-dihydroxystilbene
COPD	Chronic Obstructive Pulmonary Disease
CF	Cystic Fibrosis
BAL	Bronchoalveolar lavage
WHO	World Health Organization

## References

1. World Health Organization. *WHO Global Report on Trends in Prevalence of Tobacco Use 2000–2030*; World Health Organization: Geneva, Switzerland, 2024.
2. Dai, X.; Gakidou, E.; Lopez, A.D. Evolution of the global smoking epidemic over the past half century: Strengthening the evidence base for policy action. *Tob. Control* **2022**, *31*, 129–137. [[CrossRef](#)]
3. Varghese, J.; Muntode Gharde, P. A Comprehensive Review on the Impacts of Smoking on the Health of an Individual. *Cureus* **2023**, *15*, e46532. [[CrossRef](#)]
4. Vassallo, R. Diffuse lung diseases in cigarette smokers. *Semin. Respir. Crit. Care Med.* **2012**, *33*, 533–542. [[CrossRef](#)]
5. Kondo, T.; Nakano, Y.; Adachi, S.; Murohara, T. Effects of tobacco smoking on cardiovascular disease. *Circ. J.* **2019**, *83*, 1980–1985. [[CrossRef](#)]
6. Durazzo, T.C.; Mattsson, N.; Weiner, M.W. Smoking and increased Alzheimer’s disease risk: A review of potential mechanisms. *Alzheimers Dement.* **2014**, *10*, 122–145. [[CrossRef](#)]
7. Sloan, A.; Hussain, I.; Maqsood, M.; Eremin, O.; El-Sheemy, M. The effects of smoking on fracture healing. *Surgeon* **2010**, *8*, 111–116. [[CrossRef](#)]
8. Liu, Y.; Lu, L.; Yang, H.; Wu, X.; Luo, X.; Shen, J.; Xiao, Z.; Zhao, Y.; Du, F.; Chen, Y.; et al. Dysregulation of immunity by cigarette smoking promotes inflammation and cancer: A review. *Environ. Pollut.* **2023**, *339*, 122730. [[CrossRef](#)]
9. Walser, T.; Cui, X.; Yanagawa, J.; Lee, J.M.; Heinrich, E.; Lee, G.; Sharma, S.; Dubinett, S.M. Smoking and lung cancer: The role of inflammation. *Proc. Am. Thorac. Soc.* **2008**, *5*, 811–815. [[CrossRef](#)]
10. Xu, X.; Shrestha, S.S.; Trivers, K.F.; Neff, L.; Armour, B.S.; King, B.A. U.S. healthcare spending attributable to cigarette smoking in 2014. *Prev. Med.* **2021**, *150*, 106529. [[CrossRef](#)]
11. Gu, D.; Sung, H.Y.; Calfee, C.S.; Wang, Y.; Yao, T.; Max, W. Smoking-Attributable Health Care Expenditures for US Adults With Chronic Lower Respiratory Disease. *JAMA Netw. Open* **2024**, *7*, e2413869. [[CrossRef](#)]
12. Bancej, C.; O’Loughlin, J.; Platt, R.W.; Paradis, G.; Gervais, A. Smoking cessation attempts among adolescent smokers: A systematic review of prevalence studies. *Tob. Control* **2007**, *16*, e8. [[CrossRef](#)]
13. Torchalla, I.; Okoli, C.T.; Böttorff, J.L.; Qu, A.; Poole, N.; Greaves, L. Smoking cessation programs targeted to women: A systematic review. *Women Health* **2012**, *52*, 32–54. [[CrossRef](#)]
14. Aveyard, P.; Begh, R.; Parsons, A. Brief opportunistic smoking cessation interventions: A systematic review and meta-analysis to compare advice to quit and offer of assistance. *Addiction* **2012**, *107*, 1066–1073. [[CrossRef](#)] [[PubMed](#)]
15. Komiyama, M.; Takahashi, Y.; Tateno, H.; Mori, M.; Nagayoshi, N.; Yonehara, H.; Nakasa, N.; Haruki, Y.; Hasegawa, K. Support for Patients Who Have Difficulty Quitting Smoking: A Review. *Intern. Med.* **2019**, *58*, 317–320. [[CrossRef](#)]

16. Saad, C.; Cheng, B.H.; Takamizawa, R.; Thakur, A.; Lee, C.W.; Leung, L.; Veerman, J.L.; Aminde, L.N. Effectiveness of tobacco advertising, promotion and sponsorship bans on smoking prevalence, initiation and cessation: A systematic review and meta-analysis. *Tob. Control* **2025**. [[CrossRef](#)]
17. Siddiqi, K.; Elsey, H.; Khokhar, M.A.; Marshall, A.M.; Pokhrel, S.; Arora, M.; Crankson, S.; Mehra, R.; Morello, P.; Collin, J.; et al. Framework Convention on Tobacco Control 2030-A Program to Accelerate the Implementation of World Health Organization Framework Convention for Tobacco Control in Low- and Middle-Income Countries: A Mixed-Methods Evaluation. *Nicotine Tob. Res.* **2023**, *25*, 1074–1081. [[CrossRef](#)]
18. Lahiri, S.; Bingenheimer, J.B.; Evans, W.D.; Wang, Y.; Cislighi, B.; Dubey, P.; Snowden, B. Understanding the mechanisms of change in social norms around tobacco use: A systematic review and meta-analysis of interventions. *Addiction* **2025**, *120*, 215–235. [[CrossRef](#)] [[PubMed](#)]
19. Tabeshpour, J.; Asadpour, A.; Norouz, S.; Hosseinzadeh, H. The protective effects of medicinal plants against cigarette smoking: A comprehensive review. *Phytomedicine* **2024**, *135*, 156199. [[CrossRef](#)]
20. Hsu, C.L.; Wu, Y.L.; Tang, G.J.; Lee, T.S.; Kou, Y.R. Ginkgo biloba extract confers protection from cigarette smoke extract-induced apoptosis in human lung endothelial cells: Role of heme oxygenase-1. *Pulm. Pharmacol. Ther.* **2009**, *22*, 286–296. [[CrossRef](#)]
21. Kennedy-Feitosa, E.; Okuro, R.T.; Pinho Ribeiro, V.; Lanzetti, M.; Barroso, M.V.; Zin, W.A.; Porto, L.C.; Brito-Gitirana, L.; Valença, S.S. Eucalyptol attenuates cigarette smoke-induced acute lung inflammation and oxidative stress in the mouse. *Pulm. Pharmacol. Ther.* **2016**, *41*, 11–18. [[CrossRef](#)]
22. Sasco, A.J.; Secretan, M.B.; Straif, K. Tobacco smoking and cancer: A brief review of recent epidemiological evidence. *Lung Cancer* **2004**, *45* (Suppl. S2), S3–S9. [[CrossRef](#)]
23. Sparrow, D.; Dawber, T.R. The influence of cigarette smoking on prognosis after a first myocardial infarction: A report from the Framingham Study. *J. Chronic Dis.* **1978**, *31*, 425–432. [[CrossRef](#)]
24. Shinton, R.; Beevers, G. Meta-analysis of relation between cigarette smoking and stroke. *BMJ* **1989**, *298*, 789–794. [[CrossRef](#)]
25. Ryu, J.H.; Colby, T.V.; Hartman, T.E.; Vassallo, R. Smoking-related interstitial lung diseases: A concise review. *Eur. Respir. J.* **2001**, *17*, 122–132. [[CrossRef](#)] [[PubMed](#)]
26. Maddatu, J.; Anderson-Baucum, E.; Evans-Molina, C. Smoking and the risk of type 2 diabetes. *Transl. Res.* **2017**, *184*, 101–107. [[CrossRef](#)] [[PubMed](#)]
27. Laniado-Laborín, R. Smoking and chronic obstructive pulmonary disease (COPD). Parallel epidemics of the 21 century. *Int. J. Environ. Res. Public Health* **2009**, *6*, 209–224. [[CrossRef](#)]
28. Edderkaoui, M.; Thrower, E. Smoking and Pancreatic Disease. *J. Cancer Ther.* **2013**, *4*, 34–40. [[CrossRef](#)]
29. Roser, M. “Smoking: How Large of a Global Problem Is It? And How Can We Make Progress Against It?”. 2021. Available online: <https://ourworldindata.org/smoking-big-problem-in-brief#> (accessed on 28 May 2025).
30. Phua, Z.J.; MacInnis, R.J.; Jayasekara, H. Cigarette smoking and risk of second primary cancer: A systematic review and meta-analysis. *Cancer Epidemiol.* **2022**, *78*, 102160. [[CrossRef](#)]
31. Inoue-Choi, M.; Hartge, P.; Liao, L.M.; Caporaso, N.; Freedman, N.D. Association between long-term low-intensity cigarette smoking and incidence of smoking-related cancer in the national institutes of health-AARP cohort. *Int. J. Cancer* **2018**, *142*, 271–280. [[CrossRef](#)] [[PubMed](#)]
32. Khani, Y.; Pourgholam-Amiji, N.; Afshar, M.; Otroshi, O.; Sharifi-Esfahani, M.; Sadeghi-Gandomani, H.; Vejdani, M.; Salehiniya, H. Tobacco Smoking and Cancer Types: A Review. *Biomed. Res. Ther.* **2018**, *5*, 2142–2159. [[CrossRef](#)]
33. Shi, H.; Shao, X.; Hong, Y. Association between cigarette smoking and the susceptibility of acute myeloid leukemia: A systematic review and meta-analysis. *Eur. Rev. Med. Pharmacol. Sci.* **2019**, *23*, 10049–10057.
34. Qi, K.; Cheng, H.; Jiang, Y.; Zheng, Y. Contribution of smoking to the global burden of bladder cancer from 1990 to 2021 and projections to 2046. *Tob. Induc. Dis.* **2025**, *23*, 44. [[CrossRef](#)]
35. Wen, Q.; Wang, X.; Lv, J.; Guo, Y.; Pei, P.; Yang, L.; Chen, Y.; Du, H.; Burgess, S.; Hacker, A.; et al. Association between involuntary smoking and risk of cervical cancer in Chinese female never smokers: A prospective cohort study. *Environ. Res.* **2022**, *212 Pt C*, 113371. [[CrossRef](#)]
36. Bener, A.; Öztürk, A.E.; Dasdelen, M.F.; Barisik, C.C.; Dasdelen, Z.B.; Agan, A.F.; De La Rosette, J.; Day, A.S. Colorectal cancer and associated genetic, lifestyle, cigarette, nargileh-hookah use and alcohol consumption risk factors: A comprehensive case-control study. *Oncol. Rev.* **2024**, *18*, 1449709. [[CrossRef](#)]
37. Islam, M.O.; Thangaretnam, K.; Lu, H.; Peng, D.; Soutto, M.; El-Rifai, W.; Giordano, S.; Ban, Y.; Chen, X.; Bilbao, D.; et al. Smoking induces WEE1 expression to promote docetaxel resistance in esophageal adenocarcinoma. *Mol. Ther. Oncolytics* **2023**, *30*, 286–300. [[CrossRef](#)]
38. Yang, X.; Chen, H.; Zhang, J.; Zhang, S.; Wu, Y.S.; Pang, J. Association of cigarette use with risk of prostate cancer among US males: A cross-sectional study from NHANES 1999–2020. *BMC Public Health* **2025**, *25*, 608. [[CrossRef](#)]

39. Campi, R.; Rebez, G.; Klatte, T.; Roussel, E.; Ouizad, I.; Ingels, A.; Pavan, N.; Kara, O.; Erdem, S.; Bertolo, R.; et al. Effect of smoking, hypertension and lifestyle factors on kidney cancer—perspectives for prevention and screening programmes. *Nat. Rev. Urol.* **2023**, *20*, 669–681. [[CrossRef](#)]
40. Zuo, J.J.; Tao, Z.Z.; Chen, C.; Hu, Z.W.; Xu, Y.X.; Zheng, A.Y.; Guo, Y. Characteristics of cigarette smoking without alcohol consumption and laryngeal cancer: Overall and time-risk relation. A meta-analysis of observational studies. *Eur. Arch. Otorhinolaryngol.* **2017**, *274*, 1617–1631. [[CrossRef](#)]
41. Lee, J.; Choi, J.Y.; Lee, S.K. Heavy smoking increases early mortality risk in patients with hepatocellular carcinoma after curative treatment. *J. Liver Cancer* **2024**, *24*, 253–262. [[CrossRef](#)]
42. Tang, F.H.; Wong, H.Y.T.; Tsang, P.S.W.; Yau, M.; Tam, S.Y.; Law, L.; Yau, K.; Wong, J.; Farah, F.H.M.; Wong, J. Recent advancements in lung cancer research: A narrative review. *Transl. Lung Cancer Res.* **2025**, *14*, 975–990. [[CrossRef](#)]
43. Pérez-Leal, M.; El Helou, B.; Roger, I. Electronic Cigarettes Versus Combustible Cigarettes in Oral Squamous Cell Cancer Patients: A Systematic Review. *J. Oral Pathol. Med.* **2025**, *54*, 199–206. [[CrossRef](#)]
44. Subhan, M.; Saji Parel, N.; Krishna, P.V.; Gupta, A.; Uthayaseelan, K.; Uthayaseelan, K.; Kadari, M. Smoking and Pancreatic Cancer: Smoking Patterns, Tobacco Type, and Dose-Response Relationship. *Cureus* **2022**, *14*, e26009. [[CrossRef](#)]
45. Li, L.F.; Chan, R.L.; Lu, L.; Shen, J.; Zhang, L.; Wu, W.K.; Wang, L.; Hu, T.; Li, M.X.; Cho, C.H. Cigarette smoking and gastrointestinal diseases: The causal relationship and underlying molecular mechanisms (review). *Int. J. Mol. Med.* **2014**, *34*, 372–380. [[CrossRef](#)] [[PubMed](#)]
46. Warren, G.W.; Cartmell, K.B.; Garrett-Mayer, E.; Salloum, R.G.; Cummings, K.M. Attributable Failure of First-line Cancer Treatment and Incremental Costs Associated With Smoking by Patients With Cancer. *JAMA Netw. Open* **2019**, *2*, e191703. [[CrossRef](#)] [[PubMed](#)]
47. Petrucci, C.M.; Hyland, A. Understanding the Financial Consequences of Smoking During Cancer Treatment in the Era of Value-Based Medicine. *JAMA Netw. Open* **2019**, *2*, e191713. [[CrossRef](#)]
48. Warren, G.W. Mitigating the adverse health effects and costs associated with smoking after a cancer diagnosis. *Transl. Lung Cancer Res.* **2019**, *8* (Suppl. S1), S59–S66. [[CrossRef](#)]
49. Isaranuwatthai, W.; de Oliveira, C.; Mittmann, N.; Evans, W.K.B.; Peter, A.; Truscott, R.; Chan, K.K. Impact of smoking on health system costs among cancer patients in a retrospective cohort study in Ontario, Canada. *BMJ Open* **2019**, *9*, e026022. [[CrossRef](#)]
50. Arrieta, O.; Quintana-Carrillo, R.H.; Ahumada-Curiel, G.; Corona-Cruz, J.F.; Correa-Acevedo, E.; Zinser-Sierra, J.; de la Mata-Moya, D.; Mohar-Betancourt, A.; Morales-Oyarvide, V.; Myriam Reynales-Shigematsu, L. Medical care costs incurred by patients with smoking-related non-small cell lung cancer treated at the National Cancer Institute of Mexico. *Tob. Induc. Dis.* **2015**, *12*, 25. [[CrossRef](#)]
51. Nguyen, T.X.T.; Han, M.; Oh, J.K. The economic burden of cancers attributable to smoking in Korea, 2014. *Tob. Induc. Dis.* **2019**, *17*, 15. [[CrossRef](#)] [[PubMed](#)]
52. Lee, J.; Taneja, V.; Vassallo, R. Cigarette smoking and inflammation: Cellular and molecular mechanisms. *J. Dent. Res.* **2012**, *91*, 142–149. [[CrossRef](#)]
53. Wang, H.; Chen, H.; Fu, Y.; Liu, M.; Zhang, J.; Han, S.; Tian, Y.; Hou, H.; Hu, Q. Effects of Smoking on Inflammatory-Related Cytokine Levels in Human Serum. *Molecules* **2022**, *27*, 3715. [[CrossRef](#)]
54. Elisia, I.; Lam, V.; Cho, B.; Hay, M.; Li, M.Y.; Yeung, M.; Bu, L.; Jia, W.; Norton, N.; Lam, S.; et al. The effect of smoking on chronic inflammation, immune function and blood cell composition. *Sci. Rep.* **2020**, *10*, 19480. [[CrossRef](#)]
55. Lugade, A.A.; Bogner, P.N.; Thatcher, T.H.; Sime, P.J.; Phipps, R.P.; Thanavala, Y. Cigarette smoke exposure exacerbates lung inflammation and compromises immunity to bacterial infection. *J. Immunol.* **2014**, *192*, 5226–5235. [[CrossRef](#)]
56. Anto, R.J.; Mukhopadhyay, A.; Shishodia, S.; Gairola, C.G.; Aggarwal, B.B. Cigarette smoke condensate activates nuclear transcription factor-kappaB through phosphorylation and degradation of IkappaB(alpha): Correlation with induction of cyclooxygenase-2. *Carcinogenesis* **2002**, *23*, 1511–1518. [[CrossRef](#)]
57. Kunnumakkara, A.B.; Shabnam, B.; Girisa, S.; Harsha, C.; Banik, K.; Devi, T.B.; Choudhury, R.; Sahu, H.; Parama, D.; Sailo, B.L.; et al. Inflammation, NF-κB, and Chronic Diseases: How are They Linked? *Crit. Rev. Immunol.* **2020**, *40*, 1–39. [[CrossRef](#)]
58. Doz, E.; Noulin, N.; Boichot, E.; Guénon, I.; Fick, L.; Le Bert, M.; Lagente, V.; Ryffel, B.; Schnyder, B.; Quesniaux, V.F.; et al. Cigarette smoke-induced pulmonary inflammation is TLR4/MyD88 and IL-1R1/MyD88 signaling dependent. *J. Immunol.* **2008**, *180*, 1169–1178. [[CrossRef](#)]
59. Karimi, K.; Sarir, H.; Mortaz, E.; Smit, J.J.; Hosseini, H.; De Kimpe, S.J.; Nijkamp, F.P.; Folkerts, G. Toll-like receptor-4 mediates cigarette smoke-induced cytokine production by human macrophages. *Respir. Res.* **2006**, *7*, 66. [[CrossRef](#)]
60. Sarir, H.; Mortaz, E.; Karimi, K.; Kraneveld, A.D.; Rahman, I.; Caldenhoven, E.; Nijkamp, F.P.; Folkerts, G. Cigarette smoke regulates the expression of TLR4 and IL-8 production by human macrophages. *J. Inflamm.* **2009**, *6*, 12. [[CrossRef](#)]
61. Nadigel, J.; Préfontaine, D.; Baglolle, C.J.; Maltais, F.; Bourbeau, J.; Eidelman, D.H.; Hamid, Q. Cigarette smoke increases TLR4 and TLR9 expression and induces cytokine production from CD8(+) T cells in chronic obstructive pulmonary disease. *Respir. Res.* **2011**, *12*, 149. [[CrossRef](#)]

62. Pace, E.; Ferraro, M.; Siena, L.; Melis, M.; Montalbano, A.M.; Johnson, M.; Bonsignore, M.R.; Bonsignore, G.; Gjomarkaj, M. Cigarette smoke increases Toll-like receptor 4 and modifies lipopolysaccharide-mediated responses in airway epithelial cells. *Immunology* **2008**, *124*, 401–411. [[CrossRef](#)]
63. Yeh, H.Y.; Hung, S.H.; Chen, S.C.; Guo, F.R.; Huang, H.L.; Peng, J.K.; Lee, C.S.; Tsai, J.S. The Expression of Toll-Like Receptor 4 mRNA in PBMCs Is Upregulated in Smokers and Decreases Upon Smoking Cessation. *Front. Immunol.* **2021**, *12*, 667460. [[CrossRef](#)]
64. Hudlikar, R.R.; Chou, P.J.; Kuo, H.D.; Sargsyan, D.; Wu, R.; Kong, A.N. Long term exposure of cigarette smoke condensate (CSC) mediates transcriptomic changes in normal human lung epithelial Beas-2b cells and protection by garlic compounds. *Food Chem. Toxicol.* **2023**, *174*, 113656. [[CrossRef](#)]
65. Khan, D.; Zhou, H.; You, J.; Kaiser, V.A.; Khajuria, R.K.; Muhammad, S. Tobacco smoke condensate-induced senescence in endothelial cells was ameliorated by colchicine treatment via suppression of NF- $\kappa$ B and MAPKs P38 and ERK pathways activation. *Cell Commun. Signal.* **2024**, *22*, 214. [[CrossRef](#)] [[PubMed](#)]
66. Thaiparambil, J.; Amara, C.S.; Sen, S.; Putluri, N.; El-Zein, R. Cigarette smoke condensate induces centrosome clustering in normal lung epithelial cells. *Cancer Med.* **2023**, *12*, 8499–8509. [[CrossRef](#)]
67. Gellner, C.A.; Reynaga, D.D.; Leslie, F.M. Cigarette Smoke Extract: A Preclinical Model of Tobacco Dependence. *Curr. Protoc. Neurosci.* **2016**, *77*, 9.54.1–9.54.10. [[CrossRef](#)]
68. Hirata, N.; Horinouchi, T.; Kanda, Y. Effects of cigarette smoke extract derived from heated tobacco products on the proliferation of lung cancer stem cells. *Toxicol. Rep.* **2022**, *9*, 1273–1280. [[CrossRef](#)]
69. Amel Al-Hashimi Shah, J.; Carpenter, R.; Morgan, W.; Meah, M.; Ruchaya, P.J. An In-Vitro Standardized Protocol for Preparing Smoke 1 Extract Media from Cigarette, Electronic Cigarette and 2 Waterpipe. *bioRxiv* **2024**. [[CrossRef](#)]
70. Kim, Y.-H.; Kim, M.-S. Development and assessment of a novel standardized method for preparation of whole cigarette smoke condensate (WCSC) for toxicity testing of cigarette smoke. *Microchem. J.* **2023**, *191*, 108914. [[CrossRef](#)]
71. Higashi, T.; Mai, Y.; Noya, Y.; Horinouchi, T.; Terada, K.; Hoshi, A.; Nepal, P.; Harada, T.; Horiguchi, M.; Hatate, C.; et al. A simple and rapid method for standard preparation of gas phase extract of cigarette smoke. *PLoS ONE* **2014**, *9*, e107856. [[CrossRef](#)]
72. Wright, C. Standardized methods for the regulation of cigarette-smoke constituents. *Trends Anal. Chem.* **2015**, *66*, 118–127. [[CrossRef](#)]
73. Mathewson, H.D. The Direct Preparation of Cigarette Smoke Condensate by High Velocity Impaction. *Contrib. Tob. Nicotine Res.* **1966**, *3*, 430–437. [[CrossRef](#)]
74. Agraval, H.; Sharma, J.R.; Yadav, U.C.S. Method of Preparation of Cigarette Smoke Extract to Assess Lung Cancer-Associated Changes in Airway Epithelial Cells. *Methods Mol. Biol.* **2022**, *2413*, 121–132.
75. Li, X. In vitro toxicity testing of cigarette smoke based on the air-liquid interface exposure: A review. *Toxicol. In Vitro* **2016**, *36*, 105–113. [[CrossRef](#)]
76. Singh, A.V.; Maharjan, R.S.; Kromer, C.; Laux, P.; Luch, A.; Vats, T.; Chandrasekar, V.; Dakua, S.P.; Park, B.W. Advances in Smoking Related In Vitro Inhalation Toxicology: A Perspective Case of Challenges and Opportunities from Progresses in Lung-on-Chip Technologies. *Chem. Res. Toxicol.* **2021**, *34*, 1984–2002. [[CrossRef](#)]
77. Horiyama, S.; Kunitomo, M.; Yoshikawa, N.; Nakamura, K. Mass Spectrometric Approaches to the Identification of Potential Ingredients in Cigarette Smoke Causing Cytotoxicity. *Biol. Pharm. Bull.* **2016**, *39*, 903–908. [[CrossRef](#)]
78. Fresenius, R.E. Analysis of tobacco smoke condensate. *J. Anal. Appl. Pyrolysis* **1985**, *8*, 561–575. [[CrossRef](#)]
79. Khattri, R.B.; Thome, T.; Fitzgerald, L.F.; Wohlgemuth, S.E.; Hepple, R.T.; Ryan, T.E. NMR Spectroscopy Identifies Chemicals in Cigarette Smoke Condensate That Impair Skeletal Muscle Mitochondrial Function. *Toxics* **2022**, *10*, 140. [[CrossRef](#)]
80. Liu, G.; Wang, R.; Chen, H.; Wu, P.; Fu, Y.; Li, K.; Liu, M.; Shi, Z.; Zhang, Y.; Su, Y.; et al. Non-nicotine constituents in cigarette smoke extract enhance nicotine addiction through monoamine oxidase A inhibition. *Front. Neurosci.* **2022**, *16*, 1058254. [[CrossRef](#)]
81. Park, J.M.; Jeong, H.; Seo, Y.S.; Do, V.Q.; Choi, S.J.; Lee, K.; Choi, K.C.; Choi, W.J.; Lee, M.Y. Cigarette Smoke Extract Produces Superoxide in Aqueous Media by Reacting with Bicarbonate. *Toxics* **2021**, *9*, 316. [[CrossRef](#)]
82. Kim, Y.H.; An, Y.J.; Jo, S.; Lee, S.H.; Lee, S.J.; Choi, S.J.; Lee, K. Comparison of volatile organic compounds between cigarette smoke condensate (CSC) and extract (CSE) samples. *Environ. Health Toxicol.* **2018**, *33*, e2018012-0. [[CrossRef](#)]
83. Sun, S.C.; Ley, S.C. New insights into NF- $\kappa$ B regulation and function. *Trends Immunol.* **2008**, *29*, 469–478. [[CrossRef](#)]
84. Hacker, H.; Karin, M. Regulation and function of IKK and IKK-related kinases. *Sci. STKE* **2006**, *2006*, re13. [[CrossRef](#)]
85. Chen, F.E.; Huang, D.B.; Chen, Y.Q.; Ghosh, G. Crystal structure of p50/p65 heterodimer of transcription factor NF- $\kappa$ B bound to DNA. *Nature* **1998**, *391*, 410–413. [[CrossRef](#)]
86. Hoffmann, A.; Natoli, G.; Ghosh, G. Transcriptional regulation via the NF- $\kappa$ B signaling module. *Oncogene* **2006**, *25*, 6706–6716. [[CrossRef](#)]
87. Mathes, E.; O’Dea, E.L.; Hoffmann, A.; Ghosh, G. NF- $\kappa$ B dictates the degradation pathway of IkappaBalpha. *EMBO J.* **2008**, *27*, 1357–1367. [[CrossRef](#)] [[PubMed](#)]

88. Zhang, C.; Qin, S.; Qin, L.; Liu, L.; Sun, W.; Li, X.; Li, N.; Wu, R.; Wang, X. Cigarette smoke extract-induced p120-mediated NF- $\kappa$ B activation in human epithelial cells is dependent on the RhoA/ROCK pathway. *Sci. Rep.* **2016**, *6*, 23131. [[CrossRef](#)] [[PubMed](#)]
89. Wang, V.; Heffer, A.; Roztocil, E.; Feldon, S.E.; Libby, R.T.; Woeller, C.F.; Kuriyan, A.E. TNF- $\alpha$  and NF- $\kappa$ B signaling play a critical role in cigarette smoke-induced epithelial-mesenchymal transition of retinal pigment epithelial cells in proliferative vitreoretinopathy. *PLoS ONE* **2022**, *17*, e0271950. [[CrossRef](#)]
90. Wang, H.; Yang, T.; Shen, Y.; Wan, C.; Li, X.; Li, D.; Liu, Y.; Wang, T.; Xu, D.; Wen, F.; et al. Ghrelin Inhibits Interleukin-6 Production Induced by Cigarette Smoke Extract in the Bronchial Epithelial Cell Via NF- $\kappa$ B Pathway. *Inflammation* **2016**, *39*, 190–198. [[CrossRef](#)]
91. Wang, D.; Tao, K.; Xion, J.; Xu, S.; Jiang, Y.; Chen, Q.; He, S. TAK-242 attenuates acute cigarette smoke-induced pulmonary inflammation in mouse via the TLR4/NF- $\kappa$ B signaling pathway. *Biochem. Biophys. Res. Commun.* **2016**, *472*, 508–515. [[CrossRef](#)]
92. Wang, L.; Meng, J.; Wang, C.; Wang, Y.; Yang, C.; Li, Y. Hydrogen sulfide attenuates cigarette smoke-induced pyroptosis through the TLR4/NF- $\kappa$ B signaling pathway. *Int. J. Mol. Med.* **2022**, *49*, 56. [[CrossRef](#)]
93. Muresan, X.M.; Cervellati, F.; Sticozzi, C.; Belmonte, G.; Chui, C.H.; Lampronti, I.; Borgatti, M.; Gambari, R.; Valacchi, G. The loss of cellular junctions in epithelial lung cells induced by cigarette smoke is attenuated by corilagin. *Oxid. Med. Cell. Longev.* **2015**, *2015*, 631758. [[CrossRef](#)]
94. Geraghty, P.; Dabo, A.J.; D'Armiento, J. TLR4 protein contributes to cigarette smoke-induced matrix metalloproteinase-1 (MMP-1) expression in chronic obstructive pulmonary disease. *J. Biol. Chem.* **2011**, *286*, 30211–30218. [[CrossRef](#)]
95. Zhang, F.; Geng, Y.; Shi, X.; Duo, J. EGR3 deficiency alleviates cigarette smoke-induced pulmonary inflammation in COPD through TLR4/NF- $\kappa$ B/TIMP-1 axis. *Biochem. Biophys. Res. Commun.* **2025**, *763*, 151741. [[CrossRef](#)] [[PubMed](#)]
96. Wang, X.; Smith, C.; Yin, H. Targeting Toll-like receptors with small molecule agents. *Chem. Soc. Rev.* **2013**, *42*, 4859–4866. [[CrossRef](#)]
97. Takashima, K.; Matsunaga, N.; Yoshimatsu, M.; Hazeki, K.; Kaisho, T.; Uekata, M.; Hazeki, O.; Akira, S.; Iizawa, Y.; Ii, M. Analysis of binding site for the novel small-molecule TLR4 signal transduction inhibitor TAK-242 and its therapeutic effect on mouse sepsis model. *Br. J. Pharmacol.* **2009**, *157*, 1250–1262. [[CrossRef](#)]
98. Mio, T.; Romberger, D.J.; Thompson, A.B.; Robbins, R.A.; Heires, A.; Rennard, S.I. Cigarette smoke induces interleukin-8 release from human bronchial epithelial cells. *Am. J. Respir. Crit. Care Med.* **1997**, *155*, 1770–1776. [[CrossRef](#)] [[PubMed](#)]
99. Levänen, B.; Glader, P.; Dahlén, B.; Billing, B.; Qvarfordt, I.; Palmberg, L.; Larsson, K.; Lindén, A. Impact of tobacco smoking on cytokine signaling via interleukin-17A in the peripheral airways. *Int. Peer-Rev. J. Ther. Pharmacol.* **2016**, *11*, 2109–2116. [[CrossRef](#)]
100. Guo, J.H.; Thuong, L.H.H.; Jiang, Y.J.; Huang, C.L.; Huang, Y.W.; Cheng, F.J.; Liu, P.I.; Liu, C.L.; Huang, W.C.; Tang, C.H. Cigarette smoke promotes IL-6-dependent lung cancer migration and osteolytic bone metastasis. *Int. J. Biol. Sci.* **2024**, *20*, 3257–3268. [[CrossRef](#)]
101. Reynolds, P.R.; Cosio, M.G.; Hoidal, J.R. Cigarette smoke-induced Egr-1 upregulates proinflammatory cytokines in pulmonary epithelial cells. *Am. J. Respir. Cell Mol. Biol.* **2006**, *35*, 314–319. [[CrossRef](#)]
102. Lee, K.H.; Lee, C.H.; Woo, J.; Jeong, J.; Jang, A.H.; Yoo, C.G. Cigarette Smoke Extract Enhances IL-17A-Induced IL-8 Production via Up-Regulation of IL-17R in Human Bronchial Epithelial Cells. *Mol Cells* **2018**, *41*, 282–289.
103. Yang, S.R.; Chida, A.S.; Bauter, M.R.; Shafiq, N.; Seweryniak, K.; Maggirwar, S.B.; Kilty, I.; Rahman, I. Cigarette smoke induces proinflammatory cytokine release by activation of NF- $\kappa$ B and posttranslational modifications of histone deacetylase in macrophages. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2006**, *291*, L46–L57. [[CrossRef](#)]
104. Oltmanns, U.; Chung, K.F.; Walters, M.; John, M.; Mitchell, J.A. Cigarette smoke induces IL-8, but inhibits eotaxin and RANTES release from airway smooth muscle. *Respir. Res.* **2005**, *6*, 74. [[CrossRef](#)]
105. Ramage, L.; Jones, A.C.; Whelan, C.J. Induction of apoptosis with tobacco smoke and related products in A549 lung epithelial cells in vitro. *J. Inflamm.* **2006**, *3*, 3. [[CrossRef](#)]
106. Jiao, Z.X.; Ao, Q.L.; Xiong, M. Cigarette smoke extract inhibits the proliferation of alveolar epithelial cells and induces apoptosis. *Sheng Li Xue Bao* **2006**, *58*, 244–254.
107. Wang, J.; Wilcken, D.E.; Wang, X.L. Cigarette smoke activates caspase-3 to induce apoptosis of human umbilical venous endothelial cells. *Mol. Genet. Metab.* **2001**, *72*, 82–88. [[CrossRef](#)]
108. Messner, B.; Frotschnig, S.; Steinacher-Nigisch, A.; Winter, B.; Eichmair, E.; Gebetsberger, J.; Schwaiger, S.; Ploner, C.; Laufer, G.; Bernhard, D. Apoptosis and necrosis: Two different outcomes of cigarette smoke condensate-induced endothelial cell death. *Cell Death Dis.* **2012**, *3*, e424. [[CrossRef](#)]
109. Feng, H.; Li, M.; Altawil, A.; Yin, Y.; Zheng, R.; Kang, J. Cigarette smoke extracts induce apoptosis in Raw264.7 cells via endoplasmic reticulum stress and the intracellular Ca<sup>2+</sup>/P38/STAT1 pathway. *Toxicol. In Vitro* **2021**, *77*, 105249. [[CrossRef](#)]
110. Banerjee, S.; Maity, P.; Mukherjee, S.; Sil, A.K.; Panda, K.; Chattopadhyay, D.; Chatterjee, I.B. Black tea prevents cigarette smoke-induced apoptosis and lung damage. *J. Inflamm.* **2007**, *4*, 3. [[CrossRef](#)] [[PubMed](#)]
111. Lin, X.-X.; Yang, X.-F.; Jiang, J.-X.; Zhang, S.-J.; Guan, Y.; Liu, Y.-N.; Sun, Y.-H.; Xie, Q.-M. Cigarette smoke extract-induced BEAS-2B cell apoptosis and anti-oxidative Nrf-2 up-regulation are mediated by ROS-stimulated p38 activation. *Toxicol. Mech. Methods* **2014**, *24*, 575–583. [[CrossRef](#)]

112. Seo, Y.S.; Park, J.M.; Kim, J.H.; Lee, M.Y. Cigarette Smoke-Induced Reactive Oxygen Species Formation: A Concise Review. *Antioxidants* **2023**, *12*, 1732. [[CrossRef](#)]
113. Lyons, M.J.; Gibson, J.F.; Ingram, D.J. Free-radicals produced in cigarette smoke. *Nature* **1958**, *181*, 1003–1004. [[CrossRef](#)] [[PubMed](#)]
114. Shein, M.; Jeschke, G. Comparison of free radical levels in the aerosol from conventional cigarettes, electronic cigarettes, and heat-not-burn tobacco products. *Chem. Res. Toxicol.* **2019**, *32*, 1289–1298. [[CrossRef](#)]
115. Bartalis, J.; Chan, W.G.; Wooten, J.B. A new look at radicals in cigarette smoke. *Anal. Chem.* **2007**, *79*, 5103–5106. [[CrossRef](#)]
116. Mitra, A.; Mandal, A.K. Conjugation of para-benzoquinone of cigarette smoke with human hemoglobin leads to unstable tetramer and reduced cooperative oxygen binding. *J. Am. Soc. Mass Spectrom.* **2018**, *29*, 2048–2058. [[CrossRef](#)]
117. Ghosh, A.; Choudhury, A.; Das, A.; Chatterjee, N.S.; Das, T.; Chowdhury, R.; Panda, K.; Banerjee, R.; Chatterjee, I.B. Cigarette smoke induces p-benzoquinone-albumin adduct in blood serum: Implications on structure and ligand binding properties. *Toxicology* **2012**, *292*, 78–89. [[CrossRef](#)]
118. Chang, K.H.; Park, J.M.; Lee, C.H.; Kim, B.; Choi, K.C.; Choi, S.J.; Lee, K.; Lee, M.Y. NADPH oxidase (NOX) 1 mediates cigarette smoke-induced superoxide generation in rat vascular smooth muscle cells. *Toxicol. In Vitro* **2017**, *38*, 49–58. [[CrossRef](#)]
119. Yildiz, L.; Kayaoglu, N.; Aksoy, H. The changes of superoxide dismutase, catalase and glutathione peroxidase activities in erythrocytes of active and passive smokers. *Clin. Chem. Lab. Med.* **2002**, *40*, 612–615. [[CrossRef](#)]
120. Kondo, T.; Tagami, S.; Yoshioka, A.; Nishimura, M.; Kawakami, Y. Current smoking of elderly men reduces antioxidants in alveolar macrophages. *Am. J. Respir. Crit. Care Med.* **1994**, *149*, 178–182. [[CrossRef](#)]
121. Oriola, A.O.; Oyediji, A.O. Plant-Derived Natural Products as Lead Agents against Common Respiratory Diseases. *Molecules* **2022**, *27*, 3054. [[CrossRef](#)]
122. Li, D.; Hu, J.; Wang, T.; Zhang, X.; Liu, L.; Wang, H.; Wu, Y.; Xu, D.; Wen, F. Silymarin attenuates cigarette smoke extract-induced inflammation via simultaneous inhibition of autophagy and ERK/p38 MAPK pathway in human bronchial epithelial cells. *Sci. Rep.* **2016**, *6*, 37751. [[CrossRef](#)] [[PubMed](#)]
123. Li, D.; Xu, D.; Wang, T.; Shen, Y.; Guo, S.; Zhang, X.; Guo, L.; Li, X.; Liu, L.; Wen, F. Silymarin attenuates airway inflammation induced by cigarette smoke in mice. *Inflammation* **2015**, *38*, 871–878. [[CrossRef](#)]
124. Hoch, C.C.; Petry, J.; Griesbaum, L.; Weiser, T.; Werner, K.; Ploch, M.; Verschoor, A.; Multhoff, G.; Bashiri Dezfouli, A.; Wollenberg, B. 1,8-cineole (eucalyptol): A versatile phytochemical with therapeutic applications across multiple diseases. *Biomed. Pharmacother.* **2023**, *167*, 115467. [[CrossRef](#)]
125. Seol, G.H.; Kim, K.Y. Eucalyptol and Its Role in Chronic Diseases. *Adv. Exp. Med. Biol.* **2016**, *929*, 389–398.
126. Reis, R.; Orak, D.; Yilmaz, D.; Cimen, H.; Sipahi, H. Modulation of cigarette smoke extract-induced human bronchial epithelial damage by eucalyptol and curcumin. *Hum. Exp. Toxicol.* **2021**, *40*, 1445–1462. [[CrossRef](#)]
127. Yu, N.; Sun, Y.T.; Su, X.M.; He, M.; Dai, B.; Kang, J. Treatment with eucalyptol mitigates cigarette smoke-induced lung injury through suppressing ICAM-1 gene expression. *Biosci. Rep.* **2018**, *38*, BSR20171636. [[CrossRef](#)]
128. Kennedy-Feitosa, E.; Cattani-Cavaliere, I.; Barroso, M.V.; Romana-Souza, B.; Brito-Gitirana, L.; Valenca, S.S. Eucalyptol promotes lung repair in mice following cigarette smoke-induced emphysema. *Phytomedicine* **2019**, *55*, 70–79. [[CrossRef](#)]
129. Hewlings, S.J.; Kalman, D.S. Curcumin: A Review of Its Effects on Human Health. *Foods* **2017**, *6*, 92. [[CrossRef](#)]
130. Nelson, K.M.; Dahlin, J.L.; Bisson, J.; Graham, J.; Pauli, G.F.; Walters, M.A. The Essential Medicinal Chemistry of Curcumin. *J. Med. Chem.* **2017**, *60*, 1620–1637. [[CrossRef](#)]
131. Fanoudi, S.; Alavi, M.S.; Mehri, S.; Hosseinzadeh, H. The protective effects of curcumin against cigarette smoke-induced toxicity: A comprehensive review. *Phytother. Res.* **2024**, *38*, 98–116. [[CrossRef](#)]
132. Kokkinis, S.; De Rubis, G.; Paudel, K.R.; Patel, V.K.; Yeung, S.; Jessamine, V.; MacLoughlin, R.; Hansbro, P.M.; Oliver, B.; Dua, K. Liposomal curcumin inhibits cigarette smoke induced senescence and inflammation in human bronchial epithelial cells. *Pathol. Res. Pract.* **2024**, *260*, 155423. [[CrossRef](#)]
133. Patel, V.K.; Kokkinis, S.; De Rubis, G.; Hansbro, P.M.; Paudel, K.R.; Dua, K. Curcumin liposomes attenuate the expression of cigarette smoke extract-induced inflammatory markers IL-8 and IL-24 in vitro. *EXCLI J.* **2024**, *23*, 904–907.
134. Li, Q.; Sun, J.; Mohammadtursun, N.; Wu, J.; Dong, J.; Li, L. Curcumin inhibits cigarette smoke-induced inflammation via modulating the PPAR $\gamma$ -NF- $\kappa$ B signaling pathway. *Food Funct.* **2019**, *10*, 7983–7994. [[CrossRef](#)]
135. Ames, T.R.; Beton, J.L.; Bowers, A.; Halsall, T.G.; Jones, E. The chemistry of the triterpenes and related compounds Part XXIII the structure of taraxasterol  $\psi$ -taraxasterol (heterolupeol) and lupenol-I. *J. Chem. Soc.* **1954**, *25*, 307–318. [[CrossRef](#)]
136. Jiao, F.; Tan, Z.; Yu, Z.; Zhou, B.; Meng, L.; Shi, X. The phytochemical and pharmacological profile of taraxasterol. *Front. Pharmacol.* **2022**, *13*, 927365. [[CrossRef](#)] [[PubMed](#)]
137. Liu, X.; Yu, D.; Wang, T. Taraxasterol inhibits cigarette smoke-induced lung inflammation by inhibiting reactive oxygen species-induced TLR4 trafficking to lipid rafts. *Eur. J. Pharmacol.* **2016**, *789*, 301–307.
138. Yagishita, Y.; Fahey, J.W.; Dinkova-Kostova, A.T.; Kensler, T.W. Broccoli or Sulforaphane: Is It the Source or Dose That Matters? *Molecules* **2019**, *24*, 3593. [[CrossRef](#)]

139. Baralić, K.; Živanović, J.; Marić, Đ.; Bozic, D.; Grahovac, L.; Antonijević Miljaković, E.; Ćurčić, M.; Buha Djordjevic, A.; Bulat, Z.; Antonijević, B.; et al. Sulforaphane-A Compound with Potential Health Benefits for Disease Prevention and Treatment: Insights from Pharmacological and Toxicological Experimental Studies. *Antioxidants* **2024**, *13*, 147. [[CrossRef](#)]
140. Song, H.; Wang, Y.H.; Zhou, H.Y.; Cui, K.M. Sulforaphane alleviates LPS-induced inflammatory injury in ARPE-19 cells by repressing the PWRN2/NF-κB pathway. *Immunopharmacol. Immunotoxicol.* **2022**, *44*, 868–876. [[CrossRef](#)]
141. Gasparello, J.; Marzaro, G.; Papi, C.; Gentili, V.; Rizzo, R.; Zurlo, M.; Scapoli, C.; Finotti, A.; Gambari, R. Effects of Sulforaphane on SARS-CoV-2 infection and NF-kappaB dependent expression of genes involved in the COVID-19 'cytokine storm'. *Int. J. Mol. Med.* **2023**, *52*, 76. [[CrossRef](#)]
142. Starrett, W.; Blake, D.J. Sulforaphane inhibits de novo synthesis of IL-8 and MCP-1 in human epithelial cells generated by cigarette smoke extract. *J. Immunotoxicol.* **2011**, *8*, 150–158. [[CrossRef](#)]
143. Jiao, Z.; Chang, J.; Li, J.; Nie, D.; Cui, H.; Guo, D. Sulforaphane increases Nrf2 expression and protects alveolar epithelial cells against injury caused by cigarette smoke extract. *Mol. Med. Rep.* **2017**, *16*, 1241–1247. [[CrossRef](#)]
144. Jiao, Z.; Zhang, Q.; Chang, J.; Nie, D.; Li, M.; Zhu, Y.; Wang, C.; Wang, Y.; Liu, F. A protective role of sulforaphane on alveolar epithelial cells exposed to cigarette smoke extract. *Exp. Lung Res.* **2013**, *39*, 379–386. [[CrossRef](#)] [[PubMed](#)]
145. Hau, D.K.; Gambari, R.; Wong, R.S.; Yuen, M.C.; Cheng, G.Y.; Tong, C.S.; Zhu, G.Y.; Leung, A.K.; Lai, P.B.; Lau, F.Y.; et al. Phyllanthus urinaria extract attenuates acetaminophen induced hepatotoxicity: Involvement of cytochrome P450 CYP2E1. *Phytomedicine* **2009**, *16*, 751–760. [[CrossRef](#)]
146. Sudjaroen, Y.; Hull, W.E.; Erben, G.; Würtele, G.; Changbumrung, S.; Ulrich, C.M.; Owen, R.W. Isolation and characterization of ellagitannins as the major polyphenolic components of Longan (*Dimocarpus longan* Lour) seeds. *Phytochemistry* **2012**, *77*, 226–237. [[CrossRef](#)]
147. Okabe, S.; Suganuma, M.; Imayoshi, Y.; Taniguchi, S.; Yoshida, T.; Fujiki, H. New TNF-α releasing inhibitors, geraniin and corilagin, in leaves of acer nikoense, megusurino-ki. *Biol. Pharm. Bull.* **2001**, *24*, 1145–1148. [[CrossRef](#)]
148. Zhao, L.; Zhang, S.L.; Tao, J.Y.; Pang, R.; Jin, F.; Guo, Y.J.; Dong, J.H.; Ye, P.; Zhao, H.Y.; Zheng, G.H. Preliminary exploration on anti-inflammatory mechanism of Corilagin (beta-1-O-galloyl-3,6-(R)-hexahydroxydiphenoyl-d-glucose) in vitro. *Int. Immunopharmacol.* **2008**, *8*, 1059–1064. [[CrossRef](#)] [[PubMed](#)]
149. Kinoshita, S.; Inoue, Y.; Nakama, S.; Ichiba, T.; Aniya, Y. Antioxidant and hepatoprotective actions of medicinal herb, Terminalia catappa L. from Okinawa Island and its tannin corilagin. *Phytomedicine* **2007**, *14*, 755–762. [[CrossRef](#)]
150. Luo, T.; Zhou, X.; Qin, M.; Lin, Y.; Lin, J.; Chen, G.; Liu, A.; Ouyang, D.; Chen, D.; Pan, H. Corilagin Restrains NLRP3 Inflammasome Activation and Pyroptosis through the ROS/TXNIP/NLRP3 Pathway to Prevent Inflammation. *Oxid. Med. Cell. Longev.* **2022**, *2022*, 1652244. [[CrossRef](#)]
151. Fan, G.J.; Liu, X.D.; Qian, Y.P.; Shang, Y.J.; Li, X.Z.; Dai, F.; Fang, J.G.; Jin, X.L.; Zhou, B. 4,4'-Dihydroxy-trans-stilbene, a resveratrol analogue, exhibited enhanced antioxidant activity and cytotoxicity. *Bioorg. Med. Chem.* **2009**, *17*, 2360–2365. [[CrossRef](#)]
152. Wang, T.; Dai, F.; Li, G.H.; Chen, X.M.; Li, Y.R.; Wang, S.Q.; Ren, D.M.; Wang, X.N.; Lou, H.X.; Zhou, B.; et al. Trans-4,4'-dihydroxystilbene ameliorates cigarette smoke-induced progression of chronic obstructive pulmonary disease via inhibiting oxidative stress and inflammatory response. *Free Radic. Biol. Med.* **2020**, *152*, 525–539. [[CrossRef](#)]
153. Al-Joufi, F.A.; Shaukat, S.; Hussain, L.; Khan, K.U.R.; Hussain, N.; Al Haddad, A.H.I.; Alqahtani, A.; Alqahtani, T.; Momenah, M.A.; Ibrahim, S.A.; et al. Lavandula stoechas significantly alleviates cigarette smoke-induced acute lung injury via modulation of oxidative stress and the NF-κB pathway. *Food Biosci.* **2024**, *59*, 103834. [[CrossRef](#)]
154. Hussain, N.; Ikram, N.; Khan, K.U.R.; Hussain, L.; Alqahtani, A.M.; Alqahtani, T.; Hussain, M.; Suliman, M.; Alshahrani, M.Y.; Sitohy, B. *Cichorium intybus* L. significantly alleviates cigarette smoke-induced acute lung injury by lowering NF-κB pathway activation and inflammatory mediators. *Heliyon* **2023**, *9*, e22055. [[CrossRef](#)]
155. Zeng, L.H.; Fatima, M.; Syed, S.K.; Shaukat, S.; Mahdy, A.; Hussain, N.; Al Haddad, A.H.I.; Said, A.S.A.; Alqahtani, A.; Alqahtani, T.; et al. Anti-inflammatory and anti-oxidant properties of Ipomoea nil (Linn.) Roth significantly alleviates cigarette smoke (CS)-induced acute lung injury via possibly inhibiting the NF-κB pathway. *Biomed. Pharmacother.* **2022**, *155*, 113267. [[CrossRef](#)]
156. Barroso, M.V.; Cattani-Cavaliere, I.; de Brito-Gitirana, L.; Fautrel, A.; Lagente, V.; Schmidt, M.; Porto, L.C.; Romana-Souza, B.; Valenca, S.S.; Lanzetti, M. Propolis reversed cigarette smoke-induced emphysema through macrophage alternative activation independent of Nrf2. *Bioorg. Med. Chem.* **2017**, *25*, 5557–5568. [[CrossRef](#)]
157. Lanzetti, M.; Lopes, A.A.; Ferreira, T.S.; de Moura, R.S.; Resende, A.C.; Porto, L.C.; Valenca, S.S. Mate tea ameliorates emphysema in cigarette smoke-exposed mice. *Exp. Lung Res.* **2011**, *37*, 246–257. [[CrossRef](#)]
158. Pires, K.M.; Valenca, S.S.; Resende, A.C.; Porto, L.C.; Queiroz, E.F.; Moreira, D.D.; de Moura, R.S. Grape skin extract reduced pulmonary oxidative response in mice exposed to cigarette smoke. *Med. Sci. Monit.* **2011**, *17*, BR187–BR195. [[CrossRef](#)]
159. Imai, J.; Ide, N.; Nagae, S.; Moriguchi, T.; Matsuura, H.; Itakura, Y. Antioxidant and radical scavenging effects of aged garlic extract and its constituents. *Planta Medica* **1994**, *60*, 417–420. [[CrossRef](#)] [[PubMed](#)]

160. Serrano, J.C.E.; Castro-Boqué, E.; García-Carrasco, A.; Morán-Valero, M.I.; González-Hedström, D.; Bermúdez-López, M.; Valdivielso, J.M.; Espinel, A.E.; Portero-Otín, M. Antihypertensive Effects of an Optimized Aged Garlic Extract in Subjects with Grade I Hypertension and Antihypertensive Drug Therapy: A Randomized, Triple-Blind Controlled Trial. *Nutrients* **2023**, *15*, 3691. [[CrossRef](#)]
161. Ohkubo, S.; Dalla Via, L.; Grancara, S.; Kanamori, Y.; García-Argáez, A.N.; Canettieri, G.; Arcari, P.; Toninello, A.; Agostinelli, E. The antioxidant, aged garlic extract, exerts cytotoxic effects on wild-type and multidrug-resistant human cancer cells by altering mitochondrial permeability. *Int. J. Oncol.* **2018**, *53*, 1257–1268. [[CrossRef](#)] [[PubMed](#)]
162. Liu, X.; Wang, N.; He, Z.; Chen, C.; Ma, J.; Liu, X.; Deng, S.; Xie, L. Diallyl trisulfide inhibits osteosarcoma 143B cell migration, invasion and EMT by inducing autophagy. *Heliyon* **2024**, *10*, e26681. [[CrossRef](#)] [[PubMed](#)]
163. Ferguson, D.T.; Taka, E.; Messeha, S.; Flores-Rozas, H.; Reed, S.L.; Redmond, B.V.; Soliman, K.F.A.; Kanga, K.J.W.; Darling-Reed, S.F. The Garlic Compound, Diallyl Trisulfide, Attenuates Benzo[a]Pyrene-Induced Precancerous Effect through Its Antioxidant Effect, AhR Inhibition, and Increased DNA Repair in Human Breast Epithelial Cells. *Nutrients* **2024**, *16*, 300. [[CrossRef](#)]
164. Bentke-Imiolek, A.; Szlęzak, D.; Zarzycka, M.; Wróbel, M.; Bronowicka-Adamska, P. S-Allyl-L-Cysteine Affects Cell Proliferation and Expression of H2S-Synthetizing Enzymes in MCF-7 and MDA-MB-231 Adenocarcinoma Cell Lines. *Biomolecules* **2024**, *14*, 188. [[CrossRef](#)]
165. Kanamori, Y.; Via, L.D.; Maccone, A.; Canettieri, G.; Greco, A.; Toninello, A.; Agostinelli, E. Aged garlic extract and its constituent, S-allyl-L-cysteine, induce the apoptosis of neuroblastoma cancer cells due to mitochondrial membrane depolarization. *Exp. Ther. Med.* **2020**, *19*, 1511–1521. [[CrossRef](#)]
166. Kodera, Y.; Kurita, M.; Nakamoto, M.; Matsutomo, T. Chemistry of aged garlic: Diversity of constituents in aged garlic extract and their production mechanisms via the combination of chemical and enzymatic reactions. *Exp. Ther. Med.* **2020**, *19*, 1574–1584. [[CrossRef](#)]
167. Borek, C. Antioxidant health effects of aged garlic extract. *J. Nutr.* **2001**, *131*, 1010S–1015S. [[CrossRef](#)] [[PubMed](#)]
168. Ryu, K.; Rosen, R.T. Unique Chemistry of Aged Garlic Extract. *Orient. Foods Herbs* **2003**, *19*, 258–270.
169. El-Saadony, M.T.; Saad, A.M.; Korma, S.A.; Salem, H.M.; Abd El-Mageed, T.A.; Alkafaas, S.S.; Elsalahaty, M.I.; Elkafas, S.S.; Mosa, W.F.A.; Ahmed, A.E.; et al. Garlic bioactive substances and their therapeutic applications for improving human health: A comprehensive review. *Front. Immunol.* **2024**, *15*, 1277074. [[CrossRef](#)] [[PubMed](#)]
170. Nagae, S.; Ushijima, M.; Hatono, S.; Imai, J.; Kasuga, S.; Matsuura, H.; Itakura, Y.; Higashi, Y. Pharmacokinetics of the garlic compound S-allylcysteine. *Planta Med.* **1994**, *60*, 214–217. [[CrossRef](#)] [[PubMed](#)]
171. Agostinelli, E.; Marzaro, G.; Gambari, R.; Finotti, A. Potential applications of components of Aged Garlic Extract (AGE) in mitigating pro-inflammatory gene expression linked to human diseases. *Exp. Ther. Med.* **2025**, *30*, 134. [[CrossRef](#)]
172. Gasparello, J.; Papi, C.; Marzaro, G.; Maccone, A.; Zurlo, M.; Finotti, A.; Agostinelli, E.; Gambari, R. Aged Garlic Extract (AGE) and Its Constituent S-Allyl-Cysteine (SAC) Inhibit the Expression of Pro-Inflammatory Genes Induced in Bronchial Epithelial IB3-1 Cells by Exposure to the SARS-CoV-2 Spike Protein and the BNT162b2 Vaccine. *Molecules* **2024**, *29*, 5938. [[CrossRef](#)]
173. Papi, C.; Gasparello, J.; Marzaro, G.; Maccone, A.; Zurlo, M.; Di Padua, F.; Fino, P.; Agostinelli, E.; Gambari, R.; Finotti, A.; et al. Aged garlic extract major constituent S-1-propenyl-l-cysteine inhibits proinflammatory mRNA expression in bronchial epithelial IB3-1 cells exposed to the BNT162b2 vaccine. *Exp. Ther. Med.* **2025**, *30*, 153. [[CrossRef](#)]
174. Elmazoglu, Z.; Aydın Bek, Z.; Sarıbaş, S.G.; Özoğul, C.; Goker, B.; Bitik, B.; Aktekin, C.N.; Karasu, Ç. S-allylcysteine inhibits chondrocyte inflammation to reduce human osteoarthritis via targeting RAGE, TLR4, JNK, and Nrf2 signaling: Comparison with colchicine. *Biochem. Cell Biol.* **2021**, *99*, 645–654. [[CrossRef](#)]
175. Geng, Z.; Rong, Y.; Lau, B.H. S-allyl cysteine inhibits activation of nuclear factor kappa B in human T cells. *Free Radic. Biol. Med.* **1997**, *23*, 345–350. [[CrossRef](#)]
176. Huang, X.P.; Shi, Z.H.; Ming, G.F.; Xu, D.M.; Cheng, S.Q. S-Allyl-L-cysteine (SAC) inhibits copper-induced apoptosis and cuproptosis to alleviate cardiomyocyte injury. *Biochem. Biophys. Res. Commun.* **2024**, *730*, 150341. [[CrossRef](#)]
177. Chen, P.; Hu, M.; Liu, F.; Yu, H.; Chen, C. S-allyl-l-cysteine (SAC) protects hepatocytes from alcohol-induced apoptosis. *FEBS Open Bio* **2019**, *9*, 1327–1336. [[CrossRef](#)] [[PubMed](#)]
178. Kalayarsan, S.; Sriram, N.; Sureshkumar, A.; Sudhandiran, G. Chromium (VI)-induced oxidative stress and apoptosis is reduced by garlic and its derivative S-allylcysteine through the activation of Nrf2 in the hepatocytes of Wistar rats. *J. Appl. Toxicol.* **2008**, *28*, 908–919. [[CrossRef](#)]
179. Orozco-Ibarra, M.; Muñoz-Sánchez, J.; Zavala-Medina, M.E.; Pineda, B.; Magaña-Maldonado, R.; Vázquez-Contreras, E.; Maldonado, P.D.; Pedraza-Chaverri, J.; Chánez-Cárdenas, M.E. Aged garlic extract and S-allylcysteine prevent apoptotic cell death in a chemical hypoxia model. *Biol Res.* **2016**, *49*, 7. [[CrossRef](#)]
180. Reddy, V.P. Oxidative Stress in Health and Disease. *Biomedicine* **2023**, *11*, 2925. [[CrossRef](#)]
181. Liu, Z.; Ren, Z.; Zhang, J.; Chuang, C.C.; Kandaswamy, E.; Zhou, T.; Zuo, L. Role of ROS and Nutritional Antioxidants in Human Diseases. *Front. Physiol.* **2018**, *9*, 477. [[CrossRef](#)] [[PubMed](#)]

182. Gupta, P.; Dutt, V.; Kaur, N.; Kalra, P.; Gupta, S.; Dua, A.; Dabur, R.; Saini, V.; Mittal, A. S-allyl cysteine: A potential compound against skeletal muscle atrophy. *Biochim. Biophys. Acta Gen. Subj.* **2020**, *1864*, 129676. [[CrossRef](#)]
183. He, Y.; Xiao, L.; Zhang, J.; Zhu, Y.; Guo, Y.; Xia, Y.; Zhao, H.; Wei, Z.; Dai, Y. Diallyl trisulfide alleviates dextran sulphate sodium-induced colitis in mice by inhibiting NLRP3 inflammasome activation via ROS/Trx-1 pathway. *Basic Clin. Pharmacol. Toxicol.* **2024**, *135*, 593–606. [[CrossRef](#)] [[PubMed](#)]
184. Wang, Y.; Wang, H.L.; Xing, G.D.; Qian, Y.; Zhong, J.F.; Chen, K.L. S-allyl cysteine ameliorates heat stress-induced oxidative stress by activating Nrf2/HO-1 signaling pathway in BMECs. *Toxicol. Appl. Pharmacol.* **2021**, *416*, 115469. [[CrossRef](#)]
185. Ruiz-Sánchez, E.; Pedraza-Chaverri, J.; Medina-Campos, O.N.; Maldonado, P.D.; Rojas, P. S-allyl Cysteine, a Garlic Compound, Produces an Antidepressant-Like Effect and Exhibits Antioxidant Properties in Mice. *Brain Sci.* **2020**, *10*, 592. [[CrossRef](#)]
186. Xu, C.; Mathews, A.E.; Rodrigues, C.; Eudy, B.J.; Rowe, C.A.; O'Donoghue, A.; Percival, S.S. Aged garlic extract supplementation modifies inflammation and immunity of adults with obesity: A randomized, double-blind, placebo-controlled clinical trial. *Clin. Nutr. ESPEN* **2018**, *24*, 148–155. [[CrossRef](#)]
187. Budoff, M.J.; Ahmadi, N.; Gul, K.M.; Liu, S.T.; Flores, F.R.; Tiano, J.; Takasu, J.; Miller, E.; Tsimikas, S. Aged garlic extract supplemented with B vitamins, folic acid and L-arginine retards the progression of subclinical atherosclerosis: A randomized clinical trial. *Prev. Med.* **2009**, *49*, 101–107. [[CrossRef](#)]
188. Wlosinska, M.; Nilsson, A.C.; Hlebowicz, J.; Fakhro, M.; Malmjö, M.; Lindstedt, S. Aged Garlic Extract Reduces IL-6: A Double-Blind Placebo-Controlled Trial in Females with a Low Risk of Cardiovascular Disease. *Evid. Based Complement. Altern. Med.* **2021**, *2021*, 6636875. [[CrossRef](#)]
189. Gambari, R.; Papi, C.; Gasparello, J.; Agostinelli, E.; Finotti, A. Preliminary results and a theoretical perspective of co-treatment using a miR-93-5p mimic and aged garlic extract to inhibit the expression of the pro-inflammatory interleukin-8 gene. *Exp. Ther. Med.* **2025**, *29*, 85. [[CrossRef](#)] [[PubMed](#)]
190. Onwuzo, C.N.; Olukorode, J.; Sange, W.; Orimoloye, D.A.; Udojike, C.; Omoragbon, L.; Hassan, A.E.; Falade, D.M.; Omiko, R.; Odunaike, O.S.; et al. A Review of Smoking Cessation Interventions: Efficacy, Strategies for Implementation, and Future Directions. *Cureus* **2024**, *16*, e52102. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.