

Medicinal Chemistry, Pharmacology and Potential Therapeutic Benefits of Cannabinoid CB₂ Receptor Agonists

Mojgan Aghazadeh Tabrizi,[†] Pier Giovanni Baraldi,^{*,†} Pier Andrea Borea,[‡] and
Katia Varani[‡]

[†]Department of Chemical and Pharmaceutical Sciences, University of Ferrara, Italy

[‡]Department of Medical Science, Pharmacology Section, University of Ferrara, Italy

*To whom correspondence should be addressed.

Phone: 0039-0532- 455921 Fax: 0039-0532- 455953

E-mail: baraldi@unife.it

CONTENTS

1. INTRODUCTION	4
2. MOLECULAR CHARACTERIZATION OF CB ₂ RECEPTORS	6
3. DISTRIBUTION OF CB ₂ RECEPTORS	8
4. INTRACELLULAR PATHWAYS OF CB ₂ RECEPTORS	11
5. THE ENDOCANNABINOID SYSTEM	14
6. PHARMACOLOGICAL ROLE OF CB ₂ RECEPTORS	16
6.1. Immune System and Inflammation	18
6.2. Central Nervous System	20
6.3. Cardiovascular System	23
6.4. Atherosclerosis	23
6.5. Gastrointestinal system and liver	24
6.6. Pulmonary system	25
6.7. Osteoporosis	26
6.8. Cancer	27
6.9. Chronic and Neuropathic Pain	30
7. CHEMISTRY OF CB ₂ AGONISTS	33
7.1. Pyridine, Pyridazine	34
7.2. Sulfamoyl Benzamide	38
7.3. Heterocyclic Ylidene	40
7.4. Oxadiazole, α -Amidosulfone	43
7.5. Imidazole	45
7.6. Benzoimidazole	47
7.7. Imidazopyridine	52
7.8. Purine	54

7.9. Quinolone, Naphthyridine	55
7.10. 4-Oxo-1,4-dihydropyridine	58
7.11. Decahydroquinoline amide	60
7.12. Carboline	62
7.13. Heteroarylpyridine, Heteroarylpyrimidine	64
7.14. Indol-3-ylcycloalkyl ketone	66
7.15. Oxazinoquinoline	68
7.16. 1,4-Diazepane	70
7.17. (<i>S</i>)Proline, (<i>S</i>)Piperidine	72
7.18. Pyrazole-3-carboxamide	74
7.19. 3-Carboxamido-5-aryl-isoxazole	74
7.20. 1,3,5-Triazine	75
7.21. Analogs of HU-308	77
8. PATENTS	79
9. POTENTIAL THERAPEUTIC APPLICATIONS OF CB ₂ AGONISTS	84
10. FUTURE PERSPECTIVES AND CONCLUSIONS	92
Author Information	94
Corresponding Author	94
Notes	94
Biographies	94
References	96

1. INTRODUCTION

The endocannabinoid system is represented by a group of neuromodulatory lipids and their receptors that are involved in a variety of physiological and pathological conditions. The main endocannabinoids, or endogenous cannabis-like substances, are small molecules derived from arachidonic acid, N-arachidonoyl ethanolamine (anandamide, AEA) and 2-arachidonoyl-glycerol (2-AG).¹ Each endocannabinoid belongs to a larger class of lipids termed N-acyl-ethanolamines (NAEs) and monoacylglycerols (MAGs), respectively, where the members differ in the length and unsaturation degree of their acyl chains.² In particular, AEA and 2-AG represent polyunsaturated derivatives with potential biological activities as, for example, in the nervous system where they act as retrograde messengers.³ These endocannabinoids are not stored in vesicles and mediate intercellular signals from postsynaptic neurons back to presynaptic terminals where they inhibit the neurotransmitter release.⁴ Their levels are controlled by selective deactivating enzymes such as fatty acid amide hydrolase (FAAH) for AEA, and monoacylglycerol lipase (MGL) for 2-AG, that primarily influence the endocannabinoid tone.⁵

Two different types of G protein-coupled receptors (GPCRs) have been discovered and named as central cannabinoid (CB₁) and peripheral cannabinoid (CB₂) receptors. The CB₁ and CB₂ receptors are negatively coupled to adenylyl cyclase, exert an inhibitory effect on cyclic AMP (cAMP) production, and modulate the mitogen-activated protein kinase (MAPK) pathway.⁶ In addition, CB₁ receptors inhibit presynaptic N- and P/Q-type calcium channels, activate inwardly potassium channels, reduce neuronal excitability and neurotransmitter release.⁴ CB₁ receptors are primarily distributed in different areas of the brain related to motor control, in sites associated with pain processing, cognition, emotional responses, motivated behaviour and homeostasis.⁷

CB₂ receptors were originally believed to be present in periphery, primarily to immune cells, but nowadays it has been reported that these receptors are localized neuronally in different species. They are expressed in all hematopoietic cells such as lymphocytes, natural killer cells, macrophages

and neutrophils, and represent an attractive target for the treatment of the inflammatory status.⁸ These receptors also mediate a significant protection in brain microglia against neurotoxicity by modulating the release of anti- or pro-inflammatory cytokines.⁹

CB receptors can also be directly modulated by exogenous ligands, for example, by the main psychotropic constituent found in *cannabis sativa*, named Δ^9 -tetrahydrocannabinol (Δ^9 -THC). It is well reported that nearly 400 distinct chemical entities have been isolated from this plant, and that it contains approximately 60 separate centrally active or psychoactive components.⁵ Experimental evidence has reported the role of CB receptors in the pathophysiology, and novel developments in the medicinal chemistry of these receptor subtypes have tried to identify potential therapeutic areas for drug research.¹⁰ In vitro and in vivo studies of the specific effects of selective CB ligands have determined important progress on the role of CB receptors in various pathologies. Interestingly, selective agonists, antagonists, inverse agonists and novel allosteric modulators interacting with CB₁ or CB₂ receptors have been developed to inhibit or augment their basal tone.¹¹

Over the past decades, the discovery of CB receptors has been followed by intensive research related to identifying key proteins involved in their modulation and to investigating their interaction with endocannabinoid ligands.¹² In addition, the search related to novel CB₂ agonists has been greatly developed and a large number of ligands has been synthesized and tested with the aim to identify novel therapeutic drugs.¹³

From the pharmacological point of view, CB₂ agonists show different ways of interacting with the receptor site, suggesting various binding and functional properties well represented by pharmacodynamic concepts of full, partial and inverse agonism.¹⁴⁻¹⁶ In recent years, pharmaceutical companies and academic research laboratories have attempted to identify novel CB₂ agonists without CB₁ central effects by focusing on the discovery of high affinity, selective agonists for CB₂ receptors with a therapeutic potential in the treatment of different pathologies such as

neurodegenerative diseases, pain transduction and perception, ischemic stroke, severe inflammation, autoimmune diseases, osteoporosis and cancers.^{9,11}

The purpose of this review is to analyze the molecular characterization, distribution and signal transduction pathways of CB₂ receptors together with their pharmacological role in some of the most relevant biological systems. The structure-activity relationships of several interesting CB₂ agonists, including synthetic schemes in order to clarify the chemistry involved in this field, are reported. Moreover, the corresponding pharmacological and beneficial potentials of selected CB₂ compounds have been focused as well as the patent literature and the identification of promising therapeutic candidates.

2. MOLECULAR CHARACTERIZATION OF CB₂ RECEPTORS

The distinction between CB₁ and CB₂ receptors is primarily based on differences in their predicted amino acid sequence, signaling mechanisms, and tissue distribution. Cloning of the neuronal CB receptor initiated several years ago with a molecular approach to the study of cannabinoid actions.¹⁷ Since then, rapid progress has been made, which includes the cloning of a second CB receptor from cells of immune origin, the identification of two endogenous CB ligands and specific elucidation of their metabolic pathways.^{18,19} Briefly, CB₁ receptors have been cloned in different vertebrates including mammals, amphibians, birds and fish, exhibiting 97% to 99% amino acid sequence identity across species.²⁰ CB₁ mRNA and protein are primarily found in brain and neuronal tissues encoding protein residues of 473 and 472 amino acids in rat and human, respectively. The human and rat CB₁ receptors differ in only 13 residues, and the mouse CB₁ protein displays 99% amino acids identity compared to rats.²¹

The rat CB₂ receptors consist of 410 amino acids among transmembrane domain, and exhibit 48% homology with CB₁ receptors, less than the 70-80% usually seen between different types of GPCRs.²² The mouse CB₂ receptor has been cloned and reveals a high degree of homology with rat or human CB₂ receptor except in the carboxy terminus where rat CB₂ is 50 and 63 residues longer

than hCB₂ and mCB₂, respectively.²³ The importance of the carboxy terminus in regulating CB₂ receptor desensitization and internalization is well established and the differences identified in this specific region between species suggest caution when extrapolating experimental results from non human models to humans.²⁴ The differences between CB₁ and CB₂ receptors have been associated to their sequences with a high concentration in N-terminal and other extracellular regions.²⁵ Amino acids amenable to potential post-translational modifications include some serines/threonine residues that are phosphoacceptor sites and specific sites for asparagines N-linked glycosylation.²² These mutations generally give the same results in the ligand-receptor interaction even if they could play a key role in transmitting the binding signal to the G proteins.²⁶

A novel human CB_{2A} isoform has been discovered by sequencing RT-PCR fragments amplified from human brain cDNA that differentially encodes tissue specific CB₂ gene expression.²⁷ The CB_{2A} isoform is predominantly expressed in human testis, and the promoter of CB_{2A} is located upstream of CB_{2B} isoform promoter, that is primarily present in spleen.²⁸ The quantitative RT-PCR revealed that CB_{2A} mRNA is expressed in different human brain regions with similar levels found in muscle, spleen, intestine, leukocytes and kidney.²⁹ In contrast, CB_{2B} mRNA is mostly present in spleen and to a lesser extent in leukocytes, muscle, intestine, liver and heart.²⁷ The human CB_{2A} and CB_{2B} isoforms contain different 5' untranslated regions and are alternatively spliced to the major protein coding exon of the CB₂ gene.³⁰ The discrepancies on CB₂ mRNA sizes in the literature indicate incomplete structure of CB₂ gene in different species, or polymorphism in the same species. Moreover, species comparison has revealed that the CB₂ gene of human, rat and mouse genomes deviate in their gene structures and isoform expression patterns. The presence of tissue specific isoforms of CB₂ gene may provide tissue specific targeting of CBs in inflammatory and neuropathic pain disorders associated with CB₂ receptors in the brain and peripheral tissues.³¹

There is a large body of experimental evidence that is compatible with the presence of heterodimers and/or homodimers of the major subclasses of GPCRs with different functional

properties. In addition, allosteric interactions associated to the receptors, or across the receptor-receptor interface, could modulate the binding properties and thereby change their pharmacology.³² It has been reported that CB₁ receptors can exist as homodimers or heterodimers with one or more other classes of GPCRs, such as dopamine D₂, opioid and orexin receptors. Resulting cross talk between CB₁ and other receptors may involve the sequestration of Gai/o subunit mediating an alteration in the cellular response.^{33,34} The allosteric modulation found for CB₁ receptors suggests a new approach to the manipulation of the endocannabinoid system for therapeutic benefits based on the pharmacological effects of direct full or partial agonists and/or competitive antagonists.^{35,36} At present, no pharmacological or chemical details are present in the literature regarding homodimers, heterodimers or the allosteric modulation of the CB₂ receptors, even if, given the co-localization of CB receptors and of various GPCRs, the existence of physical interactions between these receptor subtypes that may affect CB pharmacology could be hypothesized.

3. DISTRIBUTION OF CB₂ RECEPTORS

The discovery of CB receptors was followed by the observation that CBs affect every organ system, suggesting that CB expression is present throughout the body.⁶ Further evidence has established that some of the most relevant molecular and functional responses in the central nervous system (CNS) are primarily associated to CB₁ receptors, and those in the periphery to CB₂ receptors. Briefly, CB₁ receptors are found in high concentrations within CNS and their concentration is considerably less in peripheral tissues.^{10,11}

It has been well established that CB₂ receptors are present in immune tissue, spleen, thymus, tonsils, bone marrow, pancreas, lung, splenic macrophage/monocyte preparations, mast cells and peripheral blood leukocytes (Table 1).

Table 1- Distribution and known functions in different cells or tissues of the CB₂ receptors

Cell type/ Tissue	Species	Functions
Macrophages ^a	Human	Decrease release of NO, IL-12, IL-23, TNF- α ; increase IL-10 secretion; reduce ROS production and CCL2 (chemokine C-C motif ligand 2)
B and T lymphocytes ^b	Human	Affect B and T cells differentiation TNF and the balance of pro-inflammatory cytokines; immunosuppressive effect
Stomach (enteric nerves) ^c	Human	Inhibit cholinergic and noradrenergic transmission
Liver ^d	Human	Inhibit fibrosis, reduce inflammation and injury
Myocardium ^e	Human	Protect in ischemia/reperfusion injury
Endothelial cells ^f	Human	Slow the progression of atherosclerosis
Intimal macrophages ^g	Human	Inhibit cytokine production and leukocyte migration; reduce macrophage activation and increase apoptosis
Lung and airways ^h	Human	Antitussive effect and prevent bronchoconstriction
Osteoclast ⁱ	Human	Decrease osteoclast precursor proliferation and osteoclast activity
Osteoblast ⁱ	Human	Increase osteoblast stimulation
Whole brain ^k	Rat, mouse, human	Sleepiness, alterations in cognition and memory, increase analgesia; orexigenic effect, antiemetic effect, decrease intraocular pressure; decrease inflammation and cause immunosuppression
Brainstem ^l	Rat, mouse	Modulate immunoreactivity, inhibit emesis
Cerebellum ^m	Rat, mouse, human	Inhibit T cells and adenylate cyclase, neuroprotective effect, prevent neuron loss
Cortex ⁿ	Rat, mouse, human	Mediate anti-hyperalgesia
Hippocampus ^o	Rat, mouse, human	Reduce food consumption
Other brain regions ^p	Rat	Modulate neuron responses in nociceptive transmission
Peripherical neurons/ spinal cord ^q	Rat, mouse, guinea pig, human	Suppress nociceptive transmission
Microglia ^r	Human	Decrease IL- β and IL-6 levels and TNF- α release; p38MAPK and NADPH oxidase activation; interfere with CCR2 (chemokine C-C motif receptor 2) and iNOS expression; inhibit migration and inflammation
Glia ^s	Human	Enhance release of IL-4 and IL-10

^aChuchawankul, S.; Shima, M.; Buckley, N.E.; Hartmann C.B McCoy, K.L. Role of cannabinoid receptors in inhibiting macrophage costimulatory activity. *Int Immunopharmacol.* **2004**, 4, 265–278; Han, K.H.; Lim, S.; Ryu, J.; Lee, C.W.; Kim, Y.; Kang, J.H.; Kang, S.S.; Ahn, Y.K.; Park, C.S.; Kim, J.J. CB₁ and CB₂ cannabinoid receptors differentially regulate the production of reactive oxygen species by macrophages. *Cardiovasc Res.* **2009**, 84, 378–386; Correa F, Docagne F, Mestre L, Clemente D, Hernangomez M, Loria F, Guaza C. A role for CB₂ receptors in anandamide signalling pathways involved in the regulation of IL-12 and IL-23 in microglial cells. *Biochem Pharmacol.* **2009**, 77, 86–100

^bZiring, D.; Wei, B.; Velazquez, P.; Schrage, M.; Buckley, N.E.; Braun, J. Formation of B and T cell subsets require the cannabinoid receptor CB₂. *Immunogenetics.* **2006**, 58, 714–725; Galiegue, S.; Mary, S.; Marchand, J.; Dussosoy, D.; Carriere, D.; Carayon, P.; Bouaboula, M.; Shire, D.; Le Fur, G.; Casellas, P. Expression of central and peripheral cannabinoid receptors in human immune tissues and leukocyte subpopulations. *Eur. J. Biochem.* **1995**, 232, 54–61.

^cStorr, M.; Gaffal, E.; Saur, D.; Schusdziarra, V.; Allescher, H.D. Effect of cannabinoids on neural transmission in rat gastric fundus. *Can. J Physiol Pharmacol.* **2002**, 80, 67–76; Mule, F.; Amato, A.; Baldassano, S.; Serio, R. Involvement of CB₁ and CB₂ receptors in the modulation of cholinergic neurotransmission in mouse gastric preparations. *Pharmacol. Res.* **2007**, 56, 185–192.

^dJulien, B.; Grenard, P.; Teixeira-Clerc, F.; Van Nhieu, J.T.; Li, L.; Karsak, M.; Zimmer, A.; Mallat, A.; Lotersztajn, S. Antifibrogenic role of the cannabinoid receptor CB₂ in the liver. *Gastroenterology.* **2005**, 128, 742–755; Munoz-Luque, J.; Ros, J.; Fernandez-Varo, G.; Tugues, S.; Morales-Ruiz, M.; Alvarez, C. E.; Friedman, S. L.; Arroyo, V.; Jimenez, W. Regression of fibrosis after chronic stimulation of cannabinoid CB₂ receptor in cirrhotic rats. *J. Pharmacol. Exp. Ther.* **2008**, 324, 475–483;

- ^eLepicier, P.; Bibeau-Poirier, A.; Lagneux, C.; Servant, M.J.; Lamontagne, D. Signaling pathways involved in the cardioprotective effects of cannabinoids. *J. Pharmacol. Sci.* **2006**, *102*, 155-166; Shmist, Y.A.; Goncharov, I.; Eichler, M.; Shneyvays, V.; Isaac, A.; Vogel, Z.; Shainberg, A. Delta-9 tetrahydrocannabinol protects cardiac cells from hypoxia via CB₂ receptor activation and nitric oxide production. *Mol. Cell Biochem.* **2006**, *283*, 75-83.
- ^fRajesh, M.; Mukhopadhyay, P.; Batkai, S.; Hasko, G.; Liaudet, L.; Huffman, J.W.; Csiszar, A.; Ungvari, Z.I.; Mackie, K.; Chatterjee, S.; Pacher, P. Cannabinoid-2 receptor stimulation attenuates TNF α -induced human endothelial cell activation, transendothelial migration of monocytes, and monocyte-endothelial adhesion. *Am. J. Physiol. Heart Circ. Physiol.* **2007**, *293*(4), H2210-H2218; Steffens, S.; Veillard, N. R.; Arnaud, C.; Pelli, G.; Burger, F.; Staub, C.; Karsak, M.; Zimmer, A.; Frossard, J.L.; Mach, F. Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature.* **2005**, *434*, 782-786.
- ^gMontecucco, F.; Burger, F.; Mach, F.; Steffens, S. CB₂ cannabinoid receptor agonist JWH-015 modulates human monocyte migration through defined intracellular signaling pathways. *Am. J. Physiol Heart Circ. Physiol.* **2008**, *294*, H1145-H1155; Freeman-Anderson, N.E.; Pickle, T. G.; Netherland, C.D.; Bales, A.; Buckley, N.E.; Thewke, D.P. Cannabinoid CB₂ receptor deficiency reduces the susceptibility of macrophages to oxidized LDL/oxysterol-induced apoptosis. *J. Lipid Res.* **2008**, *49*, 2338-2346.
- ^hPatel, H.J.; Birrell, M.A.; Crispino, N.; Hele, D.J.; Venkatesan, P.; Barnes, P.J.; Yacoub, M.H.; Belvisi, M.G. Inhibition of guinea-pig and human sensory nerve activity and the cough reflex in guinea-pigs by cannabinoid CB₂ receptor activation. *Br. J. Pharmacol.* **2003**, *140*, 261-268; Cui, Y.Y.; D'Agostino, B.; Risse, P. A.; Marrocco, G.; Naline, E.; Zhang, Y.; Chen, H.Z.; Finance, O.; Rinaldi Carmona, M.; Rossi, F.; Advenier, C. Cannabinoid CB₂ receptor activation prevents bronchoconstriction and airway oedema in a model of gastro-oesophageal reflux. *Eur. J. Pharmacol.* **2007**, *573*, 206-213.
- ⁱOfek, O.; Karsak, M.; Leclerc, N.; Fogel, M.; Frenkel, B.; Wright, K.; Tam, J.; Attar-Namdar, M.; Kram, V.; Shohami, E.; et al. Peripheral cannabinoid receptor, CB₂, regulates bone mass. *Proc. Natl. Acad. Sci. U. S. A.* **2006**, *103*, 696-701.
- ^jHowlett, A.C. The cannabinoid receptors. *Prostaglandins Other Lipid Mediat.* **2002**, *68-69*, 619-631; Gong, J.P.; Onaivi, E.S.; Ishiguro, H.; Liu, Q.R.; Tagliaferro, P.A.; Brusco, A.; Uhl, G.R. Cannabinoid CB₂ receptors: immunohistochemical localization in rat brain. *Brain Res.* **2006**, *1071*, 10-23.
- ^kVan Sickle, M.D.; Duncan, M.; Kingsley, P.J.; Mouihate, A.; Urbani, P.; Mackie, K.; et al. Identification and functional characterization of brainstem cannabinoid CB₂ receptors. *Science.* **2005**, *310*, 329-332.
- ^lSkaper, S.D.; Burianni, A.; Dal Toso, R.; Petrelli, L.; Romanello, S.; Facci, L.; Leon A. The aliamide palmitoylethanolamide and cannabinoids, but not anandamide, are protective in a delayed postglutamate paradigm of excitotoxic death in cerebellar granule neurons. *Proc Natl Acad Sci U S A.* **1996**, *93*, 3984-3989.
- ^mBeltramo, M.; Bernardini, N.; Bertorelli, R.; Campanella, M.; Nicolussi, E.; Fredduzzi S.; et al. CB₂ receptor-mediated antihyperalgesia: possible direct involvement of neural mechanisms. *Eur J Neurosci.* **2006**, *23*, 1530-1538.
- ⁿOnaivi, E.S.; Ishiguro, H.; Gong, J.P.; Patel, S.; Meozzi, P.A.; Myers, L.; Perchuk, A.; Mora, Z.; Tagliaferro, P.A.; Gardner, E.; Brusco, A.; Akinshola, B.E.; Hope, B.; Lujilde, J.; Inada, T.; Iwasaki, S.; Macharia, D.; Teasenfitz, L.; Arinami, T.; Uhl, G.R. Brain neuronal CB₂ cannabinoid receptors in drug abuse and depression: from mice to human subjects. *Plos One.* **2008**, *3*, 1640.
- ^oJhaveri, M.D.; Elmes, S.J.; Richardson, D.; Barrett, D.A.; Kendall, D.A.; Mason, R.; Chapman, V. Evidence for a novel functional role of cannabinoid CB₂ receptors in the thalamus of neuropathic rats. *Eur J Neurosci.* **2008**, *27*, 1722-1730.
- ^pSagar, D.R.; Kelly, S.; Millns, P.J.; O'Shaughnessey, C.T.; Kendall, D.A.; Chapman, V. Inhibitory effects of CB₁ and CB₂ receptor agonists on responses of DRG neurons and dorsal horn neurons in neuropathic rats. *Eur J Neurosci.* **2005**, *22*, 371-379.
- ^qPuffenbarger, R.A.; Boothe, A.C.; Cabral, G.A. Cannabinoids inhibit LPS-inducible cytokine mRNA expression in rat microglial cells. *Glia.* **2000**, *29*, 58-69; Ramirez, B.G.; Blazquez, C.; Gomez del Pulgar, T.; Guzman, M.; de Ceballos, M.L. Prevention of Alzheimer's disease pathology by cannabinoids: neuroprotection mediated by blockade of microglial activation. *J Neurosci.* **2005**, *25*, 1904-1913; Correa, F.; Docagne, F.; Mestre, L.; Clemente, D.; Hernangomez, M.; Loria, F.; Guaza, C. A role for CB₂ receptors in anandamide signalling pathways involved in the regulation of IL-12 and IL-23 in microglial cells. *Biochem Pharmacol.* **2009**, *77*, 86-100;
- ^rMolina-Holgado.; Molina-Holgado, E.; Guaza, C. The endogenous cannabinoid anandamide potentiates interleukin-6 production by astrocytes infected with Theiler's murine encephalomyelitis virus by a receptor-mediated pathway. *FEBS Lett.* **1998**, *433*, 139-142.
-

Moreover, they have been found in a variety of cultured immune cell models, including the myeloid cell line, U937, and granulocyte-like or macrophage-like cells.³⁷ The distribution pattern of CB₂ mRNA displays major variation in human blood cell populations, with a rank order of lymphocytes type B, natural killer cells, monocytes, neutrophils and lymphocytes.³⁸ CB₂ receptors have been localized in osteocytes, osteoblasts and osteoclasts where they modulate bone formation and turnover.³⁹ CB₂ receptors are also expressed in myofibroblasts of cirrhotic liver but their presence in normal liver is low.⁴⁰ Several studies have reported that CB₂ receptors in the enteric nervous system modulate ileum contractility.⁴¹ In rodents, CB₂ mRNA has been identified in cerebellum, cortex, brainstem and in glial cells, whilst in humans these receptors are expressed in microglial cells and in dorsal root ganglion.⁴² CB₂ receptors are implicated in microglial migration and infiltration into brain areas with active inflammation and degeneration, although the healthy brain microglia do not express these receptor subtypes.⁴³ In accordance with this evidence, CB₂ receptors are expressed in the brain only under certain conditions where inflammation is present at high levels.⁴⁴ In addition, it cannot be assumed that immune responses are solely regulated by CB₂ receptors because CB₁ receptor transcripts have also been found in immune cells and tissues such as Jurkat T cell line, human lymphoblastoid and macrophage like cells.⁴⁵

4. INTRACELLULAR PATHWAYS OF CB₂ RECEPTORS

Several reports present evidence that CB receptor molecular structure is similar to the GPCRs conformation and constituted of seven transmembrane domains with biochemical and cellular determinations of G proteins signal transduction.^{10,20,22} Both CB receptors are primarily coupled to Gi/Go proteins and inhibit adenylyl cyclase activity and cyclic adenosine monophosphate (cAMP) production.³⁰ CB₁ receptors are positively coupled to the potassium channels and decrease N-type and P/Q-type calcium channel conductance playing a key role in the suppression of neuronal excitability and neurotransmitter release.⁴⁶

The signaling pathways used by CB₂ receptors (Figure 1) depend on the type of cells and tissues where the receptor is localized, the specific G protein involved and the kinases present in the cells.²⁸ In particular, CB₂ receptor signaling mechanisms involve inhibition of adenylyl cyclase, mitogen-activated protein kinase (MAPK) activation and a transient increase in intracellular free calcium levels via a phospholipase C (PLC) modulation.⁴² CB₂ receptors are linked to three main components of MAPK pathway that have been identified as extracellular receptor kinase (ERK), Jun N-terminal kinase (JNK) and phospho-p38.⁴⁷ It is well reported that ERK–MAPK pathways are essential in mitogenesis and also associated to the cell proliferative stages of cancer. Interestingly, CB₂ receptors regulate ERK–MAPKs, depending on the cell type and experimental conditions employed.⁴⁸

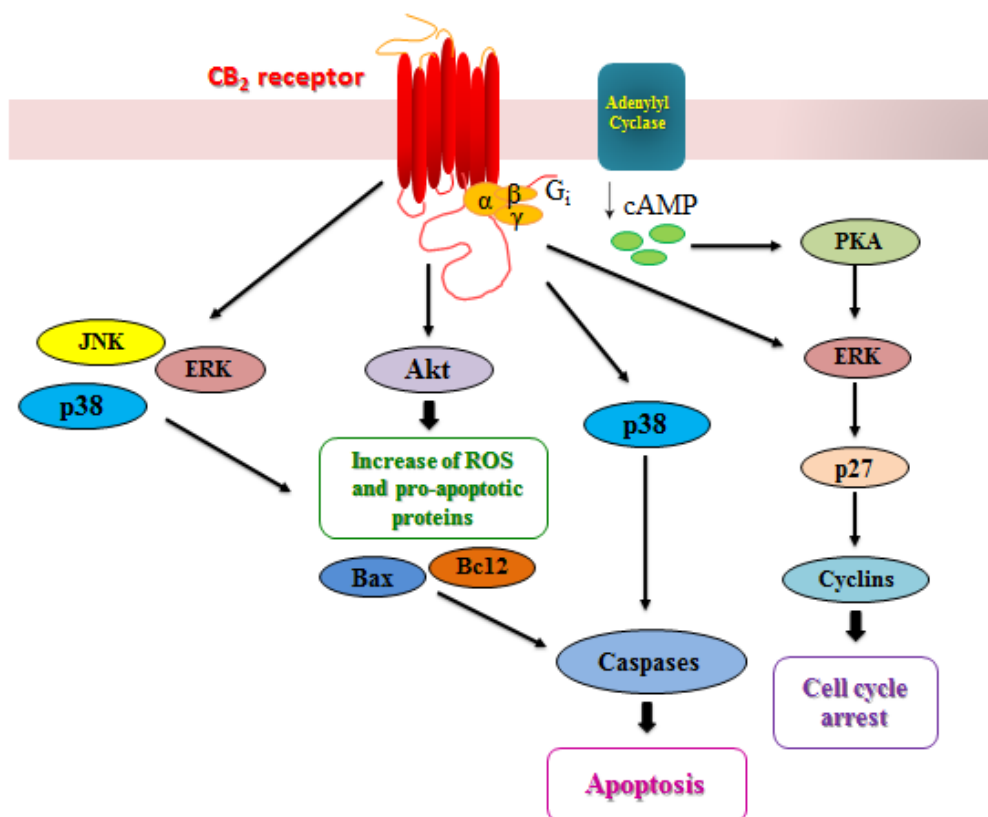


Figure 1 – Scheme of the main signaling pathways mediated by CB₂ receptors.

cAMP, cyclic adenosine monophosphate; Gi, inhibitory class of G protein; PKA, cAMP dependent protein kinase; JNK, jun N-terminal kinase; p38, p38 mitogen-activated protein kinase; ERK, extracellular receptor kinase; Akt, protein kinase B; Bax, apoptosis regulator Bcl2 like protein 4; p27, cyclin dependent kinase; ROS, reactive oxygen species.

CB₂ receptors are also involved in the inhibition of the intercellular communications through a mitogen activated protein kinase kinase (MEK)-dependent pathway and the ERK family activation involving different cellular responses like proliferation, apoptosis and differentiation.⁴⁹ It has been reported that CB₂ receptor stimulation counteracts the ability of morphine to upregulate protein kinase B or Akt and ERK1/2 activation induced by lipopolysaccharide (LPS), thus reducing nitric oxide (NO) and proinflammatory cytokine release.⁵⁰ The ability to modulate microglia and MAPK could be very interesting because their activation in the CNS and peripheral system contributes to morphine tolerance and dependence.⁵¹ It has also been demonstrated that CB₂ receptor stimulation activates the MAPK pathway, inhibits pro-inflammatory LPS-induced production of NO and the attenuation of the inducible nitric oxide synthase (iNOS).⁵²

Moreover, CB₂ receptor modulation promotes anti-inflammatory therapeutic responses in activated microglia, suggesting a regulatory role for CB₂ receptors in preventing excessive microglial cell response to injury.⁵³ Different studies have demonstrated that the presence of ceramide has been associated with decreased proliferation and apoptosis in glioma cells closely associated with CB₂ receptor stimulation via serine palmitoyl transferase, Raf-1 activation and MAPK (p42/44) activation.⁵⁴ It has been observed that CB receptors enhance the antinociceptive properties of opioids, and adolescent exposure to CB agonists blocks opiate dependence.⁵⁵ Opioid and CB receptors are coupled to similar intracellular signaling mechanisms leading to a decrease in cAMP production through the activation of Gi proteins. CB receptors also interfere with the tolerance and dependence effects induced by opioids because the administration of low-dose combinations of CB and opioid drugs seems to be an alternative regimen that reduces the need to escalate opioid dose, while increasing opioid potency.⁵⁶

It is well reported that CB₂ agonists produce pain relief in a variety of animal models by acting on glia and neurons, inhibiting the release of pro-inflammatory cytokines such as interleukin (IL) family and tumor necrosis factor- α (TNF- α). CB₂ agonists are able to prevent the stress-induced

increase in proinflammatory cytokines, nuclear factor-kB (NF-kB), cyclooxygenase-2 (COX-2) and the cellular oxidative and nitrosative damage such as lipid peroxidation.⁵⁷ CB₂ agonists inhibit IL-1 β -induced proliferation of the fibroblasts in rheumatoid arthritis (RA), as well as the production of metalloproteinases (MMP) such as MMP-3 and MMP-13 in a dose-dependent manner.⁵⁸ However, CB₂ receptor activation decreases liver fibrosis by reducing IL-17 production through Th17 lymphocytes via a STAT5-dependent pathway, and inhibits the proinflammatory effects of IL-17 by preserving IL-22 production.⁵⁹ The spinal administration of the anti-inflammatory cytokine, IL-10, abolishes pathological pain and suppresses IL-1 β and TNF- α release, similarly to CB₂ agonists which behave like anti-inflammatory drugs.⁶⁰ A novel mechanism of CB₂ receptor activation has been proposed and associated to an increase in β -endorphin release and to MAPK signaling pathway involvement, suggesting the potential role of CB₂ agonists as peripheral analgesic agents.⁶¹

5. THE ENDOCANNABINOID SYSTEM

The endocannabinoid system is modulated by several structurally related lipid mediators belonging to either the amide or ester families which are involved in the modulation of CB receptors.⁶² Among those, the most potent endogenous CBs are biologically active fatty acid ligands, namely, AEA and 2-AG.⁶³ AEA is formed by a family of membrane phospholipids such as N-arachidonoyl-phosphatidylethanolamine through multiple pathways. The production of AEA could be due to the action of a specific phosphodiesterase Ca²⁺ dependent that catalyzes the hydrolysis of the phospholipid, or to the sequential action of a hydrolase with a phosphodiesterase that converts the glycerophosphoanandamide.⁶⁴ A biosynthetic precursor, 1-acyl-2-arachidonoyl-glycerols (DAGs), produced from the hydrolysis of phosphatidylinositols, could be directly converted into 2-AG through the action of two Ca²⁺ sensitive lipases.⁶⁵

The catabolic enzymes for these endocannabinoids are also different, suggesting that their regulation is independent of each other even if they are broken down inside the cell by intracellular enzymes.⁶⁶ Different hydrolytic enzymes, such as the fatty acid amide hydrolase (FAAH), N-

acylethanolamine hydrolyzing acid amidase (NAAA) and monoacylglycerol lipase (MAGL),⁶⁷ have been identified as forming complex enzymatic cascades that regulate endocannabinoid production and inactivation. In particular, NAAA is a cysteine hydrolase belonging to the N-terminal nucleophile hydrolase superfamily that preferentially hydrolyzes palmitoylethanolamide and NAPE-selective phospholipase D catalyzes the synthesis of AEA, and the FAAH is involved in AEA degradation into arachidonate and ethanolamine.⁶⁸ The selective diacylglycerol lipases (DAGL) α and β are thought to catalyze the formation of 2-AG, and MAGL degrades this compound into arachidonate and glycerol.⁶⁹ They have relatively short half-lives that could hamper pharmacological characterization by using *in vivo* selected models, which highlights the need to identify more stable analogues.⁷⁰

Endocannabinoids are released on demand following specific and appropriate stimuli, and their lipophilic nature allows them to diffuse passively through the cell membrane.⁷¹ In addition, these hydrophobic compounds are able to cross the cell membrane by using a putative membrane transporter that facilitates their uptake between aqueous intercellular space and other intracellular carrier proteins known as fatty acid binding proteins (FABPs).⁷² Multiple lines of evidence suggest the potential use of selective inhibitors involved in enzymatic degradation of endocannabinoids that could provide selected therapeutic opportunities.⁷³

From a pharmacological point of view AEA is a partial CB₁ agonist and a weak partial CB₂ agonist, whilst 2-AG acts as a typical full CB₂ agonist.⁶⁵ Apart from their capability to bind CB receptors, endocannabinoids are able to interact with other receptors like the vanilloid receptor 1 (VR1), the transient potential vanilloid type 1 (TRPV-1) channels and other specific GPCRs such as opioid or GPR55 and GPR35 receptors.⁷⁴ The capability of the endocannabinoids to interact with different receptors allows a high number of biological and pharmacological actions. As a consequence, endocannabinoids do not regulate only the activity of CB receptors, but also control cell homeostasis through coordinated and fine interactions with different receptor sites. For

example, in several cancer types, CB₁, CB₂ and TRPV-1 receptors appear to be increased if compared with the corresponding non tumoral cells, and their activation counteracts cancer growth and cell invasiveness.⁷⁵ Moreover, AEA activating other GPCRs suggests the possibility of a multiple mechanism acting on the regulation of synaptic plasticity that could be very important in the disorders based on neuroinflammation, stress, emotionality and cognition.^{75,76}

Interestingly, alternative molecular targets for endocannabinoids have been identified, such as the inhibition of T-type Ca²⁺ channels, the modulation of nicotinic acetylcholine receptors and the allosteric enhancement of glycine receptors. Moreover, 2-AG has been reported as able to potentiate GABA-A receptors through a direct modulation with neurosteroids on the subunit located extra synaptically. An inhibitory effect of AEA and 2-AG has been shown on several types of K⁺ channels. A similar direct effect of these endocannabinoids has been verified on peroxisome proliferator activated receptor- γ (PPAR- γ) related to its pro-adipogenic function in adipocytes.⁷⁷ The discovery that the endocannabinoids interact with different receptor sites has contributed to the identification of this signaling system as a versatile tool to control various functions in the cells and tissues of the organism.

6. PHARMACOLOGICAL ROLE OF CB₂ RECEPTORS

Increasing evidence has been published implicating a role for CB₂ receptors in a variety of disease models involving central or peripheral nervous systems (Figure 2). It is well recognized that the psychoactive effects of CBs are associated with CB₁ and the CB₂ receptors which mainly mediate anti-inflammatory and immunomodulatory actions.⁷⁸ Briefly, CB₁ agonists possess neuroprotective properties primarily via diminishing excitotoxicity in postsynaptic neurons, enhancing vasodilation in vascular smooth muscle and inhibition of endothelin-1.⁷⁹ The therapeutic limitations of CB₁ agonists are related to the delicate balance between their beneficial actions and psychoactive effects.⁸⁰

Therefore, selective CB₂ agonists could represent an attractive target for the development of neuroprotective therapies and, above all provide to anti-inflammatory effects without psychoactive activities.^{81,82}

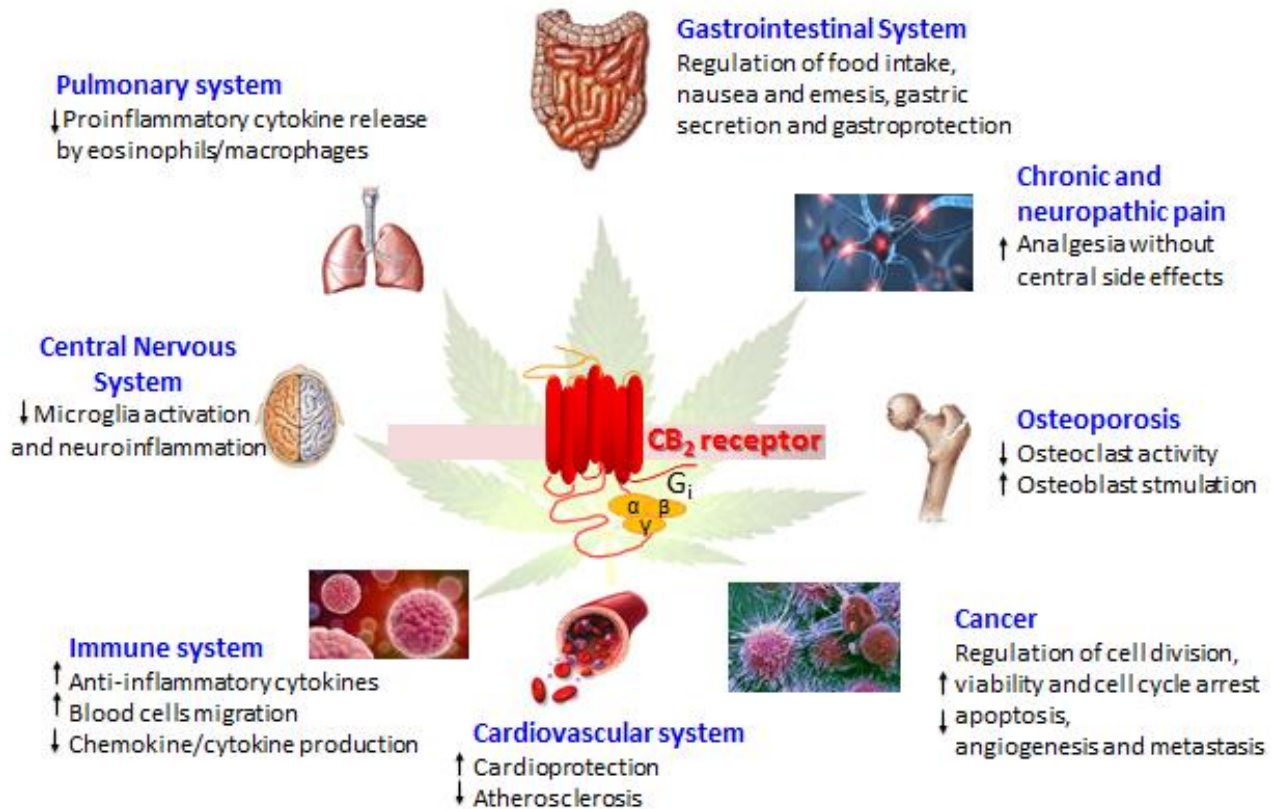


Figure 2 – Scheme for the functional role of CB₂ receptors.

The mechanisms of CB receptor regulation highlight the pathophysiology of diseases and are of interest from a therapeutic point of view. Recently, it has been reported that during cell or tissue injury CB₂ receptors are upregulated, and their activation has an important protective role, suggesting the potential use of selective agonists in different pathologies.⁸³ Although the physiological role of endocannabinoids and CB receptors in the regulation of cellular signaling has been well studied, relatively little is known about the involvement of CB receptors in human disease

processes. To better investigate the functional contribution of the CB₂ receptor to disease regulation, it may be of interest to evaluate the genetic variations in the human CB₂ gene causing an alteration in the CB₂ receptor function.^{84,85} Receptor function could be closely associated with the regulation of gene expression and/or with the presence on the cell surface of the receptors affected by the cell trafficking, receptor dimerization and desensitization/downregulation as well as an altered ligand binding and G protein coupling.³¹ It is also reported that the capability of the compounds to modulate pharmacological effects is closely associated to the drug-receptor interaction.⁸⁶ Thermodynamic analysis of the binding equilibrium permits us to evaluate the standard enthalpy and entropy that are responsible for the drug-receptor recognition phenomenon. Enthalpy is related to hydrogen bonding and multipolar interactions, whilst entropy is associated to the solvent reorganization.⁸⁶ In particular, CB₁ or CB₂ agonist binding is always totally entropy driven while antagonist binding is enthalpy and entropy driven, suggesting that CB₁ and CB₂ receptors are thermodynamically discriminated.⁸⁷ The thermodynamic discrimination of the CB ligands confirms the data obtained for other compounds interacting with GPCRs, such as adenosine, opioid, dopamine D₂ and adrenergic receptors.⁸⁷⁻⁹⁰ Experimental studies performed on several membrane receptors have demonstrated the presence of the enthalpy-entropy compensation also for ligand-gated ion channels like serotonin 5HT₃, glycine, GABA-A and nicotinic receptors.⁸⁶ Characterization of the enthalpy and entropy contribution of CB receptor binding could provide important information regarding the ligand-receptor interaction with fundamental implications for the decision-making process in drug development of novel CB ligands.

6.1. Immune system and inflammation

Most of the effects of CBs within the immune system have been attributed to CB₂ receptors, expressed in different types of leukocytes, that are able to mediate anti-inflammatory effects and immunomodulation.⁹¹ CBs modulate immune responses during inflammatory processes and their immunomodulatory effects have been studied in many disease models such as RA, multiple

sclerosis (MS), diabetes and septic shock.^{58,92} CBs exert their immunomodulatory properties through the induction of apoptosis, suppression of cell proliferation, inhibition of proinflammatory cytokine/chemokine production, increase in anti-inflammatory cytokines, and induction of regulatory T cells.⁹³

Interestingly, the levels of CB₂ receptors can be upregulated by inflammatory cytokines such as IL-1 β , IL-6 and TNF- α .⁹⁴ It has also been shown that CB₂ receptor activation inhibits the proliferation of human lymphocytes and the proliferative response of T and B cells to mitogens through induction of apoptosis.⁹⁵ Direct measurements of oxidative stress have revealed that CBs prevent serum-deprived cell death by antioxidation, confirming their ability to antagonize oxidative stress and consequent cell death.⁹⁶ Stimulation of CB₂ receptors triggers apoptosis in immune cells, suggesting the possibility of novel pharmacological approaches to the treatment of inflammatory and autoimmune diseases.⁹⁷ CB₂ receptors also regulate migration of blood cells such as lymphocytes, neutrophils, monocytes and macrophages which are influenced by CB in a dose dependent manner and by receptor stimulation.⁹⁸ In particular, CB₂ receptor expression and function is upregulated in immature B cells, fundamental in their retention and subsequent development of these cells into the bone marrow. CB₂ receptor stimulation increases the activity of the α 4 β 1 integrins and vascular adhesion molecule (VCAM-1), suggesting a selective cooperation into cell migration.⁹⁹

Pre-clinical studies have demonstrated that CB drugs have therapeutic potential in different inflammatory diseases, including RA and osteoarthritis (OA).^{100,101} CB₁ and CB₂ protein and RNA are present in the synovia of RA and OA patients. CB receptor stimulation of synovial fibroblast-like cells from RA and OA patients produce a time-dependent phosphorylation of ERK-1 and ERK-2 determining a key role of the endocannabinoid signalling system.¹⁰² Selective activation of CB₂ receptors modulate chemotaxis of human monocytes, and phosphatidylinositol 3-kinase/Akt and ERK1/2 showing an important role in the pathophysiology of RA.¹⁰³

CB₂ receptors appear to be involved in allergic dermatitis where their modulation mediates an inflammatory response elicited by antigen-specific effector T cells upon repeated allergen contact.¹⁰⁴ Contrasting preclinical studies are reported regarding the involvement of CB₂ receptors and allergic dermatitis, suggesting the potential use of both CB₂ agonists or inverse agonists/antagonists in the pharmacological therapy.¹⁰⁵

The additional role of CB₂ agonists has been studied in experimental autoimmune uveoretinitis, an animal model for human posterior uveitis induced by immunization with retinal proteins or their peptide fragments characterized by retinal infiltration in autoreactive T cells. The efficacy of the pharmacological treatment has been impressive with clinical scores positively correlated with the number of infiltrating cells.¹⁰⁶

6.2. Central nervous system

For several years CB₂ receptors were thought to be present only in the periphery, whilst CB₁ receptors were the predominant form within the brain. Recently, it has become apparent that CB₂ receptors are present in the CNS, even if in lesser quantities than CB₁ receptors. In particular, the neuroprotective effects of CB₂ agonists involve the microglia and are associated with suppression of microglia activation via inhibition of the release of neurotoxic factors and by decreasing neuronal damage in cellular or tissue culture models.^{107,108}

Increased expression of CB₂ receptors under neuroinflammatory conditions has been found in human brain.¹⁰⁹ Prominent CB₂ upregulation has been reported in brain tissues affected by MS and amyotrophic lateral sclerosis.¹¹⁰ In Alzheimer's disease, a potential use of CB₂ specific radiotracer as an imaging biomarker of neuroinflammation in the early stages of this disease has been reported.⁸² In Parkinson's disease (PD), an upregulation of CB₂ receptors in glial elements in post mortem tissues of PD patients has been studied together with marked anti-inflammatory effects.^{111,112}

CB₂ agonists prevent neuronal injury during neuroinflammation via upregulation of MAPK that results in ERK1/2 inhibition.¹⁰⁸ CB₂ receptor stimulation specifically reduces iNOS production via inhibition of ERK1/2 phosphorylation in microglia during CNS inflammation.⁵³ These findings have relevance to anti-inflammatory effects of CB₂ stimulation in brain and the regulation of neuroinflammation, that have been confirmed in different animal models for neurodegenerative or inflammatory diseases.⁷⁸ However, CBs can be considered neuroprotective via their immunomodulatory properties, which have mainly been attributed to CB₂ receptors involving the inhibition of different pro-inflammatory events such as chemotaxis of immune cells, activation of endothelium and leukocyte infiltration into tissues and injury of endothelial barriers.¹¹³ CB₂ receptors appear to play a major role in leukocyte migration where their stimulation has shown a decrease in migration and in chemokine response via down-regulation of their receptors and inhibition of interferon γ (IFN γ)-induced intercellular adhesion molecule-1 (ICAM-1) expression.¹¹⁴

CBs have been reported to inhibit chemokine-induced chemotaxis of various cell types including blood cells and microglia where endocannabinoids drive the regulation of microglial activation in normal and pathological conditions.^{103,115} It has been shown that CB₂ agonists affect dendritic cell migration in vitro and in vivo, primarily through the inhibition of MMP-9 expression.¹¹⁶ Anti-inflammatory properties of CB₂ agonists may be related to their actions on the endothelium. CB₂ receptors have been found in brain endothelium and in endothelial cells from other organs and are able to reduce TNF α -induced activation of human coronary artery endothelial cells, the secretion of monocyte chemoattractant protein 1 (MCP-1) and attenuated monocyte migration.¹¹⁷

CBs have been investigated as potential candidates for MS, a chronic inflammatory demyelinating disorder of the CNS characterized by autoreactive myelin-specific T cells and infiltrating macrophages that cause inflammation resulting in demyelinated plaques and axonal loss.¹¹⁸ An important role in the pathogenesis of MS could be played by the resident microglial cells and by CB₂ receptor stimulation that suppresses microglial cell activation.¹¹⁹ The activation of CB₂

receptors reduces IL-12 and IL-23 cytokines in human, and murine microglial cells.¹²⁰ The chronic treatment of CB₂ agonists decreases plasma and cerebrospinal fluid viral load and tissue inflammation, significantly reducing morbidity and mortality in an immunodeficiency virus-infected animal model.¹²¹ CB₂ receptor-mediated control of bone marrow-derived myeloid progenitor cell trafficking to the inflamed spinal cord has been proposed.¹²² While CB₂ agonists have been reported as having a protective effect in cerebral ischemia because they are related to improvements in vascular reperfusion through inhibition of leukocyte rolling and adhesion to cerebral microvessels, CB₁ receptor activation was found to be deleterious.¹²³

The effects of CB₂ receptors on the CNS are controversial as far as their role in depression and/or substance abuse is concerned. Behavioral and molecular methods have been performed to study the presence of a link between CB₂ receptors and drug/alcohol addiction.^{124,125} A direct association has been suggested with the polymorphism of *Cnr2* gene, encoding for CB₂ receptors, in alcoholism and neuropsychiatric disorders.¹²⁶ Mice preferring alcohol had reduced *Cnr2* gene expression in the ventral midbrain, whereas it was unaltered in mice with little or no preference for alcohol.¹²⁷ Animals treated with cocaine or heroin showed increased *Cnr2* gene transcripts in comparison with controls, indicating that this gene alteration in the brain could be influenced by substance abuse.¹²⁸ CB₂ receptors are also involved in the endocannabinoid signaling mechanisms associated with the regulation of food intake and in eating disorders, including anorexia and bulimia nervosa, that occur most commonly in young women and much less frequently in men.¹²⁹ It has been observed that the involvement of CB₂ receptors in eating disorders and food consumption may be due to a dysregulation in the immune system and to the presence of these receptors in hypothalamic nuclei linked to feeding behavior.¹³⁰ A high association between R63Q polymorphism of the *Cnr2* gene and eating disorders with a significant suppression of food intake in a time dependent manner has been reported.¹³¹

6.3. Cardiovascular system

In the literature, several papers have established a positive role of CB₁ receptors in the cardiodepression associated with various forms of stroke and heart failure, even if modulation of these receptors is associated with adverse psychiatric effects.¹³² On the other hand, the protective effect of CB₂ receptors on ischemia reperfusion (IR) injury, a pathophysiologic process due to inadequate blood supply and to hypoxic tissue damage, has also been investigated.^{133,134} CB₂ receptor knockout mice develop an increase in the IR-induced hepatic tissue damage, accompanied by elevated levels of proinflammatory markers such as TNF- α , or macrophage inflammatory proteins such as MIP-2 and MIP-1 α or ICAM-1 as an example of chemokine secretion.¹³⁵ The activation of CB₂ receptors mediates a suppression of proinflammatory cytokines and a reduction in tissue damage, low levels of lactate dehydrogenase enzymes, neutrophil infiltration, tissue lipid peroxidation and DNA fragmentation.¹³⁶ CB₂ agonists significantly reduce cerebral infarction in different animal models, suggesting a significant protective effect of CB₂ receptors.¹³⁷ A cardioprotective mechanism of CB₂ receptor is associated with the modulation of inflammatory response and myocardial remodeling during cardiac adaptation to pressure overload in a murine model of transverse aortic constriction.¹³⁸ The relevance of the endocannabinoid-CB₂ receptor axis has been underlined by the findings that CB₂ receptors are upregulated in ischemic wild type cardiomyocytes and that CBs levels are transiently increased during IR.¹³⁹ A protective role for CB₂ receptors on the cardiac myocytes from hypoxic damage and an increase in NO production by the stimulation of iNOS has been provided. The ventricular arrhythmias induced by coronary occlusion are reduced after administration of CB₂ agonists, an effect blocked by the use of selective CB₂ antagonists.¹⁴⁰

6.4. Atherosclerosis

Atherosclerosis is a chronic inflammatory disease of the vascular wall that involves a complex interplay between vascular endothelial and smooth muscle cells, mononuclear cells, growth factors,

and cytokines.¹⁴¹ Selective CB₂ agonists significantly reduce the progression of atherosclerosis, which implies a protective role for CB₂ receptors that inhibit the TNF- α activation of endothelial cells and the subsequent transendothelial migration of monocytes.¹⁴² Infiltrating macrophages as well as T cells in the atherosclerotic plaques and lesions are found to express CB₂ receptors which modulate apoptosis associated with the development and progression of atherosclerosis.¹⁴³

CB₂ receptor deficiency reduces the susceptibility of macrophages to the apoptosis induced by 7-ketocholesterol, the major oxysterol present in oxidized low-density lipoprotein (oxLDL) due to suppressed activity of caspase-3. In addition, a selective activation of CB₂ receptors reduces the oxLDL and the expression of CD36 scavenger receptor.¹⁴⁴ Moreover, CB₂ receptor activation inhibits the ICAM-1 and VCAM-1 expression in the aorta and adhesion of monocytes to aortic vascular endothelium. CB₂ agonists also attenuate cell proliferation of human vascular smooth muscle and modulate different signaling molecules such as Ras, p38 MAPK, ERK-1, ERK-2 and Akt.¹⁴⁵

CB₂ receptors attenuate many of the principal steps involved in the development and progression of atherosclerosis, and their activation modulates anti-inflammatory and protective actions within atherosclerotic vessels in both atherosclerosis human and animal models.¹⁴⁶ As a consequence, the basic and preclinical research data suggest that medications activating CB₂ function in the peripheral target organs might be a promising approach against atherogenesis.

6.5. Gastrointestinal system and liver

Cannabinoids have been used for the treatment of many gastrointestinal (GI) disorders, including anorexia, emesis, abdominal pain, diarrhea, even if their use is limited owing to the psychotropic side effects. Several cannabinoid receptors, which include CB₂ receptors, have been described in the esophagus, stomach, ileum and gut, that is an organ of host defence and has an extensive immune system.¹⁴⁷ The most abundant source of these receptors is expressed in the longitudinal muscle with the adherent myenteric plexus.¹⁴⁸ These receptors may play a role in the regulation of food intake,

nausea and emesis, gastric secretion and gastroprotection, GI motility, ion transport, visceral sensation, intestinal inflammation, and cell proliferation in the gut.¹⁴⁷

The expression of CB₂ receptors is markedly upregulated in different types of liver injury and diseases. Immunoreactivity of these receptors has been localized on myofibroblasts, in fibrotic septa, hepatocytes, biliary epithelial cells derived from active cirrhosis, steatosis and hepatocellular carcinoma.^{149,150} Molecules stimulating CB₂ receptors are promising pharmacological tools considering that liver cirrhosis and related complications represent important clinical application in the gastrointestinal system.¹⁵¹

Different reports support a functional role of CB₂ receptors in the control of stomach and intestinal motility.¹⁵² It has been demonstrated that, through the involvement of CB₂ receptors, CBs stimulate the release of vasoactive intestinal peptide from enteric synaptosomal preparations.¹⁵³ Moreover, the interaction between the immune and nervous system serves to maintain a barrier that protects the gut which, in particular conditions, may become chronically inflamed.¹⁵⁴

CB₂ receptor stimulation significantly inhibits an experimental model of colitis in mice induced by oil of mustard and dextran sodium sulfate and, in an *in vitro* model represented by HT-29 cells, suppresses the IL-8 production TNF- α induced.¹⁵⁵ Immunohistochemical analysis has revealed the presence of CB₂ receptor expression in human colonic tissue on plasma cells and macrophages in the lamina propria in inflammatory bowel diseases, ulcerative colitis and Crohn's disease.^{156,157} In particular, increased expression of CB₂ receptor has been reported in the epithelium, implying a role for CB₂ receptor in colonic inflammation.¹⁵⁸ These data further confirm that CB₂ receptor activation protects against experimental colitis, suggesting that these receptors may be a novel therapeutic target in this pathology.¹⁵⁹

6.6. Pulmonary system

Literature data have highlighted the potential importance of CB receptors on the regulation of the airways functions and cough reflex. CB₂ receptor activation prevents bronchoconstriction and

airway edema in a guinea-pig model of gastro-esophageal reflux.¹⁶⁰ Coughing is a very common clinical symptom, and current therapies are largely ineffective, indicating a major pharmacological need. CB₂ agonists appear to be protective in the cough reflex and this effect is blocked by the presence of CB₂ antagonists suggesting the direct involvement of this receptor.¹⁶¹

Stimulation of the CB₂ receptor signalling pathway diminishes cAMP accumulation and IL-8 release induced by TNF- α , suggesting that CBs could exert anti-inflammatory properties in airways by modulating cytokine release.¹⁶² It has also been reported that the activation of β 2-adrenoceptor and CB₂ receptors can inhibit sensory nerves and prevent coughing.¹⁶³ CB₂ agonists, together with TRPV-1 or P2X₃ antagonists, could represent the most promising drugs in the treatment of coughs, since they are involved in nociception, acting as MAP38 kinase inhibitors, and the reduction of the inflammatory status.¹⁶⁴

Moreover, melilotus extracts, that have been used to treat inflammatory infectious diseases, have significantly reduced lung inflammation in an animal model of acute lung injury, since they induce a significant up-regulation of CB₂ receptor expression.¹⁶⁵ As a consequence CB₂ receptor activation mediates immunosuppressive effects, which limit inflammation and associated tissue injury in various pathological conditions, suggesting that the protective actions of CB₂ receptors with endocannabinoids or synthetic agonists could be implied in the biological mechanisms involved in these diseases.¹⁶⁶

6.7. Osteoporosis

CB₂ signaling contributes to the maintenance of bone mass through direct stimulation of stromal cells/osteoblast, inhibition of monocytes/osteoclasts and the receptor activator of nuclear factor κ B ligand (RANKL) expression.³⁹ It has been reported that CB₂ receptors are well expressed in osteocytes or osteoblasts or bone forming cells, and osteoclasts or bone resorbing cells.¹⁶⁷

CB₂ agonists are able to mitigate osteoclastogenesis by inhibiting the proliferation of osteoclast precursors and mediating a mitogenic effect on osteoblasts.⁸⁵ In these cells, CB₂ and TRPV1

receptors are both expressed together and, in culture, oppositely modulate human osteoblast activity in such a way that the CB₂ receptor stimulation improves the osteogenesis, whereas TRPV1 receptor stimulation inhibits it.¹⁶⁸

CB₂ receptor knockout mice have accelerated age-related trabecular bone loss and cortical expansion, that also characterize human osteoporosis. Given the low bone mass phenotype in this mice model, it is plausible that the endocannabinoid system could play an essential role in maintaining bone mass, suggesting that CB₂ receptors could be a potential drug target for osteoporosis therapy.¹⁶⁹ Accordingly, CB₂ receptor activation might be an endogenous protective mechanism against the inflammation and the age-related bone loss, osteo-protective, especially with regard to the aging skeleton.¹⁷⁰

A molecular-modeling analysis has revealed that CB₂ receptor enantiomers have two different binding conformations able to discriminate the affinity or the potency of the orientation-targeted ligands, which could have a promising potential for the pharmacological activation of distinct processes.¹⁷¹ In addition, CB₂-selective compounds show a good efficacy in leukocyte recruitment models when added with low doses of selected anti-inflammatory agents, consistent with their biological functions and indicative of their potential therapeutic utility.¹⁷²

6.8. Cancer

A great interest in CBs as potential anticancer agents has been reported since they inhibit tumor growth in different animal models.¹⁷³ Significant upregulation of CB₂ receptors has been shown in prostate cancer cell lines if compared with normal prostate cells, and in lymphoma and breast cancer tissues.¹⁷⁴ The degree of increased expression of the CB₂ receptors correlates with tumor aggression and progression.¹⁷⁵ It is well known that CB agonists affect a range of pathways that regulate cell division and viability and a wide range of markers associated with the invasion and metastasis of cancers, including markers of migration, adhesion, invasion and metastasis.¹⁷⁶ Endocannabinoids regulate the synthesis of ceramides, lipid-based components of the cell

membrane that perform both structural and signaling functions associated with the control of apoptosis, growth arrest, differentiation, cell migration and adhesion.¹⁷⁷ CB₂ receptor activation is closely linked to an increase in ceramide levels leading to the involvement of ERK and p38MAPK activation.¹⁷⁸ It has been reported that adjuvant cannabinoid use in the treatment of adverse side effects from chemotherapy, such as neuropathic pain, loss of appetite, nausea and vomiting, is the most studied potential therapeutic application for these compounds.¹⁷⁹ Endocannabinoids are recognised for their role in the regulation of the key processes involved in the development of cancer.¹⁸⁰ For example, the endocannabinoid system is reported to induce apoptosis, cell cycle arrest and the inhibition of angiogenesis and metastasis in animal models and in cell lines.^{181–183} The variability of endocannabinoid effects in different tumor models is highly incongruous and most likely derives from the differential expression of CB receptors, playing a determining role in the progression and/or inhibition of malignancy. High levels of CB₂ mRNA have been detected by *in situ* hybridization in well differentiated human hepatocellular carcinoma and in cirrhotic liver samples, while a low expression of these receptors has been revealed in poorly differentiated hepatocellular carcinoma.¹⁵⁰

In addition, increased expression of CB₁ and/or CB₂ receptors has been noted in human lymphoma, breast cancer, acute myeloid leukaemia, hepatocellular carcinoma and prostate cancer cell lines.^{184–186} In general, a relationship between CB receptor expression and the outcome of cancer has been documented. In astrocytoma cells, 70% of cells expressing CB₁ and/or CB₂ receptors directly correlates with the degree of tumor malignancy, whilst in gliomas a higher expression of CB₂ compared to CB₁ receptors is found which is closely related to the tumor grade.¹⁸⁷ Moreover, tumor-associated endothelial cells of human glioblastoma multiforme indicate the high presence of CB₂ receptors that are able to alter blood vessel morphology.¹⁸⁸ As a consequence, CBs may produce a dual attack on the development of tumor blood vessels through the inhibition of pro-angiogenic regulators, such as vascular endothelial growth factor (VEGF), and

through a direct effect on endothelial cells.¹⁸⁹ It has been found that the CB₂ receptor stimulation decreases the release of NO and other pro-inflammatory mediators in macrophage cell lines and NF-kB-dependent apoptosis in immune cells.¹⁹⁰ The local administration of CB₂ agonist reduces chronic granuloma formation induced by carrageenan-soaked rat implant associated with angiogenesis by inhibiting NF-kB activation.¹⁹¹ Similarly, increased expression of both CB₁ and CB₂ receptors has been documented in non-Hodgkin lymphoma when compared to reactive lymphonodes.¹⁸⁴

CB₂ receptor stimulation, which has been investigated in skin melanoma, reduces growth, proliferation, angiogenesis and metastasis.¹⁹² Activation of these receptors decreases melanoma cell proliferation via inhibition of Akt, a key element of a major prosurvival pathway that is deregulated in many types of tumors including melanoma.¹⁹³ A reduced expression of CB₁, but not of CB₂, compared with the normal adjacent mucosa has been noted in colon cancer.¹⁹⁴ Of interest is the importance of CB₂ receptor in mediating protective effects by inhibition of colorectal cancer cell proliferation.¹⁹⁵ It seems that CBs might regulate the tissue response to gut inflammation via CB₂ receptor activation by involving the smooth-muscle response to pro-inflammatory mediators that affect gastrointestinal transit time, and by causing the direct suppression of pro-inflammatory mediator production.¹⁹⁶

In breast carcinoma, a relationship between CB₂ expression, the histological grade of the cancer, and other markers of prognostic and predictive value, such as HER-2 oncogene, oestrogen, and progesterone receptors, has been reported.¹⁹⁷ CB₂ receptors have been found in different breast carcinoma cell lines, such as MCF-7, T-47D, MDA-MB-231, MDA-MB-468, EVSA-T and SkBr3 and in human breast tissues using RT-PCR, immunofluorescence and/or Western blot assays.¹⁸⁰

The activation of CB₂ receptors in prostate cancer, the second most frequently diagnosed malignancy in men, where the cancer cells may metastasize to other body parts, mediates

significant anti-proliferative and pro-apoptotic properties.¹⁹⁸ As a consequence, endocannabinoids may be a beneficial option for the treatment of prostate cancer.¹⁹⁹

These studies imply a role for CB₂ receptors and their expression in relation to disease prognosis and outcome, closely associated with the specific cancer, and could help to translate this knowledge into the clinical setting to develop new pharmacological therapies.

6.9. Chronic and neuropathic pain

Available therapies often provide incomplete pain relief, and treatment-related side effects are common in chronic and neuropathic pain. Preclinical neuropathic pain models have facilitated identification of several promising targets, which have progressed to human clinical phases of evaluation.²⁰⁰ Some of the most important drugs that could be used are summarized: calcium channel antagonists, vanilloid receptor antagonists, NMDA antagonists, opioid receptor agonists, histamine H₃ receptor antagonists, serotonin modulators, acetylcholine and adrenoreceptor agonists, nitric oxide synthase inhibitors, orexin antagonists, apoptosis inhibitors, fatty acid amide hydrolase inhibitors and CB agonists.^{200,201} From these it is widely accepted that neurotrophin, CB₁ and CB₂, and thermo-transient receptor potential such as TRPs, TRPV1, TRPA1 and TRPM8 receptors could play a pivotal role in this pathology.^{202,203} In particular, endocannabinoids appear to be a first line of defence against pain, while neurotrophins and thermoTRPs are the major generators of painful signals.²⁰³ Recently, it has been reported in a clinical review on 28 randomized clinical trials of cannabinoids as pharmacotherapy that medical marijuana is used to treat a host of indications among them the treatment of chronic and neuropathic pain.²⁰⁴ Randomized clinical trials of cannabinoid drugs reported their potential use to treat disease or alleviate symptoms such as nausea and vomiting due to chemotherapy, depression, anxiety and sleep disorder, psychosis, treatment of chronic pain and spasticity.²⁰⁵ Although the diversity of pharmacological mechanisms of interest emphasise the complexity of neuropathic pain transmission, the considerable number of agents under development reflect a continued enthusiasm in drug development for neuropathic pain.²⁰⁶

The relevance of the therapeutic effects mediated by CB₂ agonists is well established for the treatment of inflammatory and neuropathic pain and neurodegenerative disorders.²⁰⁷ Commercially available CBs are subject to psychotomimetic and addictive adverse effects, largely through activation of the CB₁ receptors. Regional and peripherally restricted CBs could reduce the side effects of CBs. Spinal CBs may increase the therapeutic index by limiting the dose necessary for response and minimize drugs exposure to supraspinal sites where CB side effects originate. CB bifunctional ligands or the combination of a CB₂ agonist with a TRPV-1 antagonist may improve the therapeutic index of the CB₂ agonist. Selective enzyme inhibitors plus TRPV-1 blockers could be further explored to limit CB hydrolyzing enzyme inhibitors.²⁰⁸

Recently, it has been reported that the systemic administration of a CB₂ agonist produces robust analgesia in different pain models, such as writhing and formalin assays, streptozotocin (STZ)-induced diabetic neuropathy and bone cancer pain, providing an interesting approach to analgesic therapy in inflammatory and chronic pain without CB₁ mediated central side effects.²⁰⁹ These data have also been confirmed by the use of other CB₂ agonists in neuropathic pain of STZ-induced diabetic mice, suggesting that these compounds might have an ability to resolve diabetic neuropathic pain.²¹⁰ The combined administration of subthreshold doses of a COX inhibitor and a selective FAAH inhibitor increases CBs levels and decreases prostaglandin levels in whole brain. These data confirm that the dual blockade of FAAH and COX represents a potential therapeutic strategy for the treatment of neuropathic and inflammatory pain states.²¹¹ It has been also reported that the prolonged use of CB₂ agonists for managing chemotherapy-induced allodynia mediates a favorable therapeutic ratio marked by sustained efficacy and absence of tolerance, physical withdrawal, or CB₁-mediated side effects decreasing mRNA levels of TNF- α and MCP-1 in mice spinal cord.²¹²

Obtaining detailed information on the internalization and trafficking of the human CB₂ receptor in response to an agonist could have a significant impact on drug discovery. The internalized CB₂

receptors are co-localized with the early endosome probe and are recycled to the cell surface after the removal of the agonist, but treatment with a specific cell-permeable proteasome inhibitor reduces the recycling of internalized CB₂ receptor, suggesting that the proteasome-mediated degradation pathway may be involved in CB₂ internalization.²¹³ A novel CB₂ agonist has also been identified with high affinity versus CB₂ receptors, which failed to promote CB₂ internalization, suggesting a potential starting point for further investigations of CB₂ pharmacology and pain treatment.²¹⁴

The ability of CBs to produce signs of analgesia in chronic and neuropathic pain reveal important clinical implications. A major point in the clinic is the need to identify strategies that could reduce or abolish the adverse central effects of CBs without attenuating their clinical effects. One possibility could be to focus on CBs that induce analgesia by acting on biological targets, for example, through the use of selective CB₂ agonists. Another strategy could be activate the endocannabinoid system indirectly by inhibiting the tissue uptake or metabolism of endogenous CBs to increase their levels to CB₂ receptors. Moreover, the synergistic interactions between CBs and opioid or TRPV1 ligands for antinociception could be explored. The potential use of dualistic drugs would obtain therapeutic advantage in comparison with administering combine therapy with the primary aim to limit the unwanted side effects. A summary of the involvement of CB₂ receptors in different systems is reported in Table 2 to correlate their main pharmacological role with the potential therapeutic implications.

Table 2. Potential involvement of CB₂ receptors and their pharmacological role in various diseases.

System/ Disease	Proposed Role	Potential use in therapy	Ref
Immune and metabolic system	Inhibition of inflammation	RA, OA, allergic dermatitis, uveoretinitis, septic shock	91,92,215,216
Central nervous system	Inhibition of inflammation	MS, ALS, AD, PD, cerebral ischemia, neuropsychiatric diseases	109,122,123,217
Chronic and neuropathic pain	Inhibition of inflammation	Chronic pain associated to different disease state	206,211,218,219
Cardiovascular system	Decrease plaque progression	Stroke, heart failure, ischemia reperfusion, atherosclerosis	132,137,139,220
Gastrointestinal system and liver	Inhibition of inflammation	Colitis, diabetes	148–150,221
Pulmonary system	Reduce bronchoconstriction and edema	Coughs, lung injury	160–163
Bone system	Increase osteoblast activity	Osteoporosis	169,222–224
Cancer	Apoptosis and cell cycle regulation	Prostate, breast and colon cancer, hepatocellular carcinoma, myeloid leukemia, astrocytoma, glioblastoma	173,174,176,177

RA, rheumatoid arthritis; OA, osteoarthritis, MS, multiple sclerosis, ALS, amyotrophic lateral sclerosis, AD, Alzheimer disease, PD, Parkinson disease

7. CHEMISTRY OF CB₂ AGONISTS

The discovery of CB₁ and CB₂ receptors was followed by rigorous research efforts to investigate this interesting biochemical system and identify the key proteins implicated in its modulation. At the same time, the pharmaceutical industries and academic laboratories produced numerous new and structurally related compounds with very potent biological activities.

In the past years, the discovery of tricyclic and bicyclic analogs of tetrahydrocannabinol as well as the aminoalkylindole derivatives increased evidence to support the potential utility of selective CB₂ agonists for the treatment of pain. Compounds such as JWH133,^{225–230} HU308^{231,232} and AM1241^{233–235} have been extensively used in preclinical research and have demonstrated efficacy in preclinical models of inflammatory, moderate to severe post-operative and neuropathic pain.

The development of CB₂ selective ligands represents an important advance in cannabinoid therapeutics. In literature, it is well reported that clinical trials with published results showed the

presence of inactive drugs without efficacy. The reasons for this contrast may be linked to the localization of the CB₂ receptors that appeared to be present only peripherally, and to the use of unspecific antibodies that did not permit an exact mapping of these receptors. Another problem was that CB₂ knockout mice had only a small portion of the CB₂ gene which was partially translated into CB₂ proteins able to interact with other receptors.

Moreover, compounds which have significant selectivity and affinity for the CB₂ receptor belong to different structural groups. These include the traditional CB₂ ligands and the recently discovered CB₂ agonists, commonly characterized by lipophilic molecules, such as aromatic heterocycles, linked to bulky alkyl or aryl moieties.

The chemical section of the present review will provide an update of the progress made in structure-activity relationships in the field with a particular focus on the CB₂ agonists developed from 2008 onwards. We have included some synthetic schemes in order to show the chemistry involved in this field. Finally, a review of patent publications collects the most important cores of the CB₂ agonist molecules.

7.1. Pyridine, Pyridazine

Following the discovery of pyrimidine-5-carboxamide derivative **1** (GW842166X, Figure 3) in GlaxoSmithKline Research Laboratories as a clinical candidate, a program was begun to reach a CB₂ agonist back-up candidate with improved aqueous solubility over **1**.²³⁶ The replacement of the central pyrimidine core of compound GW842166X with a pyridine nucleus led to the pyridine-3-carboxamide CB₂ agonists.²³⁷ An extended SAR of the new pyridine-3-carboxamide template was concluded in the identification of analogue 6-(2,4-dichlorophenylamino)-4-cyclopropyl-N-(tetrahydro-2*H*-pyran-4-yl)methylpyridine-3-carboxamide **2** (Figure 3, Scheme 1), which demonstrated efficacy in an in vivo model of inflammatory pain, despite its low aqueous solubility. Boehringer Ingelheim also discovered a series of 2-aminopyridines bearing chiral morpholine ring

(compound **3**) which was reported potent and selective. (CB₂ EC₅₀ = 50 nM, efficacy (E_{max}) = 100%; CB₁ EC₅₀/CB₂ EC₅₀ > 2000).²³⁸

2-Amino-5-aryl-pyridines had been identified as a synthetically tractable series of CB₂ agonists from a high-throughput screen of the GlaxoSmithKline compound collection. Replacement of the pyridine core of the initial lead **4** (CB₂ EC₅₀ = 79 nM)²³⁹ with a pyridazine (compound **5**) brought about the discovery of a number of analogues with increased in vitro metabolic stability. Introduction of the *cis*-2,5-dimethylpyrrolidine moiety was found to significantly increase CB₂ activity. Combining this group with a number of bicyclic heterocycles led to the identification of compound **6** (Figure 1, Scheme 2), which showed high potency in a model of inflammatory pain.²⁴⁰

The CB₂ ligands based on the 3-carbamoyl-2-pyridone derivatives was also discovered. The structure-activity relationship around this template by adjusting the size of side chains at 1-, 5- and 6-positions led to the identification of **7** (S-777469, Figure 3) as a selective CB₂ receptor agonist. This compound exhibited moderate potency for CB₂ receptor (hCB₂ K_i = 36 nM) and good selectivity for CB₁ receptor (>120).²⁴¹

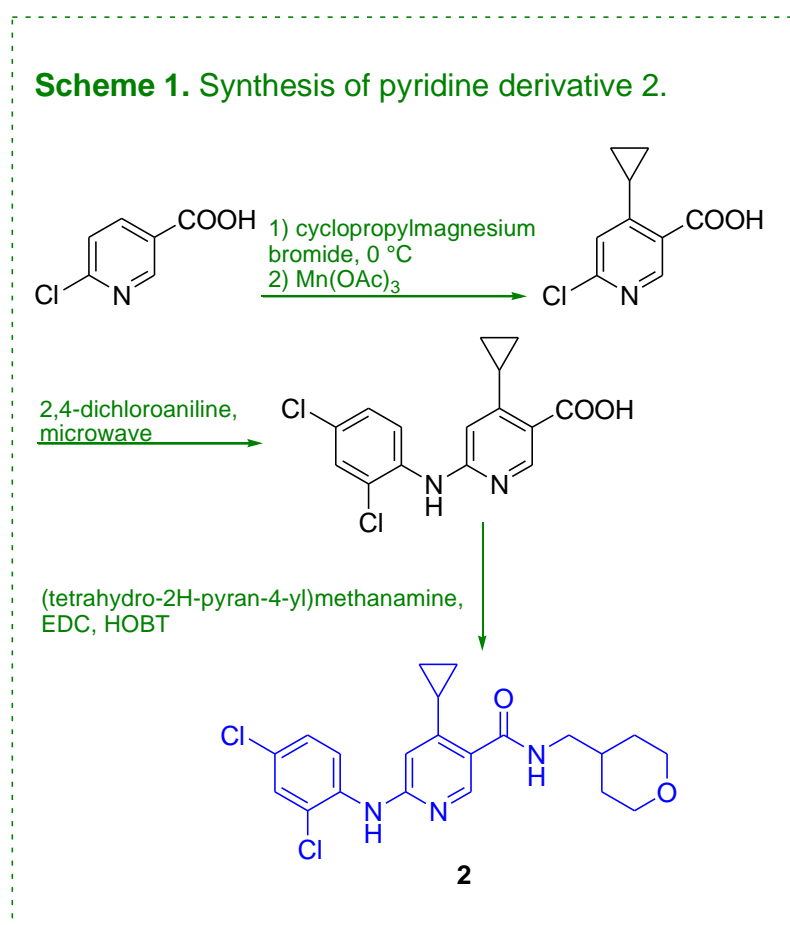
.....Merck Research Laboratories disclosed the 1-(4-(pyridin-2-yl)benzyl)imidazolidine-2,4-dione series as a novel chemotype cannabinoid CB₂ receptor agonist. The optimization of pharmacokinetic properties and human ether-a-go-go-related gene (hERG) affinity of this novel class of CB₂ agonists by systematically modulating physicochemical properties led to the synthesis of **8** (LU-101, Figure 3) as an orally bioavailable CB₂ agonist. It was shown in vivo activity in a spinal nerve ligation model of neuropathic pain.²⁴²

.....Adolor Corporation discovered a novel chemical series of pyridine-based potent and selective CB₂ agonists by replacement of the phenyl ring in their previous (morpholinomethyl)aniline carboxamide cannabinoid receptor ligands with a pyridine ring. Compound **9** (Figure 3), that is 2,2-dimethyl-N-(5-methyl-4-(morpholinomethyl)pyridin-2-yl)butanamide showed good affinity at the CB₂ receptor (K_i = 24 nM), 160-fold selectivity versus CB₁ and moderate metabolic stability in rat

and human liver microsomes. This compound displayed antiallodynic activity after oral administration in a rat model of neuropathic pain.²⁴³

Hoffmann-La Roche disclosed in a patent publication the pyridine-oxadiazoles as CB₂ agonists with EC₅₀ below 0.05 μM and selectivity versus CB₁ in the corresponding assay of at least 500 fold. within a large number of compounds the most representative are the 4-cyclopropylmethoxypyridines exemplified by cyclopropyl-4-(cyclopropylmethoxy)pyridin-2-yl]-3-methyl-1,2,4-oxadiazole **10** (Figure 3) that showed EC₅₀ value of 25 nM for the CB₂ receptor.²⁴⁴

*Preparation of 6-(2,4-dichlorophenylamino)-4-cyclopropyl-N-(tetrahydro-2H-pyran-4-yl)methylpyridine-3-carboxamide (2).*²³⁷ The addition of cyclopropylmagnesium bromide to the 6-chloronicotinic acid, followed by reaction of this with 2,4-dichloroaniline by standard amide formation gave the desired final compound **2** (Scheme 1).



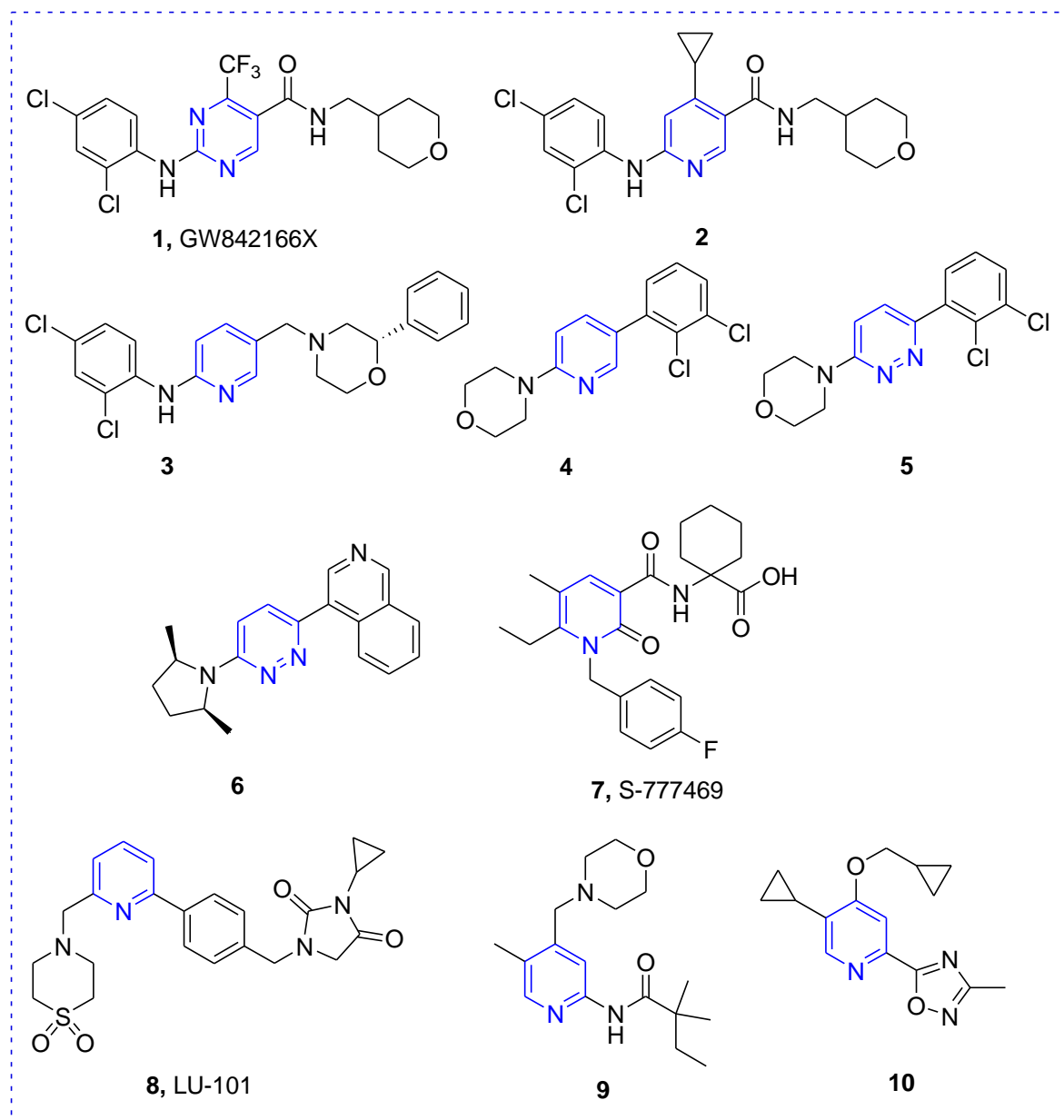
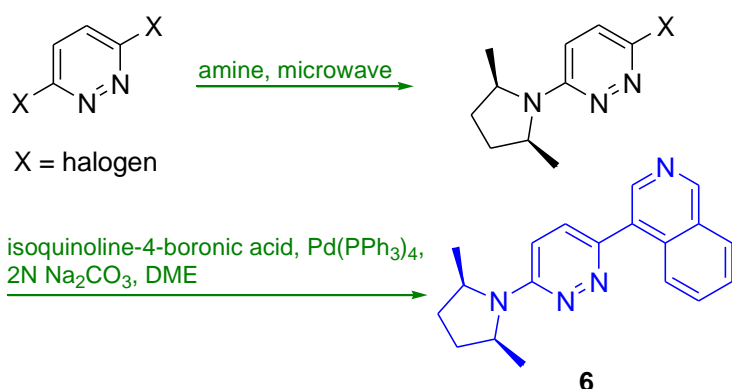


Figure 3. Pyridine and pyridazine derivatives.

*Preparation of 4-(6-((2R,5S)-2,5-dimethylpyrrolidin-1-yl)pyridazin-3-yl)isoquinoline (6).*²⁴⁰

Pyridazine analogue 6 was synthesized from the commercially available 3,6-dihalogen pyridazine according to Scheme 2. Synthesis proceeded via a palladium catalyzed cross coupling combined with an aromatic halide displacement. Halide displacement with the amine was carried out at high temperature in a microwave reactor.

Scheme 2. Synthesis of pyridazine 6.



7.2. Sulfamoyl Benzamide

Starting from a screening hit **11** (Figure 4) with modest affinity for the CB₂ receptor; Adolor Corporation identified sulfamoyl benzamides as a novel series of CB ligands. Several benzamides were prepared by introducing large lipophilic amide substituents, and increased selectivity for the CB₂ receptor was achieved by the introduction of an *S*-fenchyl residue such as in compound **12** (Figure 4). This had a 120-fold functional selectivity for the CB₂ receptor and produced robust antiallodynic activity in rodent models of postoperative pain and neuropathic pain without the habitual cannabinergic side effects. Small changes in the sulfonamide part of the molecule produced a switch from full agonist to inverse agonist.²⁴⁵

Further SAR studies and the optimization of the amide linkage led to the reverse amide derivative **13** with superior selectivity over the CB₁ receptor while retaining good affinity for CB₂. The best compound of this study, the tetramethylcyclopropy derivative **14** (Figure 4, EC₅₀ = 11 nM, E_{max} = 84%), displayed poor metabolic stability in rat pain model.²⁴⁶ This compound exhibited robust antiallodynic activity in a rodent pain model when administered intraperitoneally. Efficacy after oral administration was observed only when pretreated with aminobenzotriazole (ABT), a cytochrome P450 suicide inhibitor, before administration. The isosteric replacement of the amide functionality

of **11** with various heterocycles (thiazoles, oxadiazoles, pyrazoles, imidazoles, triazoles and tetrazoles) did not significantly improve affinity or selectivity for the CB₂ receptor in comparison to the lead compound **11**. Further efforts to improve the in vitro metabolic stability profile in the sulfonamide series led to the identification of *N*-[3,4-dimethyl-5-(morpholin-4-ylsulfonyl)phenyl]-2,2-dimethyl butanamide **15** (Figure 4, Scheme 3) that displayed the best overall in vitro profile and demonstrated robust efficacy in an animal model of post-operative pain.²⁴⁷

Preparation of N-[3,4-dimethyl-5-(morpholin-4-ylsulfonyl)phenyl]-2,2-dimethyl butanamide (**15**).²⁴⁷ Compound **15** was prepared according to Scheme 3. Chlorosulfonylation of commercially available substituted nitrobenzene was achieved using chlorosulfonic acid. Condensation of the corresponding sulfonyl chlorides with morpholine followed by reduction of the nitro group yielded the substituted aniline derivative. Classical amide formation led to the target compound **15**.

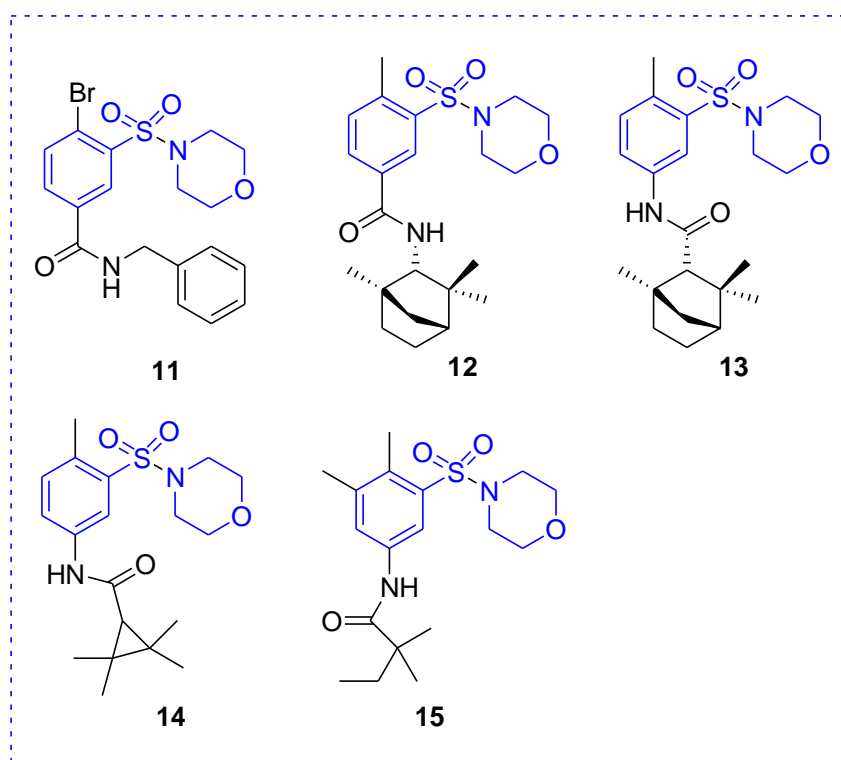
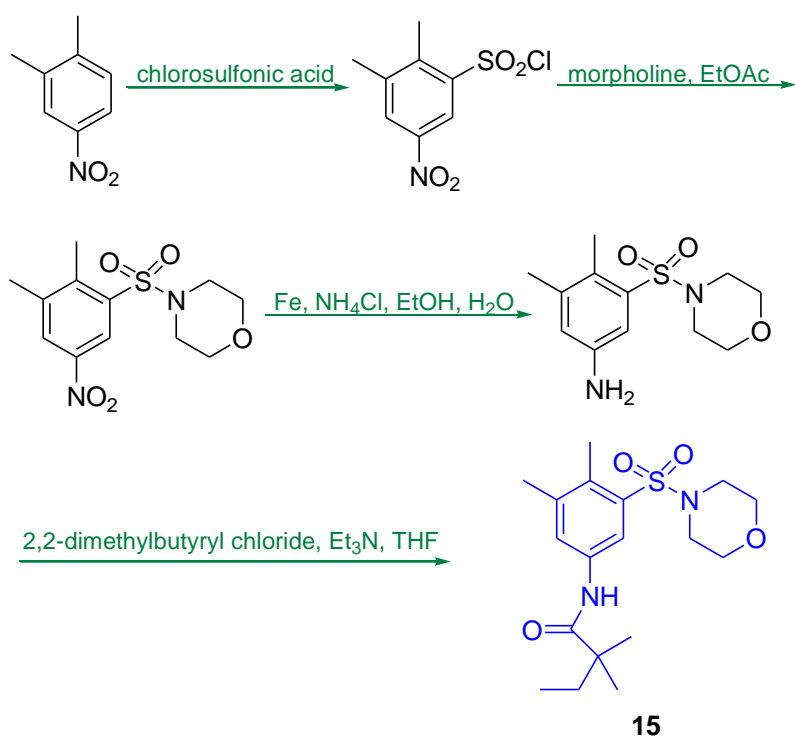


Figure 4. Sulfamoyl benzamide derivatives.

Scheme 3. Synthesis of Sulfamoyl benzamide 15.



7.3. Heterocyclic Ylidene

Taisho Pharmaceuticals published the five-membered heterocyclic cores as CB₂ selective agonists. The synthesis and SARs of thiazole derivatives as potent CB₂ agonist showed that the functional groups at 3- and 5-position in the thiazole ring and the aromatic group in the amide deeply influence the affinity and selectivity for the CB₂ receptor. Thiazolylidene compounds such as **16** (Figure 5) with a bulky functional group like trifluoromethyl exhibited much higher CB₂ affinities (hCB₂ IC₅₀ = 13 nM, 270 fold selectivity)²⁴⁸

Modifying the heteroring of **16** to improve binding affinity and selectivity for the CB₂ receptor led to the preparation of five-membered heterocyclic derivatives **17-21** (Figure 5). Thiadiazole **17** exhibited lower affinity for the CB₂ receptor than the thiazole derivative. Oxazolylidene **18**, isothiazol-ylidene **19**, and isoxazolylidene **20** had good affinity for the CB₂ receptor (**18**: IC₅₀ = 24 nM, **19**: IC₅₀ = 9.5 nM, **20**: IC₅₀ = 46 nM). Incorporation of the pyrazole reduced the overall

lipophilicity of this chemical series and led to an improvement in aqueous solubility. These studies led to compound **21** ($IC_{50} = 51$ nM) and to the *N*-(3-*tert*-butyl-1-(cyclopropylmethyl)-1,2-dihydro-2-methylpyrazol-5-ylidene)-2-fluoro-3-(trifluoromethyl)benzamide **22** (CBS0550, Figure 5, Scheme 4), which exhibited particularly high affinity and selectivity for the CB₂ receptor ($IC_{50} = 2.9$ nM, $E_{max} = 85\%$, $SI = 1400$). This compound had good solubility in water, exhibited good metabolic stability in human and rat liver microsomes. A dose dependent (10 and 30 mg/kg) compound **22** significantly reversed mechanical hyperalgesia in the Randall–Selitto model of inflammatory pain in rats.²⁴⁹

*Preparation of (14E)-N-(3-tert-butyl-1-(cyclopropylmethyl)-1,2-dihydro-2-methylpyrazol-5-ylidene)-2-fluoro-3-(trifluoromethyl)benzamide (22)*²⁴⁹ According to Scheme 4, cyclopropylmethyl alcohol was treated with methanesulfonyl chloride (MsCl) and triethylamine (Et₃N) to yield mesylate intermediate. This was transformed into alkylhydrazine by treatment with hydrazine. Cyclization of 1-(cyclopropylmethyl)hydrazine by treatment with pivaloylacetone nitrile yielded the 5-aminopyrazole. The latter was treated with trifluoroacetic anhydride and pyridine to obtain *N*-(3-*tert*-butyl-1-(cyclopropylmethyl)-1*H*-pyrazol-5-yl)-2,2,2-trifluoroacetamide. The 1-position of the pyrazole ring was alkylated, followed by deprotection under basic conditions, producing the imine intermediate which was easily transformed into pyrazole final compound **22** in one step by treatment with the corresponding aryl carbonyl chloride and Et₃N.

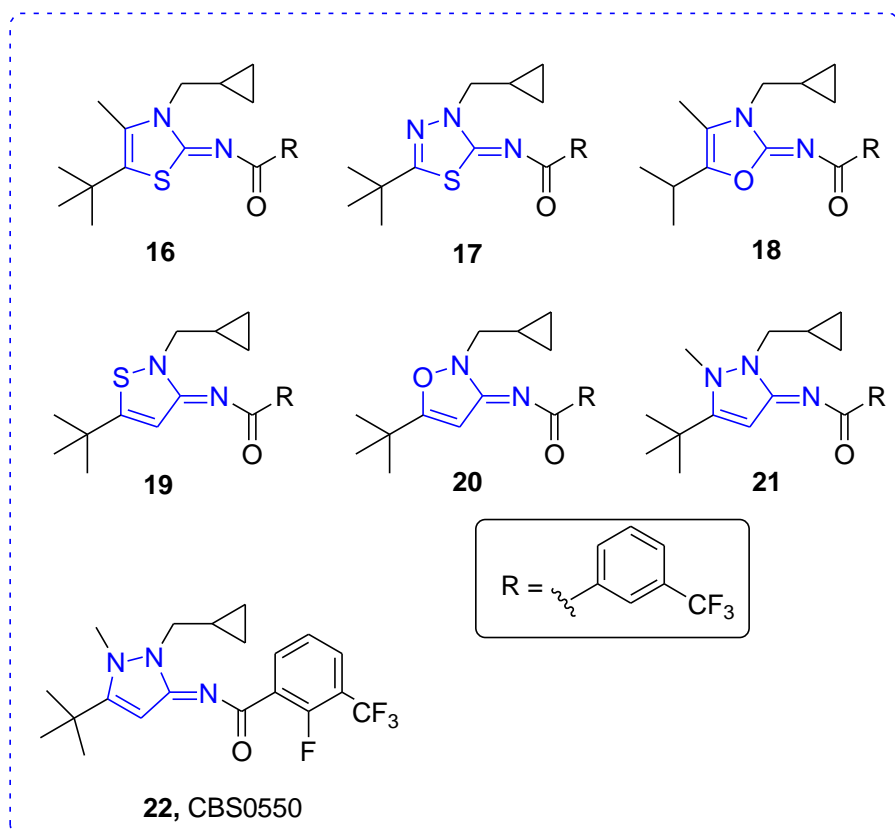
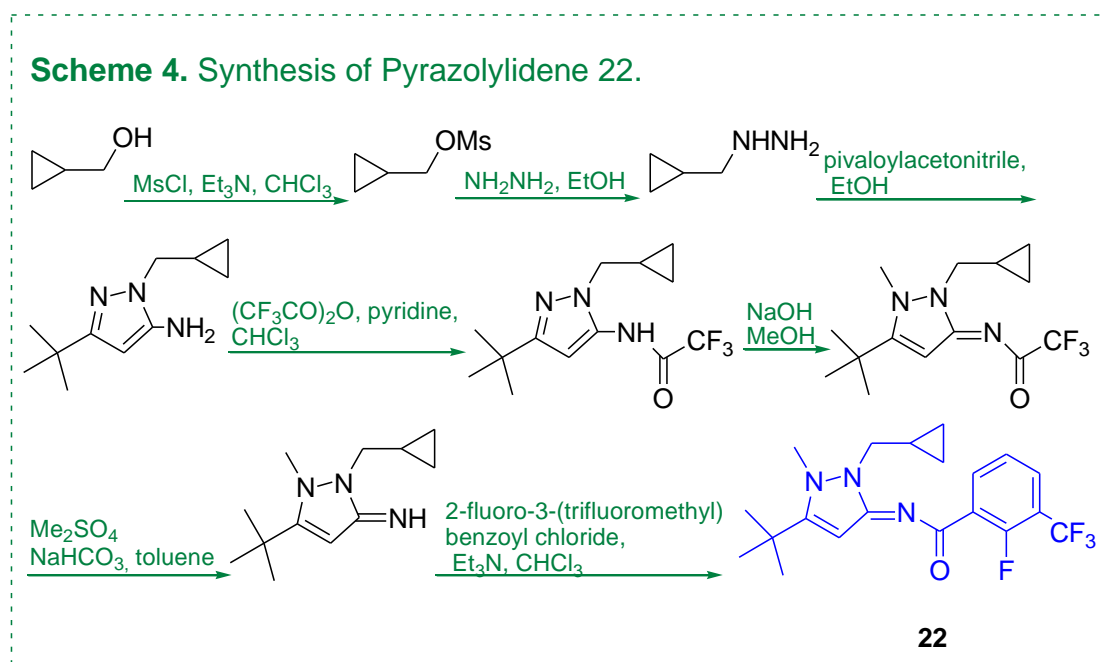


Figure 5. Heterocyclic ylidene derivatives.



7.4. Oxadiazole, α -Amidosulfone

The five-membered oxadiazole core as a novel class of potent and selective CB₂ agonists by a targeted library screening and subsequent hit assessment was identified.²⁵⁰ The SARs investigation into this group of compounds showed that the oxadiazole core was fundamental, and substitutions at the 2- and 4- positions of the phenyl ring joined to the oxadiazole were important for desired CB₂ activity. The amino quinoline derivative **23** (Figure 6, Scheme 5) was a potent and selective CB₂ agonist (EC₅₀ = 10 nM, Emax = 104%), which displayed an excellent pharmacokinetic profile with oral bioavailability in rats.²⁵⁰ Structural modifications in the central portion of the *N*-arylamide oxadiazole scaffold led to the identification of *N*-arylpiperidine oxadiazoles as conformationally constrained analogs, such as compound **24** (Figure 6), that offered improved stability with comparable potency and selectivity (EC₅₀ = 80 nM, Emax = 101%).²⁵¹ The strictly correlated α -amidosulfones were found to be CB₂ potent and selective agonists. The effect of the amide substituent on CB₂ activity and selectivity was explored. 3,4-Disubstituted phenyl ring, such as in 2-(4-chlorophenylsulfonyl)-*N*-(4-chloro-3-(trifluoromethyl)phenyl)-2-methylpropanamide **25** (Figure 4), enhanced the potency and selectivity for CB₂ receptor (EC₅₀ = 20 nM, Emax = 102%). For the five-membered heterocycles it was found that rings with *tert*-butyl group induced activity and selectivity. This series is represented by 2-(4-chlorophenylsulfonyl)-*N*-(3-*tert*-butyl-1-methyl-1*H*-pyrazol-5-yl)-2-methylpropanamide **26** (Figure 6) (EC₅₀ = 20 nM, Emax = 104%). α -Amidosulfones behaved as selective CB₂ full agonists and showed high functional and cellular activity on CB₂ receptors.²⁵²

*Preparation of 6 3-(4-(3-(2-chloro-4-fluorophenyl)-1,2,4-oxadiazol-5-yl)piperidin-1-yl)quinoline (24).*²⁵¹ Condensation of 2-chloro-4-fluorobenzonitrile and hydroxylamine afforded *N*-hydroxybenzamidine, which was condensed with 1-(*tert*-butoxycarbonyl)piperidine-4-carboxylic acid to afford the 4-(3-(2-chloro-4-fluorophenyl)-1,2,4-oxadiazol-5-yl)piperidine. The piperidine

intermediate was coupled with 3-bromoquinoline using a Buchwald-Hartwig reaction to afford 24.

(Scheme 5)

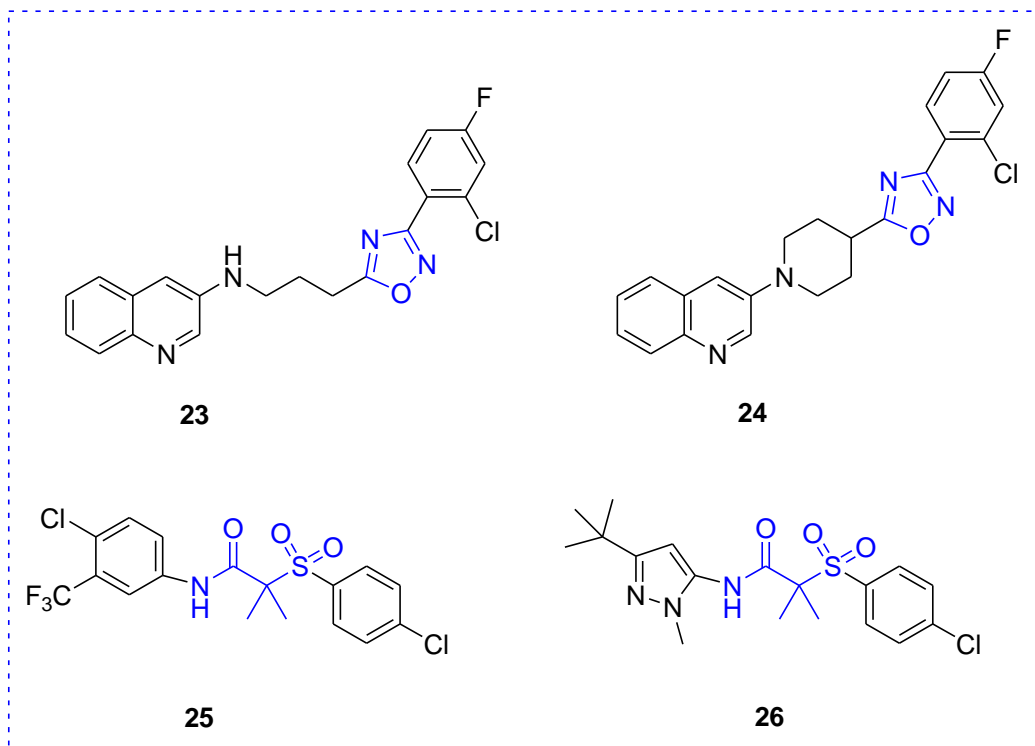
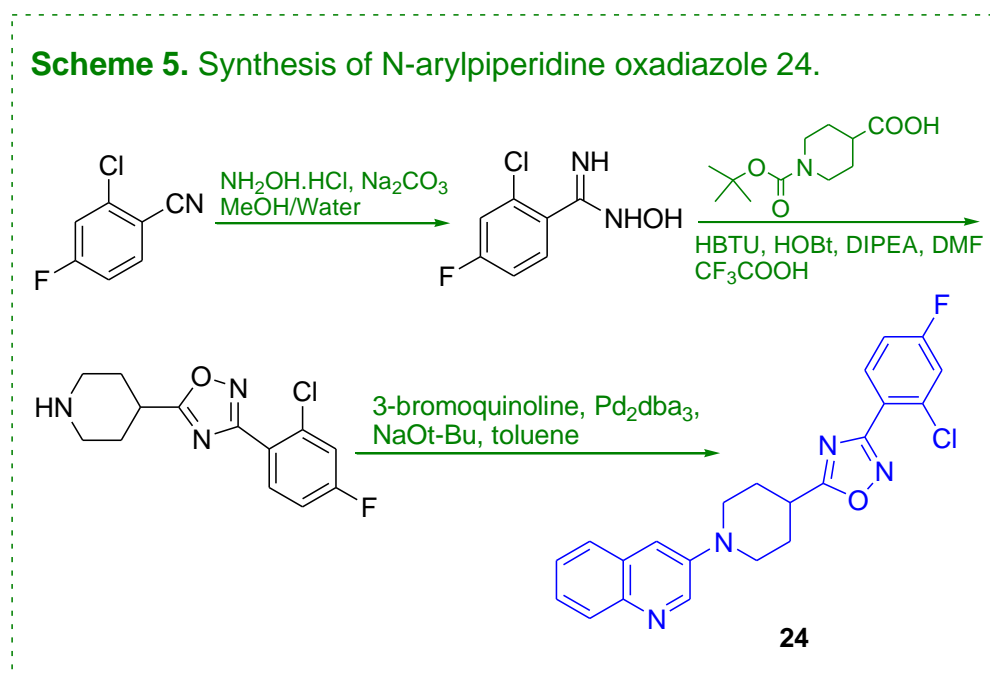


Figure 6. Oxadiazole and α -Amidosulfone derivatives.



7.5. Imidazole

Merck Research Laboratory published 2,4-diphenyl-1*H*-imidazole analogues as CB₂ receptor ligands. Compound **27** (Figure 7, Scheme 6), one of the most active compounds in this series (hCB₂ EC₅₀ = 8 nM) displayed high selectivity over hCB₁, but did not possess high plasma exposure in rats. The replacement of morpholine side chain with the 4-fluoro or 4,4-di-fluoro piperidinyl moiety yielded compounds **28** and **29**, respectively, as potent CB₂ full agonists (**28**: hCB₂ EC₅₀ = 9 nM, **29**: 5 nM). Replacement of the 3-trifluoromethyl group of phenyl ring with 2,4-dichlorine atoms (**30**: hCB₂ EC₅₀ = 10 nM) did not affect the potency compared to that of **28**.²⁵³

*Preparation of 4-(3-(5-(3-(trifluoromethyl)phenyl)-1*H*-imidazol-2-yl)benzyl)morpholine (27).*²⁵³

The synthetic route to the 2,4-diphenyl-1*H*-imidazole **27** is summarized in Scheme 6. The commercially available 1-(3-(trifluoromethyl)phenyl)ethanone was brominated and then cyclized with formamide to form 4-phenylimidazole. Consequent N-protection with 2-(trimethylsilyl)ethoxymethyl (SEM) chloride and iodination using iodine were achieved under basic conditions to afford 1-((2-(trimethylsilyl)ethoxy)methyl)-4-(3-(trifluoromethyl)phenyl)-2-iodo-1*H*-imidazole. The iodo displacement with 3-phenylaldehyde was conducted under Suzuki coupling condition using Pd(dppf)₂Cl₂ as the catalyst. Reductive amination and final removal of the SEM yielded the target compounds **27**.

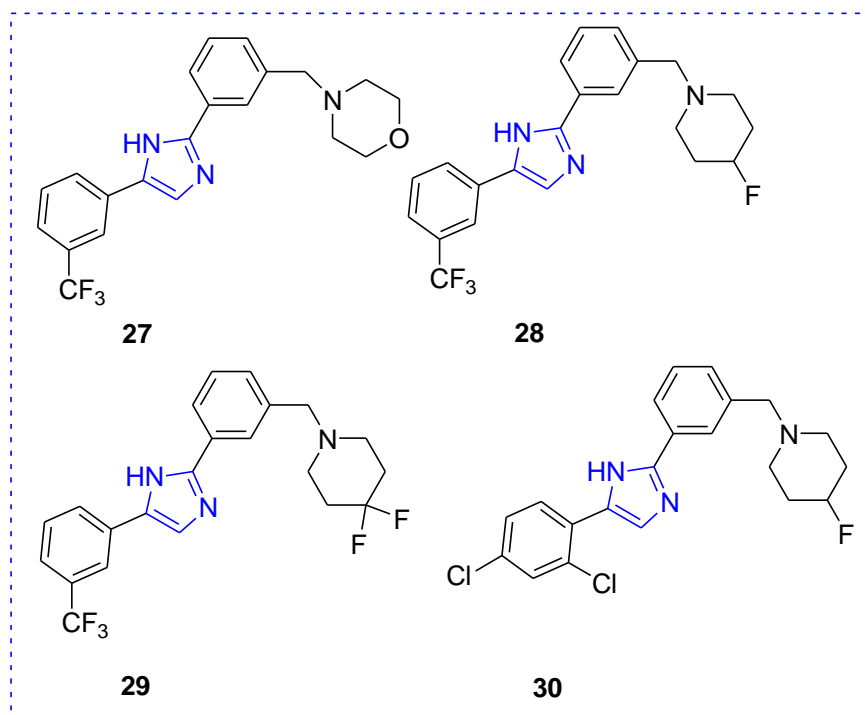
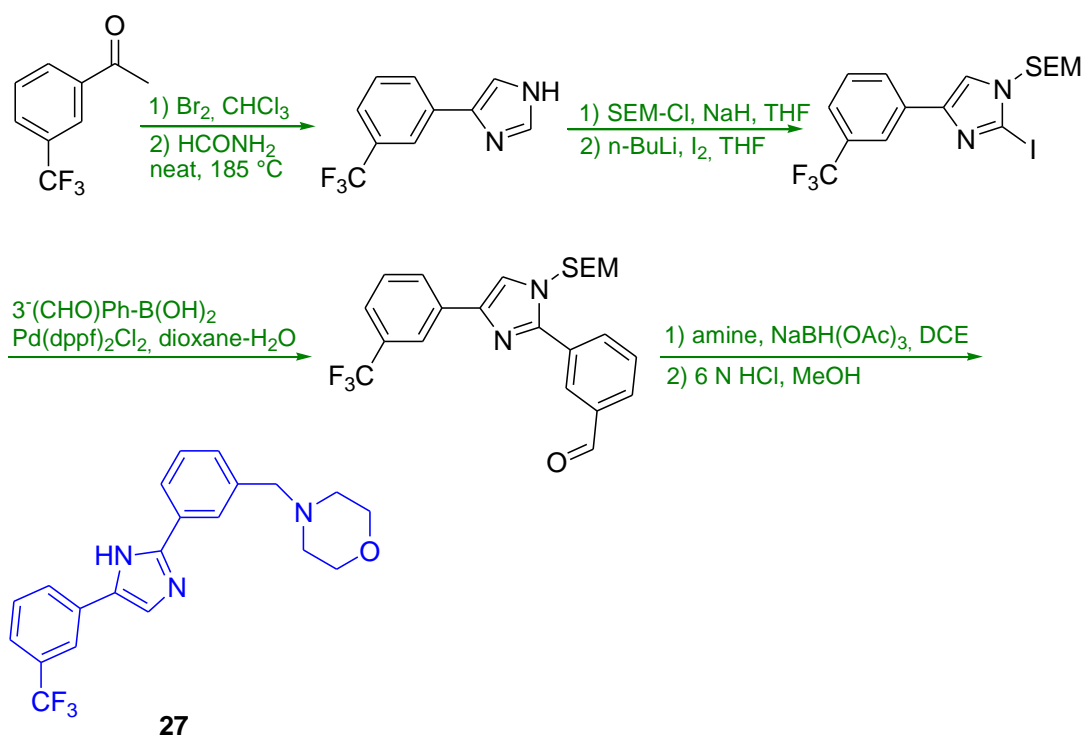


Figure 7. Imidazole derivatives.

Scheme 6. Synthesis of Imidazole 27.



7.6. Benzoimidazole

CB₂ agonists based on benzimidazole template have been reported by Page et al. from AstraZeneca. One of the first, discovered as part of an HTS campaign, was 2-(4-ethoxybenzyl)-1-(2-(dimethylamino)ethyl)-*N,N*-diethyl-1*H*-benzo[*d*]imidazole-5-carboxamide **31** (Figure 8). SAR studies around compound **31** revealed that on the N-1 position various alkyl and aromatic groups were tolerated. In this series, with *N*-cyclopropylmethyl (**32**: hCB₂ K_i = 4.1, hCB₁ K_i = > 5000 nM) and *N*-isopentyl (**33**: hCB₂ K_i = 4.5 hCB₁ K_i = > 5000 nM) an optimal potency and selectivity was obtained.²⁵⁴ The diethyl amide modifications were all well tolerated demonstrating probably the occurrence of a large binding pocket at the receptor site. Reducing the size of the group to a secondary amide showed about a 10-fold decrease in CB₂ binding affinity, having only a primary amide on the left-hand side resulted in a total loss of activity.²⁵⁴ Further studies by Pfizer on the benzimidazole scaffold led to the preparation of a large number of sulfonyl derivatives with CB₂ receptor agonistic activity. Incorporation of neopentyl chain into the 2-position together with introduction of a sulfonyl group into the 5-position led to one of the best compound of series 1-(1-(cyclopropylmethyl)-2-neopentyl-1*H*-benzo[*d*]imidazol-5-ylsulfonyl)cyclopropanecarboxamide (**34**, Figure 8), a selective and potent full agonist of CB₂ receptor. From lead compound **34**, a structurally similar series based on saturation of the benzene ring was designed. The alkyl, carbamate, urea, amide and sulfonamide derivatives of the tetrahydro-1*H*-imidazo[4,5-*c*]pyridin-5-ium scaffold were prepared. Lipophilic Efficiency (LipE) was introduced as a parameter that combines both potency and lipophilicity as a helpful instrument for design of potent and metabolically stable CB₂ agonists. The sulfonamide derivatives such proved to be better than the other functionalities being the most potent full agonist at the hCB₂ receptor. Among this series, the isoxazole **35** (Figure 8) displayed agonist activity at 10 μM on the hCB₁ receptor (EC₅₀ **35** = 7.5 μM).²⁵⁵ In a related effort, RaQualia Pharma claimed *N*-substituted saturated heterocyclic sulfone compounds with CB₂ agonistic activity. One of the representative derivative is 3-(1-(2-

(dimethylamino)ethyl)-2-neopentyl-1*H*-benzo[*d*]imidazol-5-ylsulfonyl)azetidine-carboxamide **36** (Figure 8).²⁵⁶

Subsequently, the structural modifications to improve Human Liver Microsomes (HLM) stability and membrane permeability produced compound **37** (RQ-00202730, Figure 8), which bears a hydroxyethyl group as a side chain on the azetidine ring. RQ-00202730 demonstrated the best overall profile in terms of potency, selectivity ($EC_{50} = 19$ nM, SI >1300-fold) over the CB₁ receptor showing a dose dependent analgesic effect on TNBS-induced visceral hypersensitivity in rats by oral administration.²⁵⁷ Following investigations focused on the benzoimidazole scaffold improved the pharmacokinetic profile of compound **38** by removing polar functionality at the 5-position to improve CNS penetration. Optimization of the benzimidazole substituents of **38** led to the identification of compound **39** (Figure 8), a potent CB₂ full agonist ($EC_{50} = 2.7$ nM) with excellent selectivity over the CB₁ receptor (>3000 fold). Compound **39** demonstrated good CNS penetration in rat. Further optimization led to the discovery of the fully CNS penetrant CB₂ agonist **40** (Figure 8) with reduced human *Ether-à-go-go*-Related Gene (hERG) activity.²⁵⁸

The benzimidazole CB₂ receptor agonists were synthesized and developed by Janssen Pharmaceuticals and the structure–activity relationship was explored. The size of the substituent on the 2-position determined the level of agonism, ranging from inverse to partial or full agonism, which was more pronounced for the rat CB₂ receptor. The benzoimidazole cannabinoid agonists bearing a substituted aryl group, in particular, the pyridyl sulfone, such as compound 2-*tert*-butyl-5-(2-ethoxypyridin-4-sulfonyl)-1-tetrahydropyran-4-ylmethyl)-1*H*-benzimidazole **41** (Figure 8), had excellent binding affinity and selectivity for CB₂ (h $EC_{50} = 0.3$ nM, hCB₁ $EC_{50} = 1280$ nM).²⁵⁹ Pyridyl-sulfone **40** was reported to be 43% orally bioavailable in rat and to have a low brain to plasma ratio.²⁶⁰ Further optimization of this series led to formation of peripherally acting CB₂ agonists, such as **42** (Figure 8, Scheme 7) and **43** (Figure 8). While both compounds were not active in acute pain models, the less selective compound **41** (hCB₂ $EC_{50} = 8.4$ nM, hCB₁ $EC_{50} >10000$

nM) displayed activity in a chronic model of neuropathic pain.²⁵⁹ In a related effort, *N*-{1-(cyclohexylmethyl)-2-[(5-ethoxypyridin-2-yl)methyl]-1*H*-benzimidazol-5-yl}-*N*-methylthiophene-2-sulfonamide (**44**, AZ-11713908, Figure 8) was produced by AstraZeneca as a potent CB₁ agonist and as a partial CB₂ agonist. It displayed poor blood-brain barrier penetration and was consequently a peripherally selective compound exhibiting analgesic effects in inflammatory and neuropathic pain models.²⁶¹ Subsequently, researchers in Merck designed a series of benzimidazole CB₂ agonists from imidazopyridine scaffold.²⁶² Evaluation of the SAR revealed that introduction of the amide group at the 2-position, as well as the *N*-alkyl imidazoles, provided more potent CB₂ agonism when compared to *N*-unsubstituted benzimidazoles. The lipophilic adamantyl amide gave a very potent CB₂ agonist, but its selectivity over CB₁ was poor. The most potent compound in this series was obtained by introduction of an aliphatic amide bearing hydroxyl group (compound **45**: hCB₂ EC₅₀ = 6.3 nM, hCB₁ EC₅₀ >17,000, Emax 75%, Figure 8). Although the new structural class provided CB₂-selective compounds, the pharmacokinetic profiles were moderate.²⁶³

*Preparation of 1-(4-((2-tert-butyl-5-(ethylsulfonyl)-1*H*-benzo[d]imidazol-1-yl)methyl)piperidin-1-yl)ethanone (42).*²⁵⁹ Starting from 5-chloro-2-nitro-aniline, 4-methoxybenzylthiol (PMBSH) was introduced as a protecting group via nucleophilic aromatic substitution and subsequent reaction with pivaloylchloride gave *N*-(5-(4-methoxybenzylthio)-2-nitrophenyl)pivalamide. Reduction of the nitro group was followed by reductive amination with piperidine-4-carbaldehyde and subsequent condensation under acidic conditions resulted in debocylation, which allowed for subsequent introduction of acetyl group. Deprotection of the *p*-methoxybenzyl group with trifluoroacetic acid (TFA), ethyl introduction via nucleophilic substitution, and finally, the oxidization with *m*-chloroperoxybenzoic acid yielded the sulfone 42. (Scheme 7)

*Preparation of 1-ethyl-*N*-(2,3-dihydroxypropyl)-4-phenyl-1*H*-benzo[d]imidazole-2-carboxamide 45.*²⁶³ Phenyl boronic acid was coupled to 3-chloro-2-nitroaniline which was then reduced with tin(II)chloride to give *o*-diamine intermediate. Treatment with methyl-2,2,2-trichloroacetimidate,

and ethanolysis gave benzimidazole-2-ethyl ester. Alkylation by iodoethane produced exclusively one alkylated product, which was then hydrolyzed and subsequent peptide coupling conditions yielded the desired amide 45. (Scheme 8)

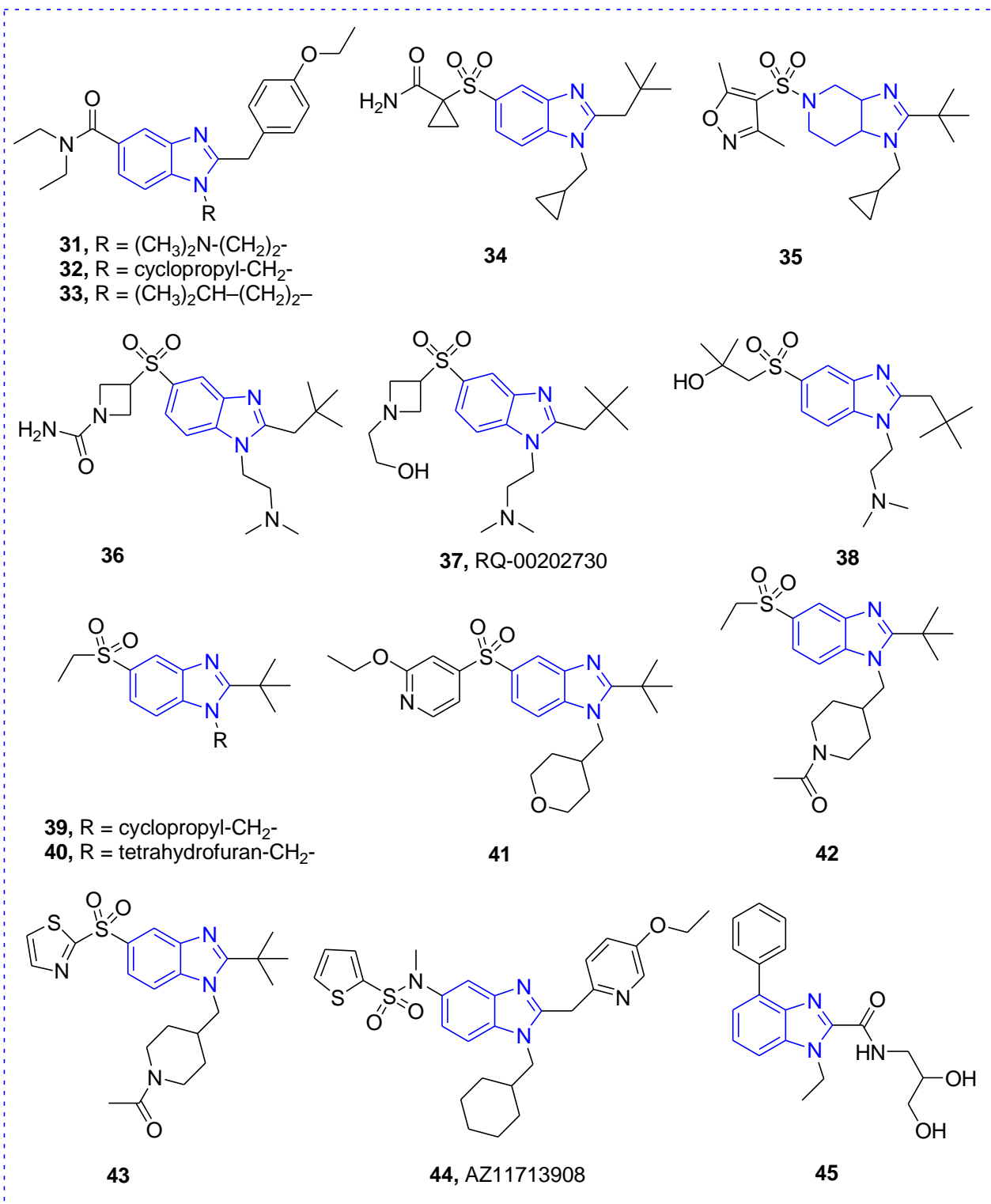
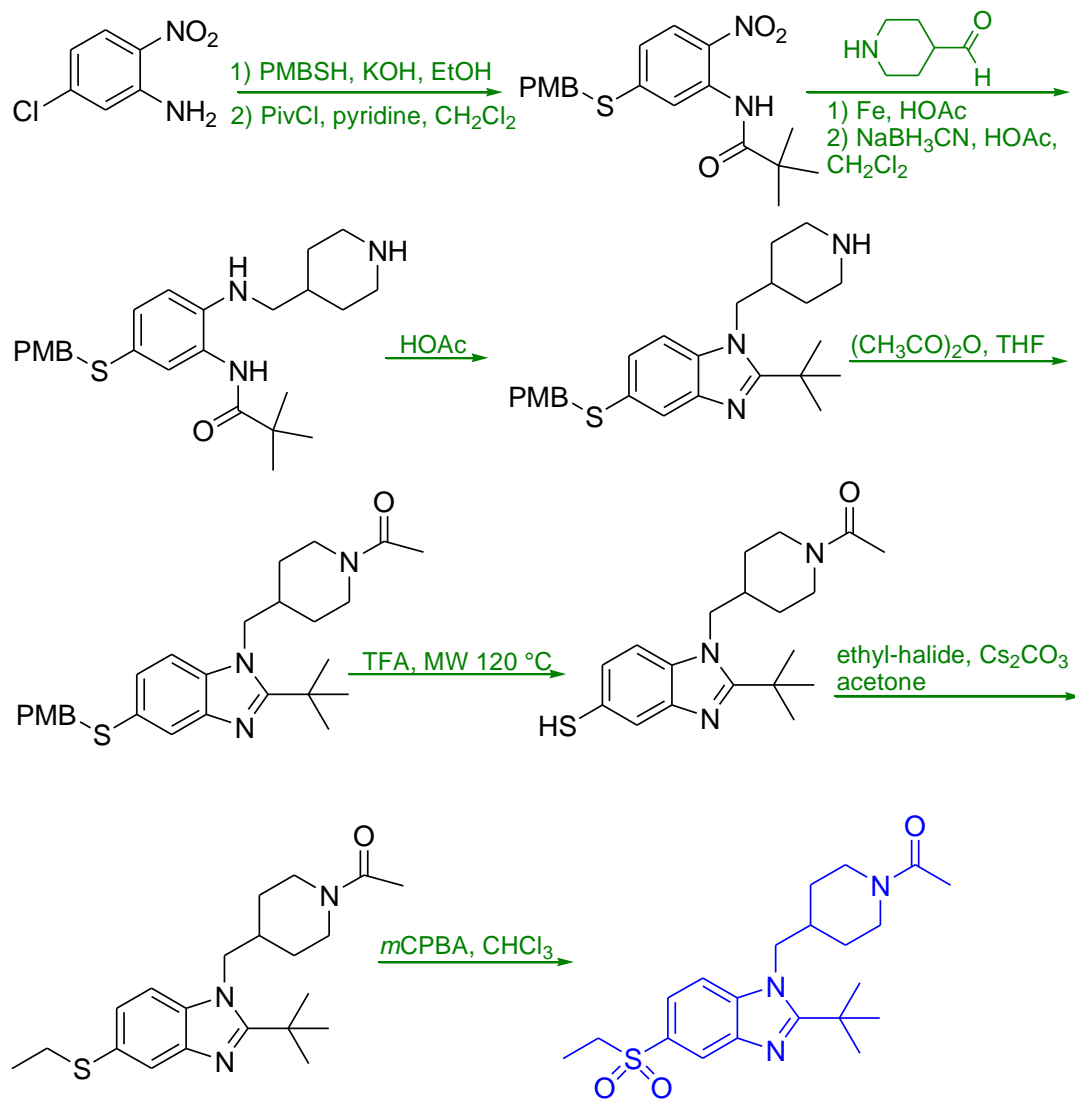
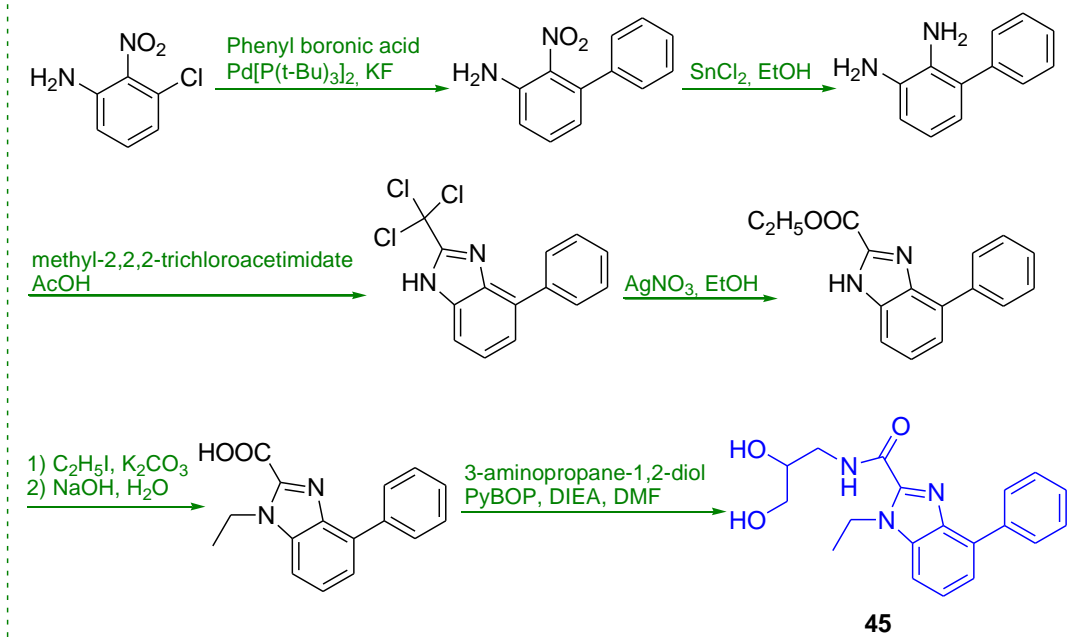


Figure 8. Benzimidazole derivatives.

Scheme 7. Synthesis of 5-Ethylsulfonyl benzoimidazole 42.



Scheme 8. Synthesis of 4-Phenyl benzoimidazole 45.



7.7. Imidazopyridine

Imidazopyridine CB₂ agonists have been described as closely related scaffolds of indole based CB₂-selective ligands. Exploration of SAR in this chemical series by variation of substituents on both carbon positions of the imidazole ring of the core produced CB₂ agonists. Compounds N-((1-(hydroxymethyl)cyclopentyl)methyl)-3-morpholino-*H*-imidazo[1,5-*a*]pyridine-1-carboxamide **46** (Figure 9) (hCB₂ IC₅₀ = 33 nM, hCB₁ IC₅₀ > 17000 nM, Emax 99%) and 2,3-dichlorophenyl(1-(morpholinomethyl)-*H*-imidazo[1,5-*a*]pyridin-3-yl)methanone **47** (hCB₂ IC₅₀ = 5 nM, Emax 94%; hCB₁ IC₅₀ = 2565 nM, Emax 66%) (Figure 9, Scheme 9), which were completely selective for CB₂ versus CB₁, are representatives of this series. In vivo evaluation of these compounds indicates a significant impact of the degree of selectivity for CB₂ on analgesic effects.²⁶⁴ A related series of 3-arylimidazopyridine analogues was reported, wherein 3-(3-(trifluoromethyl)phenyl)-1-(4,4-difluoropiperidin-1-yl)methyl)-*H*-imidazo[1,5-*a*]pyridine **48** (Figure 9) was the most potent CB₂ agonist disclosed.²⁶⁵

Preparation of (2,3-dichlorophenyl)(1-(morpholinomethyl)H-imidazo[1,5-a]pyridin-3-yl)methanone (47).²⁶⁶ As depicted in Scheme 9, condensation of 2-aminomethyl pyridine with ethyl oxalyl chloride provided the ester intermediate. Formylation with the Vilsmeier reaction gave imidazopyridine aldehyde which, by means of reductive amination, yielded the ethyl 1-(morpholinomethyl)H-imidazo[1,5-a]pyridine-3-carboxylate, that was then transformed into Weinreb amide intermediate. The hydrolysis and Grignard reaction yielded the final compound 47.

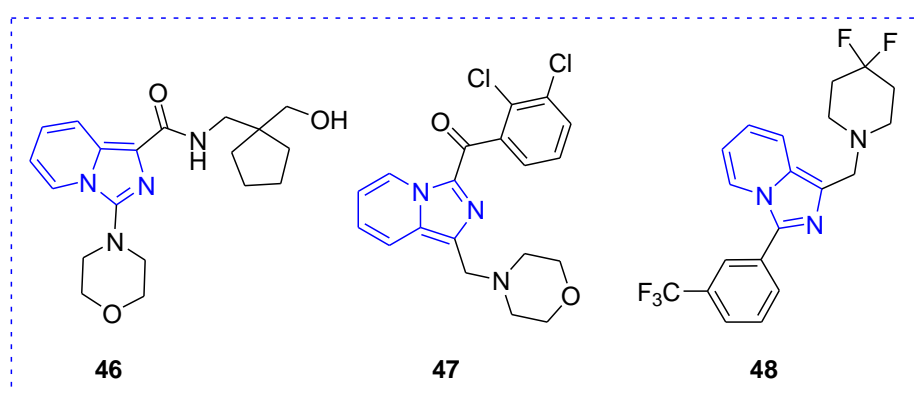
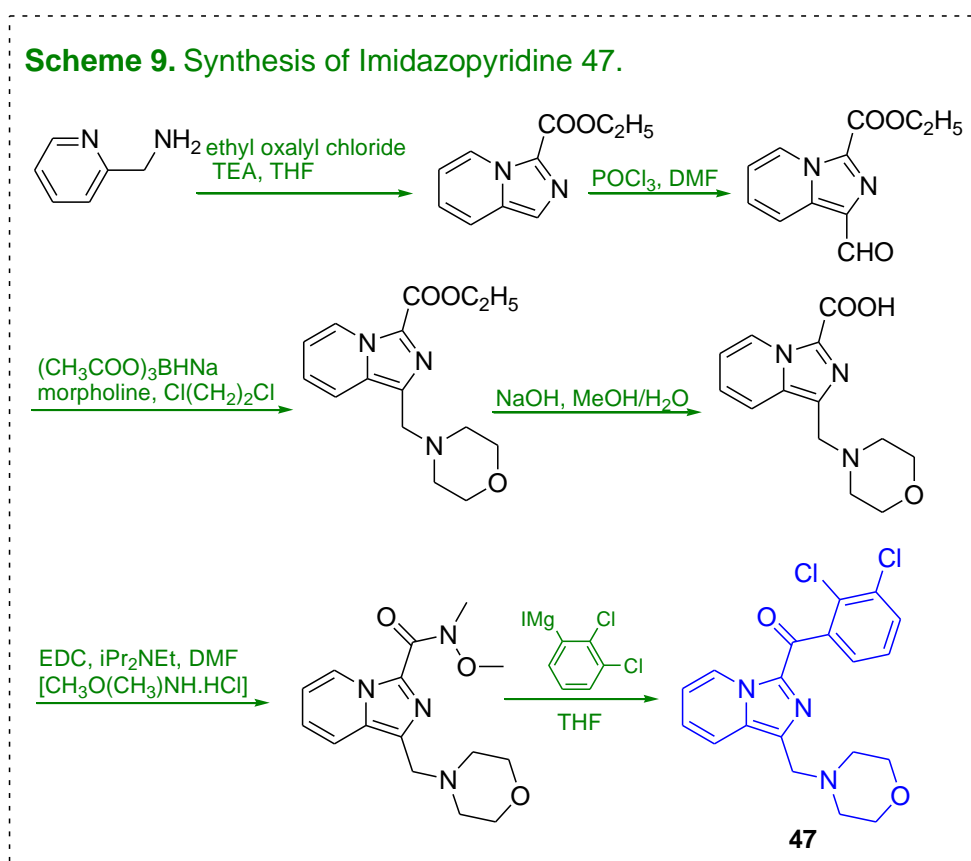


Figure 9. Imidazopyridine derivatives.



7.8. Purine

Lilly Research Laboratories discovered the purine derivatives as CB₂ receptor agonists based on the thieno[2,3-*d*]pyrimidine scaffold **49** (hCB₂ EC₅₀ = 25 nM, SI = 104, Figure 10) with moderate selectivity and poor in vitro metabolic profile. The selectivity for CB₂ and metabolic profile were further optimized by varying the substituent at the 9-position of the purine core yielding a number of potent CB₂ agonists including compounds (*R*)-8-(2-chlorophenyl)-9-((*S*)-tetrahydrofuran-3-yl)-2-methyl-6-(4-methylpiperazin-1-yl)-9*H*-purine **50** (hCB₂ EC₅₀ = 8.8 nM, SI > 1130, Figure 10) and the 9-tetrahydro-2*H*-pyran derivative **51** (Figure 10, Scheme 10), which possessed good biopharmaceutical properties and potent oral activity in a preclinical model of joint pain.^{267,268}

*Preparation of 8-(2-chlorophenyl)-9-(tetrahydro-2*H*-pyran-4-yl)-2-methyl-6-(4-methylpiperazin-1-yl)-9*H*-purine hydrochloride (51).*²⁶⁸ The starting material 5-amino-4,6-dichloro-2-methylpyrimidine was reacted with tetrahydro-2*H*-pyran-4-amine to afford the monoamino intermediate (Scheme 10). Subsequent reaction with 2-chlorobenzaldehyde was followed by cyclization by means of FeCl₃-SiO₂. The subsequent oxidation by 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) ensured the complete conversion to the desired purine compound. Successive reaction with 1-methylpiperazine gave the desired product which was then converted into the corresponding HCl salt 51.

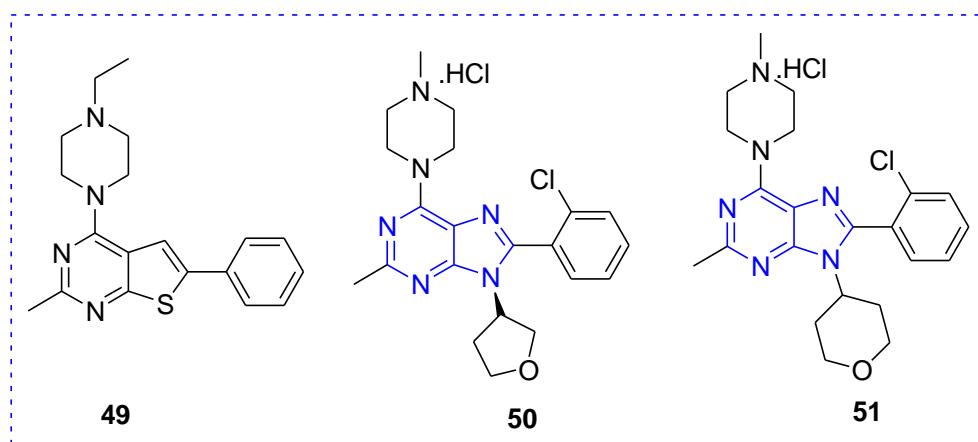
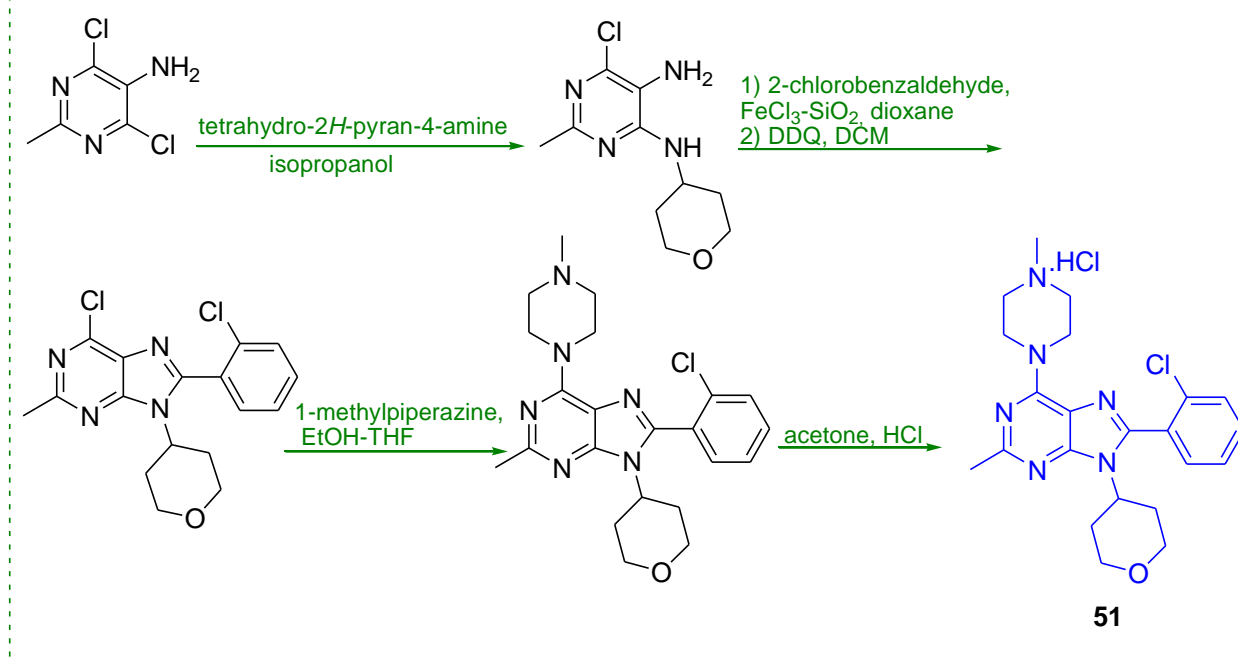


Figure 10. Purine derivatives.

Scheme 10. Synthesis of Purine 51.



7.9. Quinolone, Naphthyridine

The CB_2 receptor agonists also include 4-oxo-1,8-naphthyridine-3-carboxamide and 4-oxo-1,4-dihydroquinoline-3-carboxamide derivatives endowed with high affinity and selectivity toward CB_2 receptors.^{269,270} Some of these analogues were demonstrated to act as agonists or inverse agonists in functional activity assays, depending on the nature of the substituents on the different positions of the heterocyclic scaffold. A closely related series of 1,8-naphthyridin-2(1*H*)-on-3-carboxamide analogues were described in subsequent publications.²⁷¹ In the series, the *cis*-4-methylcyclohexylamido analogue **52** (Figure 11) exhibited the highest affinity, with K_i values of 0.7 nM. Very recently, a number of additional derivatives in which the same central scaffold was variously functionalized in position 1 or 6, have been reported.²⁷² The best results in terms of both CB_2 affinity and selectivity were obtained with the 6-methoxyphenyl analogue **53** (Figure 11, Scheme 11) showing high CB_2 affinity and selectivity (CB_2 K_i = 1.47 nM, CB_1 K_i > 10000 nM).

Within this series, the functional activity of compounds is controlled by the presence of the substituents at position C-6 of the naphthyridine scaffold; introduction of substituents in this position determined a functionality switch from agonist to antagonists/inverse agonists.²⁷²

For the new 1,8-naphthyridin-2(1*H*)-one-3-carboxamide derivatives, docking studies have shown that both the antagonists/inverse agonists and the agonists bind in the TMH2-3-6-7 regions of CB₂ receptor, suggesting that the difference between the pharmacology of these ligands depends on their ability/inability to block the Toggle Switch W6.48 (χ^1 g⁺ → trans) transition.²⁷²

The naphthyridine CB₂ agonist **54** (CB13, Figure 11) was shown to induce apoptosis through ceramide de novo synthesis in colon cancer cells.²⁷³ The closely related 4-quinolone-3-carboxamides CB₂ chemotype have also been described. The nature of substituents at positions N-1, C-6 or C-8 were studied, and one of the most potent CB₂ agonist was **55** (hCB₂ K_i = 6.3 nM, hCB₁ K_i = 1220 nM, Figure 11), which demonstrated significant analgesic effects in a mouse formalin test of acute peripheral and inflammatory pain.²⁷⁴

Introduction of 2-furyl in position 6 (analogue **56**, Figure 11) produced a significantly improved CB₂/CB₁ selectivity profile relative to **55**. It behaved as a potent CB₂ neutral antagonist in the formalin test of acute and inflammatory pain.²⁷⁵ The 8-substituted-4-quinolone-3-carboxamide derivatives were also investigated, the 8-methoxy derivative **57** (hCB₂ K_i = 0.6 nM, hCB₁ K_i > 10000 nM) (Figure 11) was assayed in vivo in the formalin test of analgesia in mice, behaved as an inverse agonist and showed antihyperalgesic effects.²⁷⁶

*Preparation of 1,2,3,4-tetrahydro-6-(4-methoxyphenyl)-N-(4-methylcyclohexyl)-1-(2-morpholinoethyl)-2,4-dioxo-1,8-naphthyridine-3-carboxamide (53).*²⁷² The synthetic route to obtain 1,8-naphthyridin-2(1*H*)-one-3-carboxamide **53** is outlined in Scheme 11. Aminonicotinaldehyde was treated with bromine in glacial acetic acid to obtain the corresponding 6-bromo derivative, which was refluxed with diethyl malonate and in the presence of piperidine in ethanol to afford ethyl 6-bromo-1,8-naphthyridin-2(1*H*)-one-3-carboxylate intermediate. The reaction of ethyl ester

with a *cis/trans* diastereoisomeric mixture of 4-methylcyclohexylamine at 150 °C provided the carboxamide. N-Alkylation with 4-(2-chloroethyl)morpholine afforded the desired 1,8-naphthyridin-2-one derivative. Suzuki reaction by generating in situ Pd(PPh₃)₄, in aqueous Na₂CO₃ and boronic acid yielded the final compound.

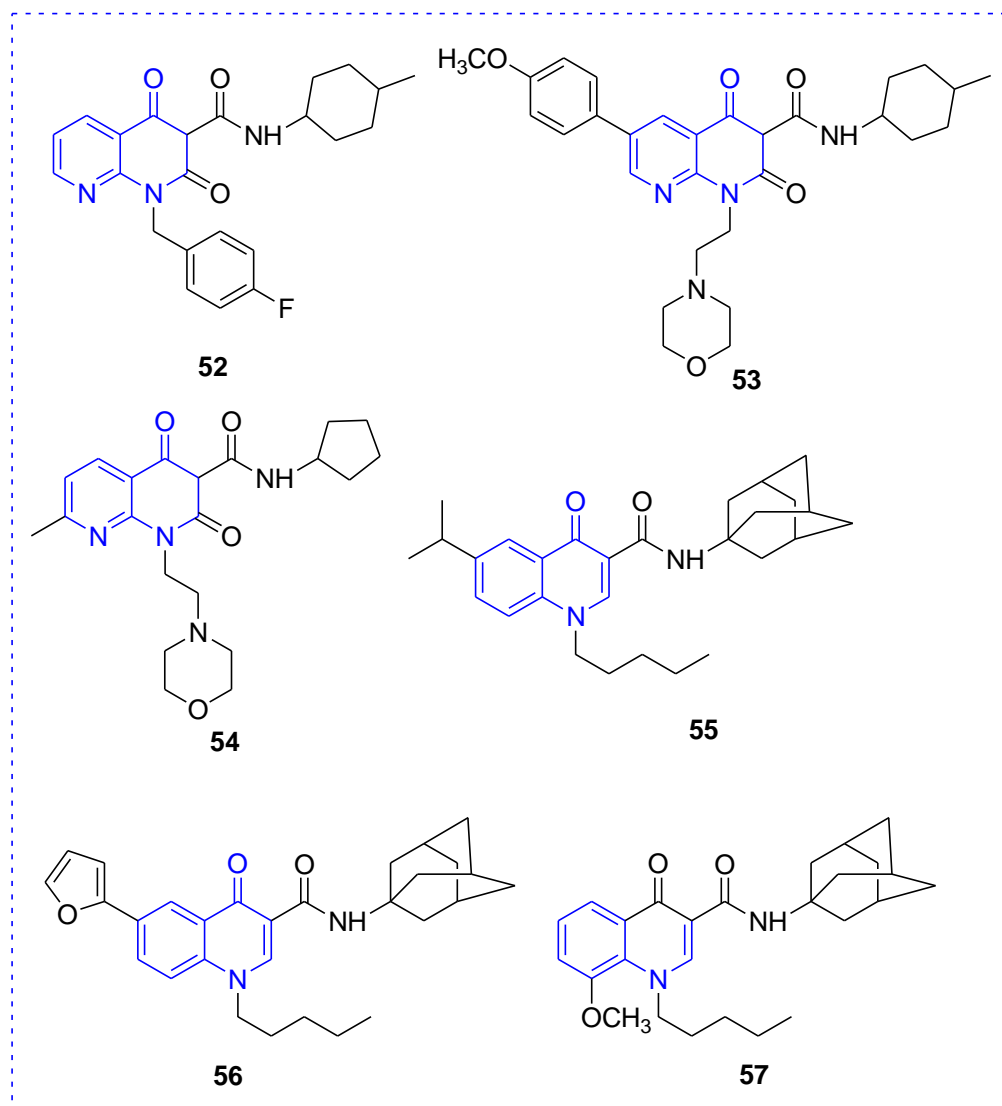
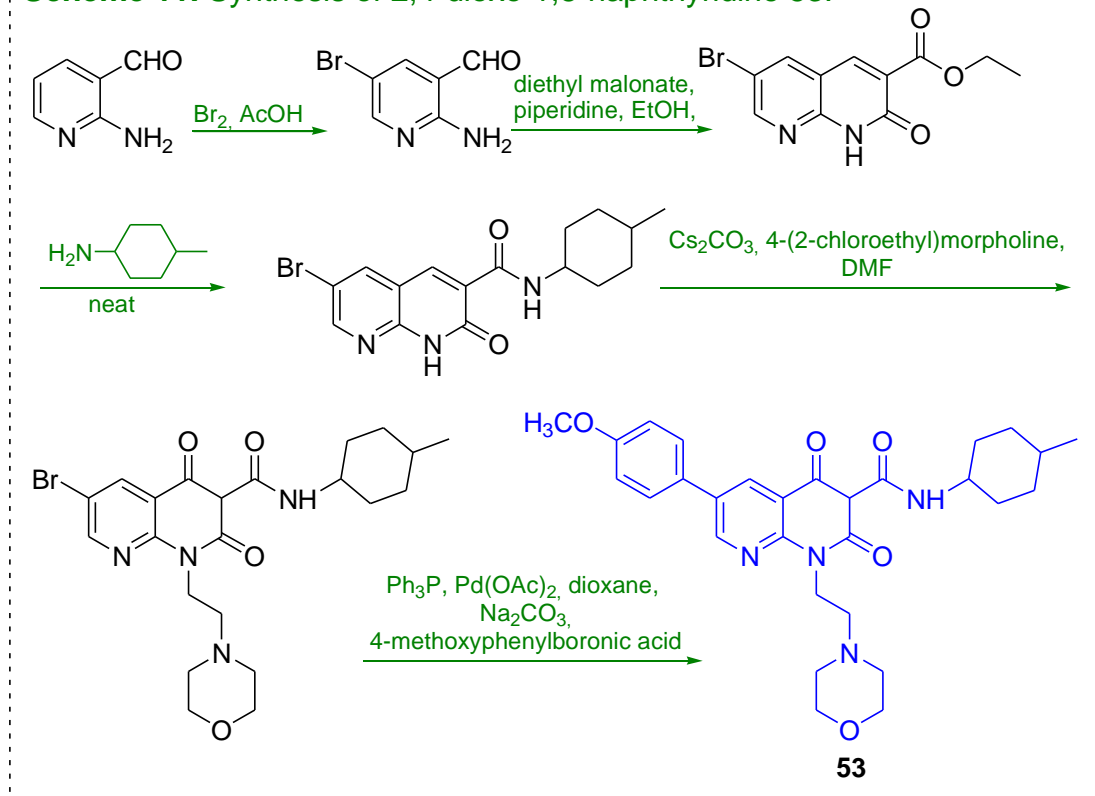


Figure 11. Quinolone and Naphthyridine derivatives.

Scheme 11. Synthesis of 2,4-dioxo-1,8-naphthyridine 53.



7.10. 4-Oxo-1,4-dihydropyridine

4-Oxo-1,4-dihydropyridines, as exemplified by the compound **58** (Figure 12, Scheme 12), has been described as potent and selective CB_2 receptor agonist (hCB_2 $K_i = 20$ nM, hCB_1 $K_i > 3000$ nM, Emax 148%).²⁷⁷ Affinity and functionality of this group of compounds was shown to be dependent on the nature of substituents around the heterocycle and the C-6 substituent was crucial in controlling the functionality of this series of compounds. Using a β_2 -adrenergic receptor-based CB_2 receptor model, the authors suggested that the phenyl at C-6 confers the inverse agonist profile by blocking the χ_1 torsion of Trp6.48 side chain in its inactive conformation. The substituents at N-1 and C-3 position were important for affinity but not for functionality. Further optimization efforts focused on the introduction of substituent at C-5, such as a pyridine heterocycle (**59**, Figure 12), was well tolerated with a K_i of 36 nM. Compound **58** failed to act on $\text{TNF-}\alpha$ and $\text{IL-1}\beta$ levels, had

drastically decreased macroscopic scores, histological damage, and Myeloperoxidase (MPO) activity, and had a protective effect against colitis.²⁷⁸

Very recently, the 2*H*-pyrazolo[4,3-*c*]quinolin-3(5*H*)-one scaffold was developed as constrained analogues of 4-oxo-1,4-dihydroquinoline-3-carboxamide series with improved affinity for the hCB₂ and selectivity over the hCB₁ receptors. The lead of this series, (**60**: hCB₂ K_i = 0.39 nM, hCB₁ K_i > 3000 nM, Figure 12) was found to protect mice against experimental colitis after oral administration.²⁷⁹

*Preparation of N3-(1-Adamantyl)-6-methyl-1-pentyl-4-oxo-1,4-dihydropyridine-3-carboxamide (58).*²⁷⁷ The synthesis of compound 58 is outlined in Scheme 12. The commercially available 4-hydroxy-6-methyl-2-pyrone was reacted with N,N-dimethylformamide dimethyl acetal to give (3*Z*)-3-((dimethylamino)methylene)-3,4-dihydro-4-hydroxy-6-methylpyran-2-one which, treated with *n*-pentylamine, under alkaline conditions gave the carboxylic acid derivative. Under peptide coupling conditions with 1-aminoadamantane hydrochloride the target amide was obtained.

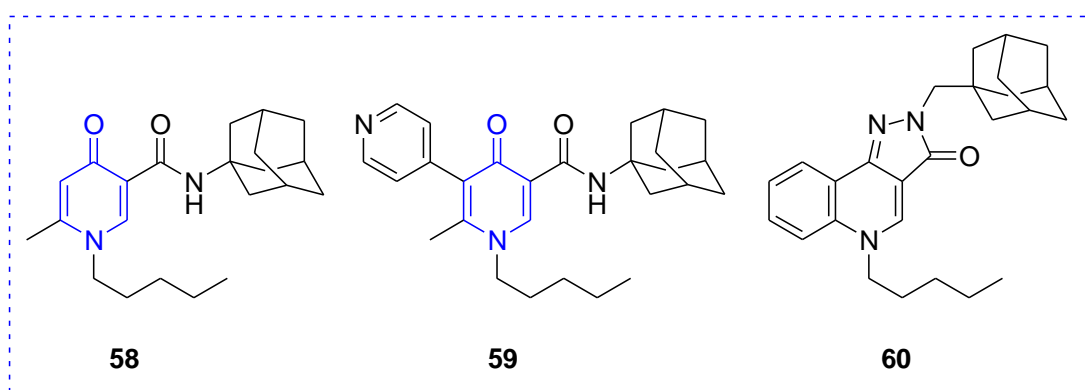
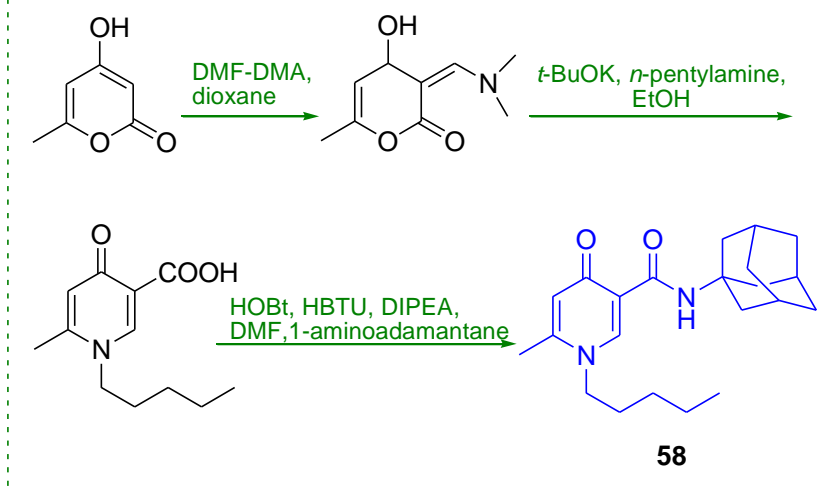


Figure 12. 4-Oxo-1,4-dihydropyridine derivatives

Scheme 12. Synthesis of 4-oxo-1,4-dihydropyridine 58.



7.11. Decahydroquinoline amide

Decahydroquinoline CB₂ agonists were reported by the Merck Research Laboratories. Optimization of the amide substituent led to improvements in CB₂ selectivity over CB₁ receptors as well as physicochemical properties. Two compounds were examined in a rat model of acute inflammatory pain where the quite selective CB₂ agonist **61** (ratCB₂ EC₅₀ = 6.3 nM, ratCB₁ EC₅₀ = 1887 nM) (Figure 13, Scheme 13) displayed a dose-dependent analgesic effect. The CB₂ agonist (**62**, Figure 13), (ratCB₂ EC₅₀ = 81 nM, ratCB₁ EC₅₀ > 17000 nM) lacking functional CB₁ activity was inactive in this model even with high in vivo exposure both peripherally and centrally.²⁸⁰

*Preparation of ((4R,4aS,8aR)-octahydro-4-hydroxy-4-phenylquinolin-1(2H)-yl)(6-methylpyridin-3-yl)methanone (61).*²⁸⁰ 1-Acetylcyclohexene was reacted with paraformaldehyde and dimethylamine hydrochloride under Mannich conditions to give the aminoketone, which was treated with concentrated aqueous ammonia under pressure to yield decahydroquinolinone intermediate. Acylation with benzyloxycarbonyl chloride gave a primarily *trans* mixture of (4a*R*,8a*R*)-benzyloctahydro-4-oxoquinoline-1(2*H*)-carboxylate. Isomerization of the center adjacent to the

ketone was achieved by treatment with K_2CO_3 in MeOH to give *cis*-isomer in 30%. Addition of phenylmagnesium bromide at low temperature gave alcohol intermediate. Deprotection by hydrogenolysis gave racemic (4*R*,4*aS*,8*aR*)-decahydro-4-phenylquinoline which was acylated in standard peptide coupling conditions. (Scheme 13)

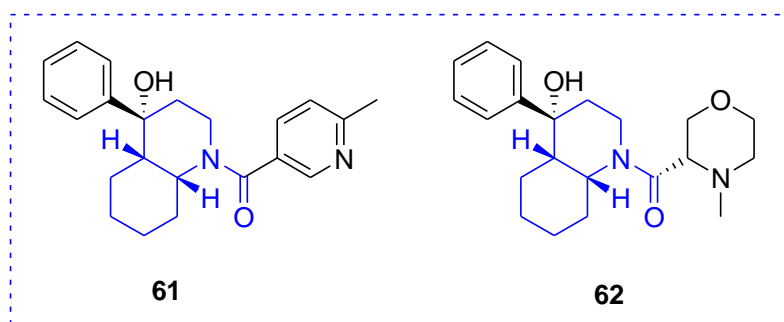
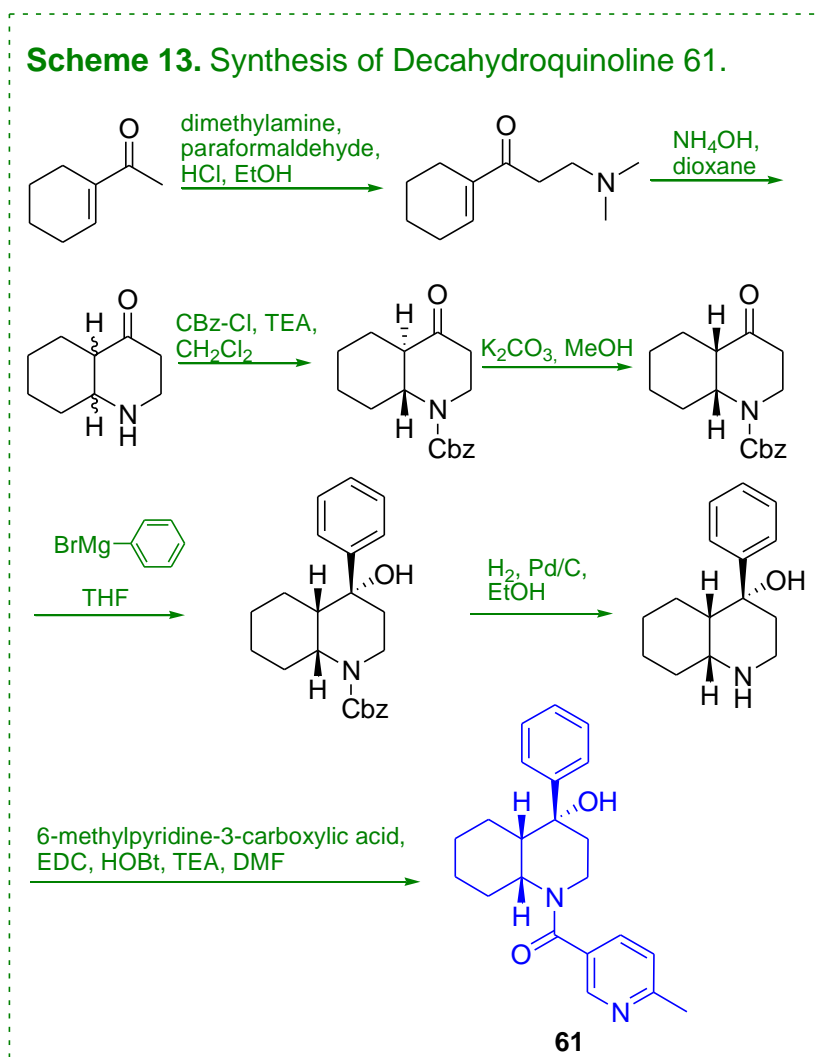


Figure 13. Decahydroquinoline amide derivatives.



7.12. Carboline

The γ -carboline series was identified by scaffold-hopping from the bicyclic benzimidazole core as mixed CB₁ and CB₂ agonists.²⁸¹ This new structural class of cannabinoid agonists showed good physicochemical properties and low CNS penetration. The SAR of the tetrahydro-1*H*-pyrido[4,3-*b*]indole motif of compound **63** (Figure 14) was extensively examined and the cyclopentyl and 4-tetrahydropyran analogs **64** and **65** (Figure 14) showed a >10 fold increase in agonist activity and preserved reasonable metabolic stability. Optimization of physicochemical properties was achieved by the ethylsulfonyl analogue, compound **66** (Figure 14). This molecule was found to be a potent agonist on hCB₁ (EC₅₀ = 49 nM, Emax 120%) and in rat brain tissue (ratCB₁ EC₅₀ = 85 nM, Emax 156%) exhibiting significant anti-hyperalgesia in a rat inflammatory pain model with low levels of CNS penetration.

*Preparation of 5-(ethylsulfonyl)-8-[(4-methylpiperidin-1-yl)carbonyl]-2-(tetrahydro-2*H*-pyran-4-yl)-2,3,4,5-tetrahydro-1*H*-pyrido[4,3-*b*]indole (66).*²⁸¹ 4-Hydrazinobenzoic acid was heated with 4-piperidinone in dioxane and hydrochloric acid, then treated with BOC-anhydride to give 2-(*tert*-butoxycarbonyl)-2,3,4,5-tetrahydro-1*H*-pyrido[4,3-*b*]indole-8-carboxylic acid. Coupling reaction with methylpiperidine furnished the amide intermediate, which was treated with ethylsulfonyl chloride and sodium hydride to yield the 5-ethylsulfonyl compound. Subsequent debocylation with trifluoroacetic acid gave the free amine which was transformed to the target compound by means of 4-tetrahydropyranone and sodium triacetoxyborohydride in dichloromethane. (Scheme 14)

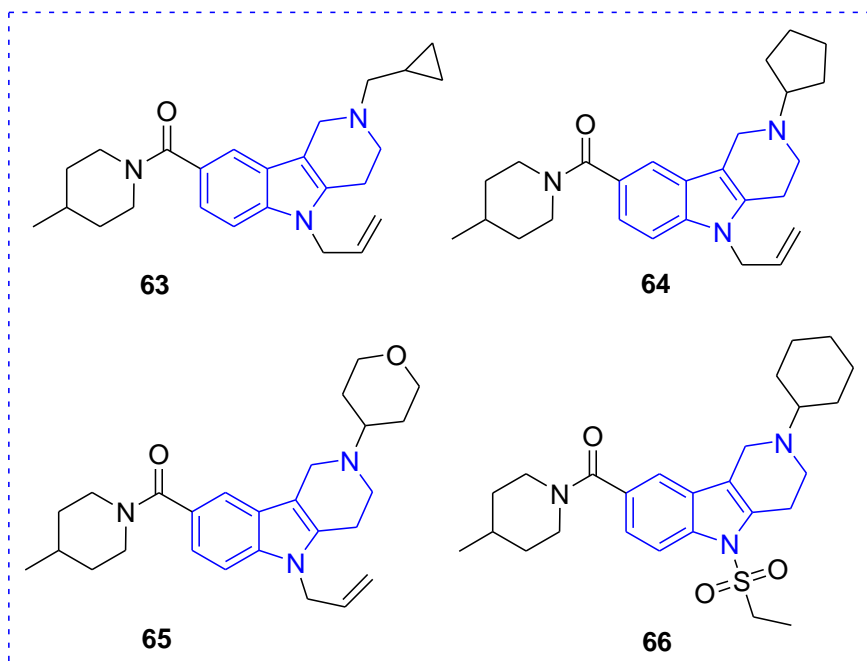
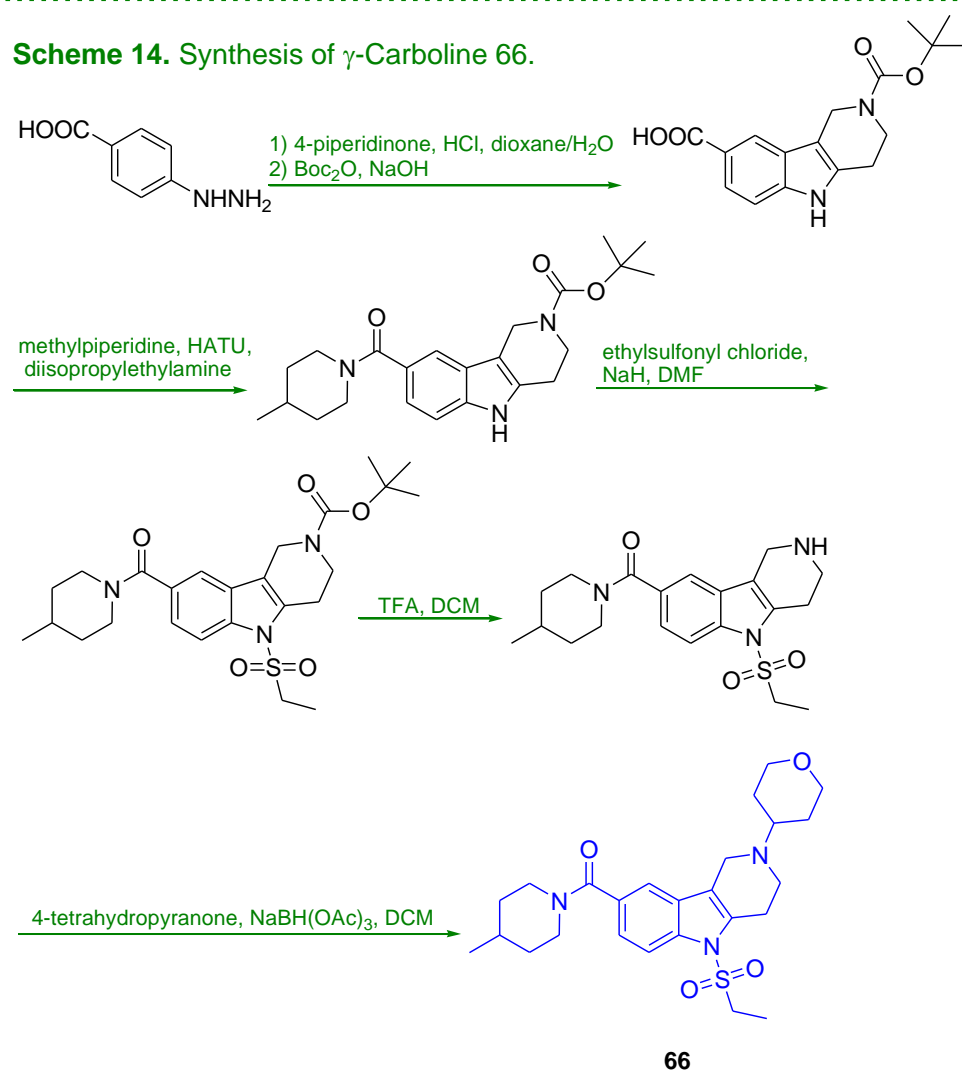


Figure 14. γ -Carboline derivatives.

Scheme 14. Synthesis of γ -Carboline 66.



7.13. Heteroarylpyridine, Heteroarylpyrimidine

The heteroaryl-4-oxopyridine/pyrimidine derivatives have been described.¹⁵ In this family, the core structure of molecules defined the type of activity to be seen, while the substituents around the core structure were used to modulate the potency. Compound **67** (Figure 15, Scheme 15), a representative analog of the heteroarylpyridine series, displayed high affinity and selectivity for CB₂ receptor (hCB₂ K_i = 11.4 nM, hCB₁ K_i = 4568 nM). This compound was found to act as partial agonist (E_{max} = 14%).

In this study, further CB₂ ligands were synthesized by replacing the pyrazolo ring with different heterocycles that were found to be potent CB₂ receptor ligands. Replacement of the pyrazolo ring of the parent nucleus with differently substituted 5-membered heterocycles allowed further SAR studies. The isoxazolopyridines, such as compound **68** (Figure 15), behaved as full agonists and the thienopyridines (for example **69**, Figure 15) acted as inverse agonists. The inverse agonists thieno[2,3-*b*]pyridine derivatives showed high affinity at the CB receptors (hCB₂ K_i values from 1.12 to 12.3 nM) with an apparent decrease in selectivity. The calculation of log P indicated the absence of a significant correlation between the lipophilicity and biological activities within this series.

A major focus on the optimization effort was to increase selectivity of the previous series of heteroarylpyridine/pyrimidine derivatives. The 7-oxopyrazolo[1,5-*a*]pyrimidine-6-carboxamides, structural isomers of the previously pyrazolo[3,4-*b*]pyridines were synthesized, exemplified by structures such as **70** (hCB₂ K_i = 2.5 nM, hCB₁ K_i > 10000 nM), and **71** (hCB₂ K_i = 4.9 nM, hCB₁ K_i > 10000 nM) (Figure 15), which showed stimulatory effects on forskolin-induced cAMP production acting as inverse agonists.¹⁶

*Preparation of trans-4,7-dihydro-1,3-dimethyl-N-(4-methylcyclohexyl)-4-oxo-7-pentyl-1H-pyrazolo[3,4-*b*]pyridine-5-carboxamide (67).*¹⁶ The synthetic route to obtain the target compound **67** is outlined in Scheme 15. Replacement of the ethoxy group of diethyl ethoxymethylenemalonate

by the 5-amino function of the pyrazole afforded the intermediate diethyl 5-pyrazolylaminomethylenemalonate, which was cyclized giving the ethyl pyrazolo[3,4-*b*]pyridine-5-carboxylate. Hydrolysis yielded the corresponding 5-carboxylic acid intermediate which by coupling reaction afforded the amide (The separation of *cis* and *trans* isomers was obtained by flash chromatography on silica gel). Alkylation at N-7 in the presence of potassium carbonate with pentylbromide was performed to give the target compound 67.

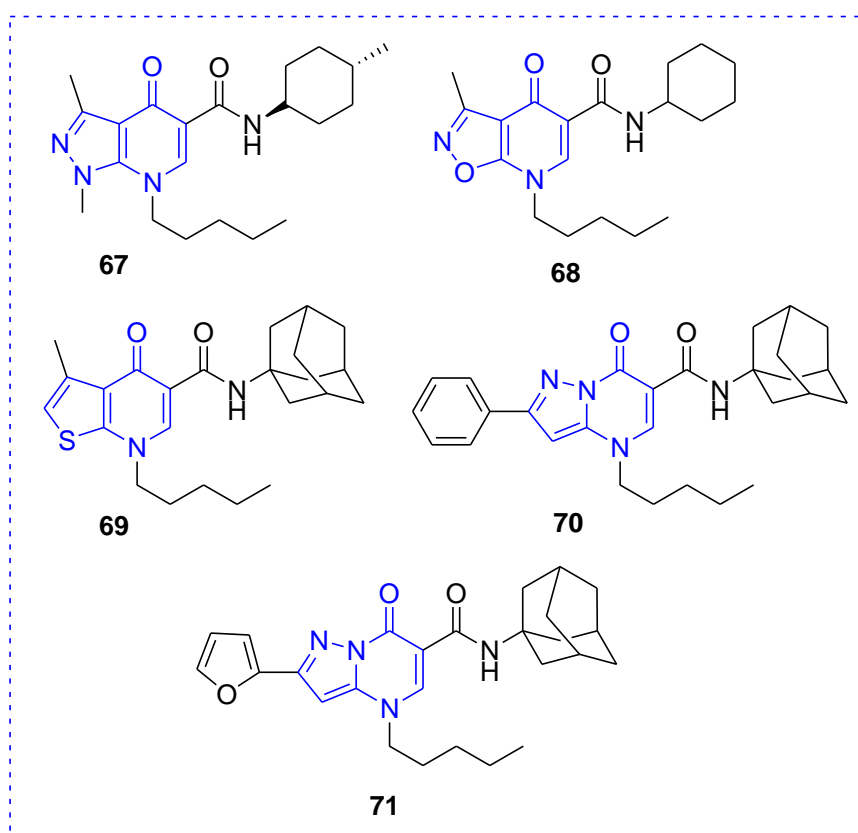
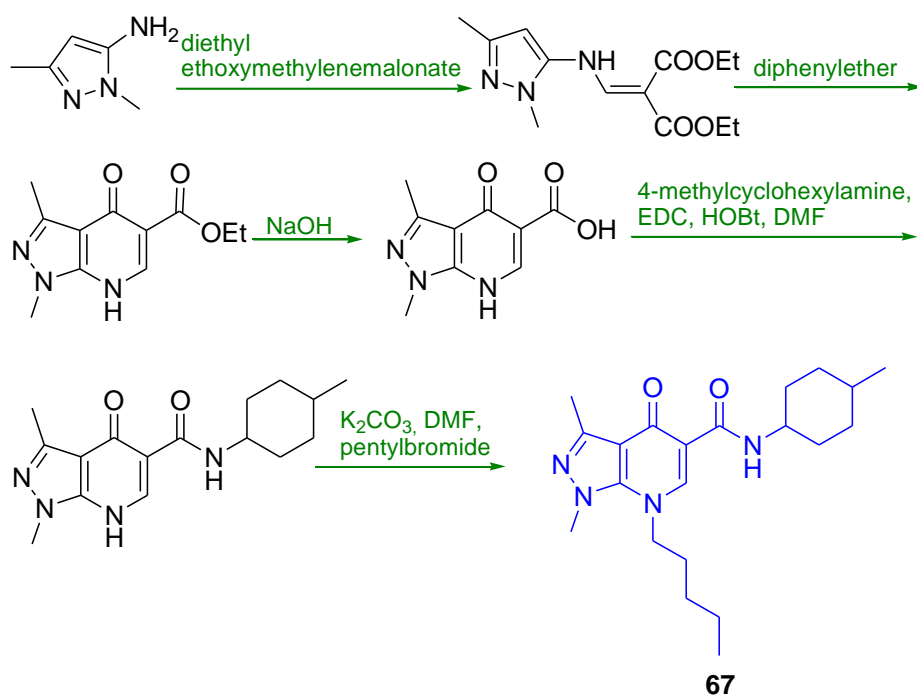


Figure 15. Heteroarylpyridone, Heteroarylpurimidone derivatives.

Scheme 15. Synthesis of pyrazolopyridine 67.



7.14. Indol-3-ylcycloalkyl ketone

Huffman and co-workers have widely investigated indole cannabinoid ligands and were the first to illustrate that N-1 alkyl side chains are tolerated in place of the aminoalkyl side chains that were previously thought to be necessary for activity at the cannabinoid receptors.^{282,283} They also found that the size of the N-1 alkyl side chain had a significant impact on selectivity toward CB₂ receptors. Makriyannis's group also discovered the aminoalkylindoles, including the CB₂ selective ligand **72** (AM1241, Figure 16). Compound **72** was efficacious in a range of preclinical inflammatory and neuropathic pain models.^{234,233} While many of the indole derivatives have commonly favored 3-naphthoyl or 3-aroil groups, a diverse chemical series comprising a tetramethyl cycloalkyl ketone at this position (compound **73**, A-796260, Figure 16, Scheme 16) was discovered.²⁸⁴ Compound **73** was found to possess receptor affinity and selectivity, having a CB₂ K_i of 4.6 nM versus 945 nM at CB₁ receptor. It has potent analgesic and anti-inflammatory actions in models of neuropathic pain without producing cannabis-like side effects.²⁸⁵ The

tetrahydropyranylmethyl analogue **74** (Figure 16), one of the most potent CB₂ agonists in this series, did not exhibit good selectivity for the CB₂ receptor versus the CB₁ receptor (CB₂ K_i = 0.2 nM, CB₁ K_i = 12 nM).²⁸⁴ A stereochemical effect was noted on binding selectivity with the tetrahydrofuranylmethyl **75** (Figure 16), the *R*-enantiomer exhibited more selectivity for the CB₂ receptor than the *S*-enantiomer.

*Preparation of (2,2,3,3-tetramethylcyclopropyl)(1-(2-morpholinoethyl)-1H-indol-3-yl)methanone (73).*²⁸⁴ As depicted in Scheme 16, the unsubstituted indole core was acylated with 2,2,3,3-tetramethylcyclopropanecarbonyl using ethylmagnesium bromide and zinc(II) chloride (ZnCl₂). The C-3 acylated product then underwent N-alkylation with 2-morpholin-4-ylethyl methanesulfonate in DMF by means of sodium hydride.

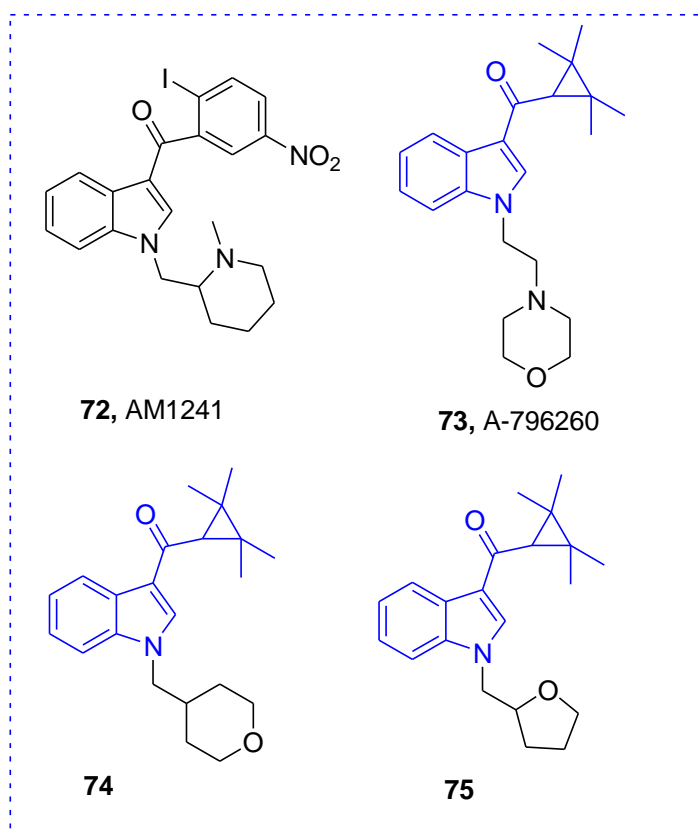
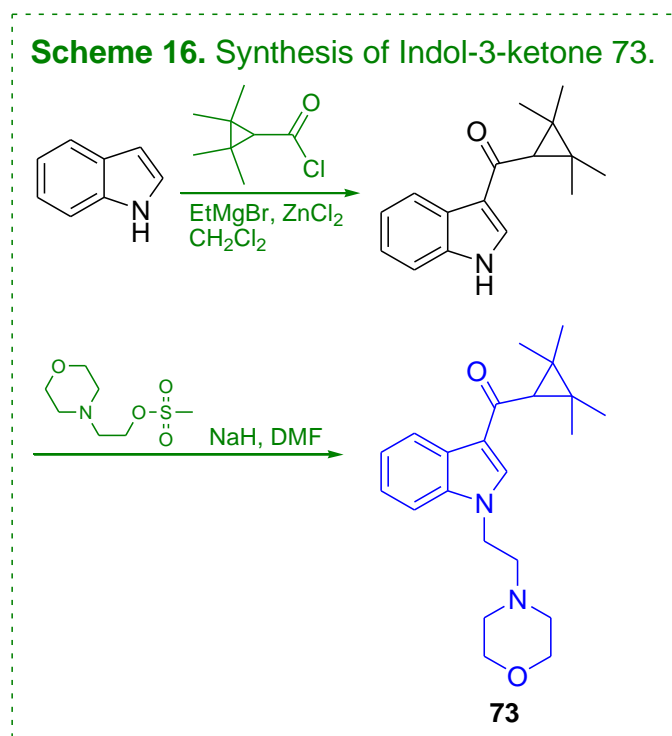


Figure 16. Indol-3-ylcycloalkyl ketone derivatives.



7.15. Oxazinoquinoline

7-Oxo-[1,4]oxazino[2,3,4-*ij*]quinoline-6-carboxamide chemotype is reported as a novel cannabinoid ligand possessing high CB_2 receptor affinity.¹⁴ This scaffold was designed as a hybrid chemical structure that incorporated the structural features of known cannabinoid ligands. The design of the new oxazinoquinolines was based on the quinolone compounds and the cannabimimetic indole derivative WIN55,212-2.²⁸⁶

The N-cycloheptyl-3,7-dihydro-7-oxo-3-phenyl-2*H*-[1,4]oxazino[2,3,4-*ij*]quinoline-6-carboxamide (**76**, Figure 17) exhibited high affinity and selectivity for the CB_2 receptor (hCB_2 $K_i = 0.8$ nM, hCB_1 $K_i = 310$ nM). Compound **77** (MT178, Figure 17, Scheme 17), in which a pyrrolidine moiety was introduced in position 10, showed high affinity and selectivity at the CB_2 receptor (hCB_2 $K_i = 8.1$ nM, $\text{SI} > 1231$). MT178 produced a robust analgesia in inflammatory and chronic pain models via CB_2 receptors without CB_1 -mediated central side effects.²⁰⁹ The SAR studies were extended by replacement of the C-3 phenyl ring with a series of aliphatic moieties. The effect of a chiral center

on the biological activity was also investigated, and it was found that the *R*-enantiomers exhibited greater affinity at the CB₂ receptor than the *S*-enantiomers. Compound (*R*)-**78** (Figure 17), the enantiopure derivative bearing an isobutyl moiety at C-3, was found to bind to the CB₂ receptor with high affinity and excellent selectivity (hCB₂ K_i = 9.2 nM, SI > 1082). In 3,5- cyclic adenosine monophosphate assays, the novel series behaved as full agonists, exhibiting functional activity at the human CB₂ receptor.¹⁴

*Preparation of N-Adamant-1-yl-3,7-dihydro-3-ethyl-7-oxo-10-pyrrolidin-1-yl-2H-[1,4]oxazino[2,3,4-ij]quinoline-6-carboxamide (77).*¹⁴ The target oxazinoquinoline **77** was prepared following the synthetic route depicted in Scheme 17. Ethyl 3-(2,3,4-trifluorophenyl)-3-oxopropanoate was prepared from the corresponding benzoic acids by reaction with carbonyldiimidazole (CDI) in THF to afford the imidazolide, followed by condensation with the magnesium salt of monoethyl malonate. Subsequent condensation with triethyl orthoformate in refluxing acetic anhydride produced the ethyl 2-(ethoxymethylene)propionate. This intermediate was reacted with racemic 2-aminobutanol in an addition–elimination sequence to afford the ethyl trifluorobenzoyl acrylate. Cyclization using potassium carbonate in dimethylformamide (DMF) at high temperature and then saponification afforded the carboxylic acid that was coupled with 1-adamantanamine to yield the amide derivative. The target compound **77** was obtained by displacement of the fluorine substituent by treating the amide with nucleophile pyrrolidine in alkaline conditions.

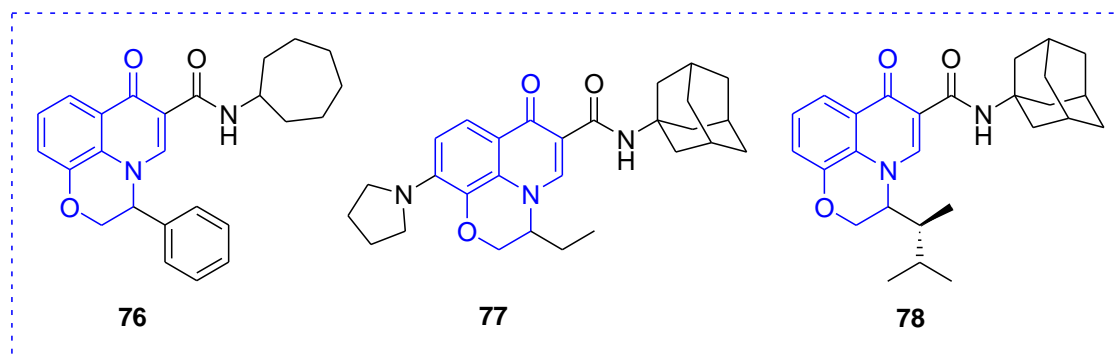
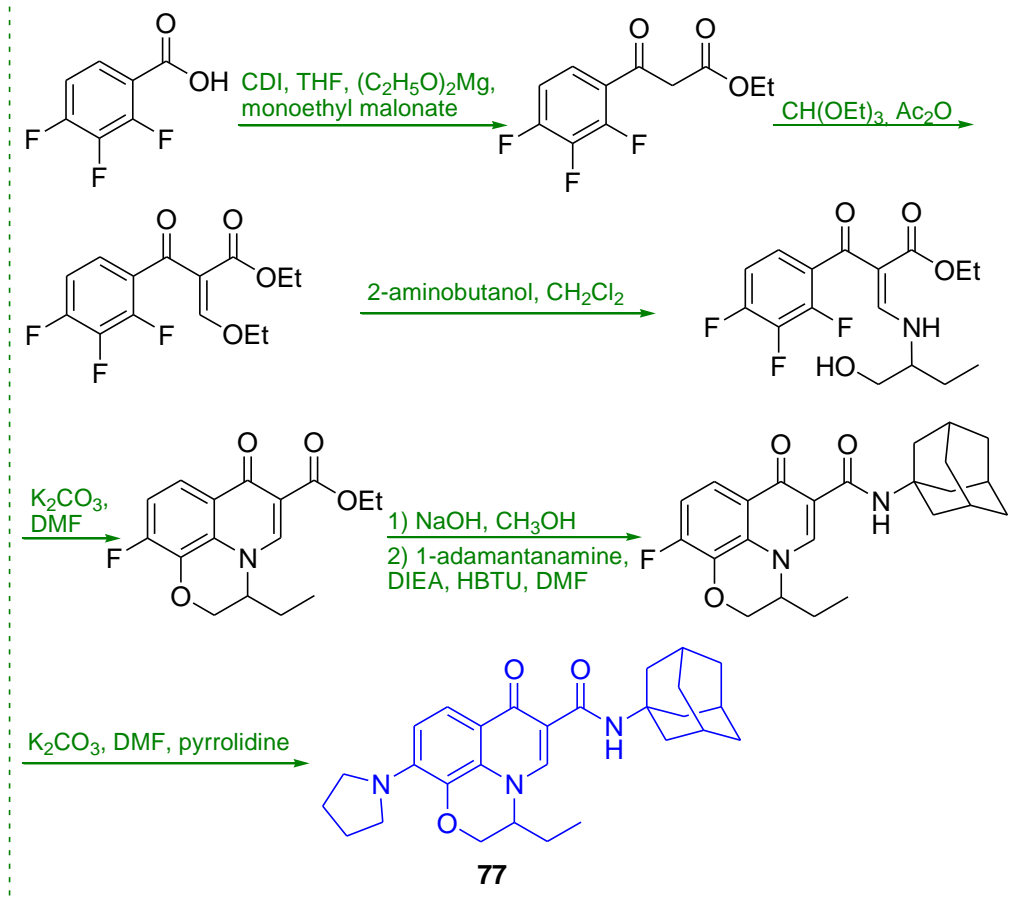


Figure 17. Oxazinoquinoline derivatives.

Scheme 17. Synthesis of Oxazinoquinoline 77.



7.16. 1,4-Diazepane

From a high-throughput screen to identify CB₂ agonists, the 1,4-diazepane hit compound **79** (EC₅₀ = 136 nM, E_{max} 67%) was identified (Figure 18, Scheme18).²⁸⁷ It was revealed to be a partial agonist and did not activate the CB₁ receptor up to 20 μM, but suffered from low metabolic stability. The optimization effort on the diazepane scaffold, achieved by decreasing lipophilicity, generated molecules which improved in vitro clearance as compared to the hit **79**. The pharmacokinetic profile of compound **80** (EC₅₀ = 67 nM) bearing an isoxazole (Figure 18), more polar substituent in concomitance with the tetrahydropyran fragment in various in vitro assays were more favorable than those of the phenyl urea **79**. Expanding upon the tetrahydropyran substituent, analog **81** (Figure 18) was synthesized and greatly enhanced CB₂ potency (EC₅₀ = 1 nM), selectivity and improved the aqueous solubility.²⁸⁸

Preparation of 4-(3-cyclopentylpropanoyl)-N-(4-butylphenyl)-1,4-diazepane-1-carboxamide (79).²⁸⁷ The commercially available *tert*-butyl homopiperazine-1-carboxylate was coupled with 3-cyclopentylpropanoic acid employing either amide coupling conditions or via acid chloride, followed by BOC deprotection using HCl. The urea coupling was performed using the 1-butyl-4-isocyanatobenzene which was generated in situ using triphosgene. (Scheme 18)

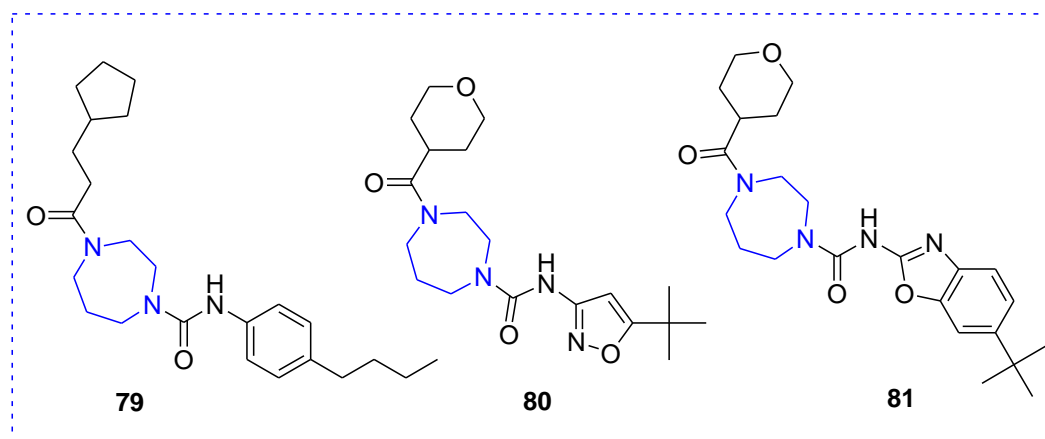
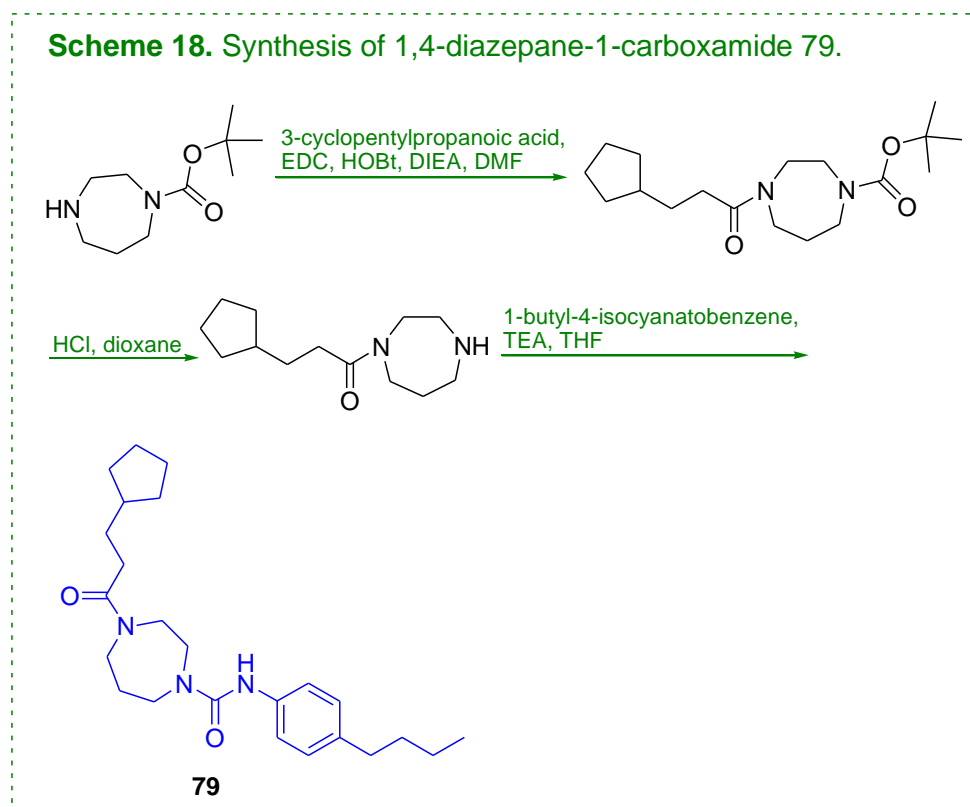


Figure 18. 1,4-Diazepane derivatives.



7.17. (S)Proline, (S)Piperidine

Boehringer Ingelheim Pharmaceuticals analyzed the diazepane and the β -sulfonylacetamide compounds, through computational and experimental approaches in search of the bioactive conformation common to these chemical series. Additionally, computer-aided drug design (CADD) strategies were adopted toward the identification of new cores to replace the diazepane scaffold. The parallels in SAR between the two series led to the synthesis of the proline-, and piperidine-based series as CB₂ full agonists (compounds **82** and **83**, respectively, Figure 19). Analogs containing the new proline scaffold exhibited picomolar CB₂ activity, and high selectivity over CB₁ receptors (**82**: CB₂ EC₅₀ = 0.07 nM, CB₁ EC₅₀ = 53 nM; **83**: CB₂ EC₅₀ = 0.09 nM, CB₁ EC₅₀ = 250 nM).²⁸⁹ Further optimization within the series led to oxo-proline compound **84** (Figure 19, Scheme 19), which showed a favorable pharmacokinetic profile of which the antinociceptive effect of this CB₂ selective agonist in a diabetic neuropathy model was evaluated.²⁹⁰ The correlated CB₂ agonist based on a (*S*)-piperidine is represented by compound **85** (Figure 19) that displayed selectivity over CB₁ receptor and suitable drug like properties (hCB₂ EC₅₀ = 10 nM, hCB₁ EC₅₀ > 20000 nM).²⁹¹ In rats, compound **85** demonstrated a favorable pharmacokinetic profile and efficacy in a Streptozotocin-induced diabetic neuropathy model, with full reversal of mechanical hyperalgesia.

*Preparation of (S)-N-(3-tert-butylisoxazol-5-yl)-1-(4-(trifluoromethyl)phenyl)-5-oxopyrrolidine-2-carboxamide (84).*²⁹⁰ As shown in Scheme 19, (*L*)-pyroglutamic acid and 4-trifluoromethyl phenylboronic acid are converted in the presence of catalyst di- μ -hydroxo-bis[(*N,N,N',N'*-tetramethylethylenediamine)copper(II)] chloride (CuTMEDA) to the *N*-aryl substituted intermediate. Typical peptide coupling conditions were applied to introduce 3-*tert*-butylisoxazol-5-amine to provide the final compound.

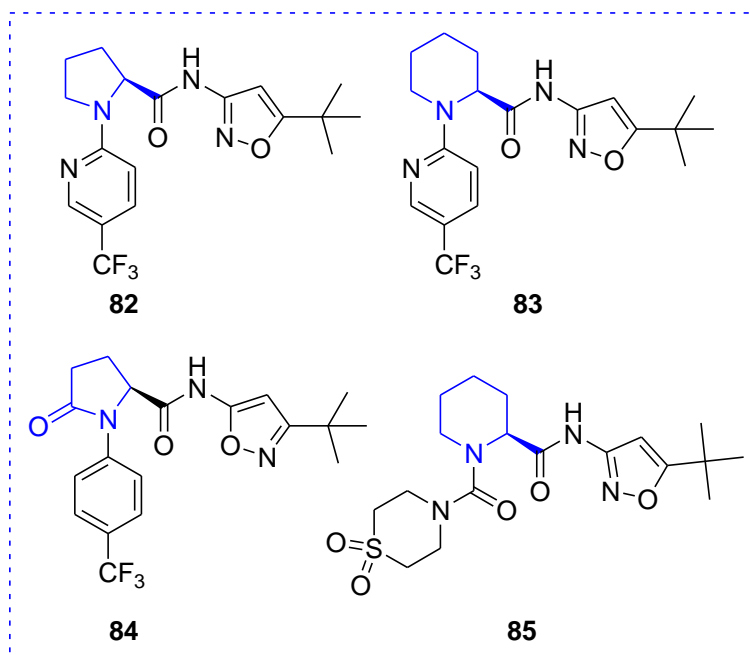
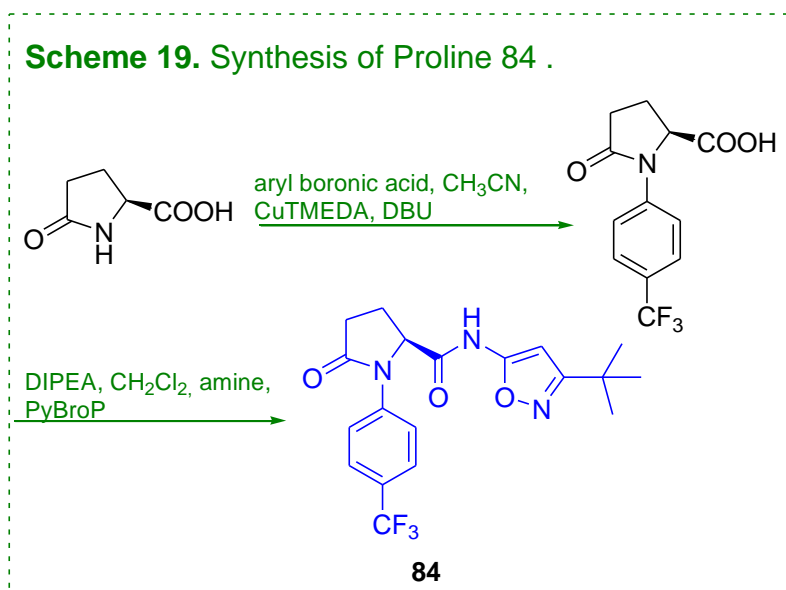


Figure 19. Proline, Piperidine derivatives.



7.18. Pyrazole-3-carboxamide

Arena Pharmaceuticals designed tricyclo pyrazole 3- carboxamides that displayed agonism at both CB receptors, and also suffered from poor in vitro and in vivo properties. Starting from this series, investigation on N-1 pyrazole substituents, through introduction of a variety of alkyl and aryl moieties, led to improvements in CB₂ potency and microsomal stability. SAR studies led to the finding of the favored compound (*R,R*)-**86** (Figure 20) which had good activity at CB₂ (hEC₅₀ = 3.7 nM) and was selective against CB₁ (SI > 8100). This compound exhibited an overall favorable ADME, and was efficacious at reversing inflammatory pain after oral administration in the rat.²⁹²

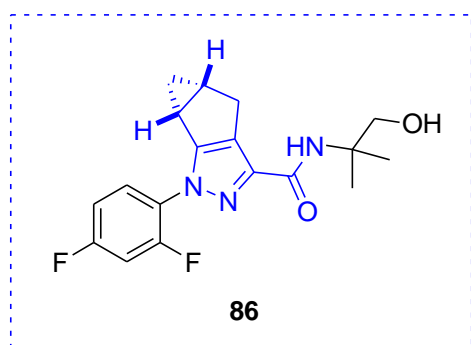


Figure 20. Pyrazole-3-carboxamide derivative.

7.19. 3-Carboxamido-5-aryl-isoxazole

The 3-carboxamido-5-aryl-isoxazole is reported as a good scaffold to design selective CB₂ agonists and FAAH inhibitors. In this family, the substituents on the heterocycle define the activity on CB₂ receptor or on FAAH enzyme. The synthesis and SARs of isoxazole derivatives show that the presence and position of the alkoxy chain deeply influence the biological activity of the molecule. Compounds such as **87** (ALIAE809, Figure 21) with a 2-substituted phenyl group at position 5 and a bulky aliphatic group on the carboxamide function have a K_i value of of 9.0 nM on hCB₂ receptors. Some of the isoxazole compounds have shown, in vivo, anti-inflammatory activities in a colitis mouse model.²⁹³ When the alkoxy chain is missing, or is at positions 3 or 4 (compounds **88**

and **89**, Figure 21), the molecules show potent FAAH inhibitor activity. Compounds **88** and **89** inhibit the development of dextran sulfate sodium (DSS)-induced acute colitis in mice.²⁹⁴

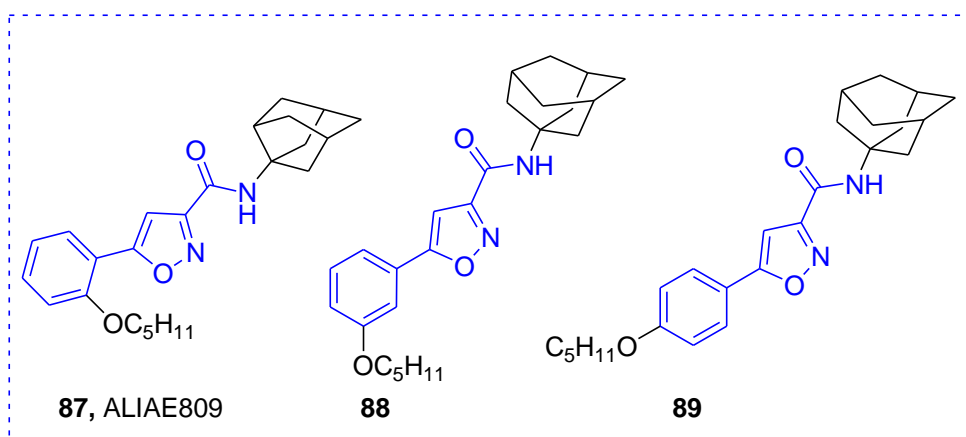


Figure 21. 3-Carboxamido-5-aryl-isoxazole

7.20. 1,3,5-Triazine

2,4,6-Trisubstituted 1,3,5-triazines were identified as potent CB₂ agonists by 3D ligand-based virtual screening. Within this series, several CB₁ receptor antagonists/ inverse agonists and CB₂ receptor agonists, of which *N*-cyclopentyl-4-ethoxy-6-(4-methylpiperidin-1-yl)-1,3,5-triazin-2-amine (**90**) (Figure 22, Scheme 20) was selected for further development. SAR study of the analogs exposed that replacing of the cyclopentyl substituent by a bulkier *N*-adamantan-1-yl chain was useful, such as in compound **91** of this series. It possessed the best potency and selectivity with only a weak CB₁ agonist activity (CB₂ EC₅₀ = 3.2 nM).²⁹⁵ Subsequently, an additional series of more polar analogues, which were found to possess high CB₂ agonist activity with enhanced water solubility, were described. The most potent compounds in the series were *N*-(adamantan-1-yl)-4-ethoxy-6-(4-(2-fluoroethyl)piperazin-1-yl)-1,3,5-triazin-2-amine (**92**, Figure 22) and *N*-(adamantan-1-yl)-4-ethoxy-6-(4-methylpiperazin-1-yl)-1,3,5-triazin-2-amine (**93**, Figure 22) with EC₅₀ value of 0.60 and 1.6 nM, respectively.²⁹⁶ Compound **93** induced cell viability decreased in prostate and leukemia cell lines, and diminished proliferation of C8161 melanoma cells.

Preparation of *N*-cyclopentyl-4-ethoxy-6-(4-methylpiperidin-1-yl)-1,3,5-triazin-2-amine (87).²⁹⁵As shown in Scheme 20, displacement of chlorine atoms in cyanuric chloride with different substituents in basic conditions and tetrahydrofuran as solvent led to the preparation of target compound.

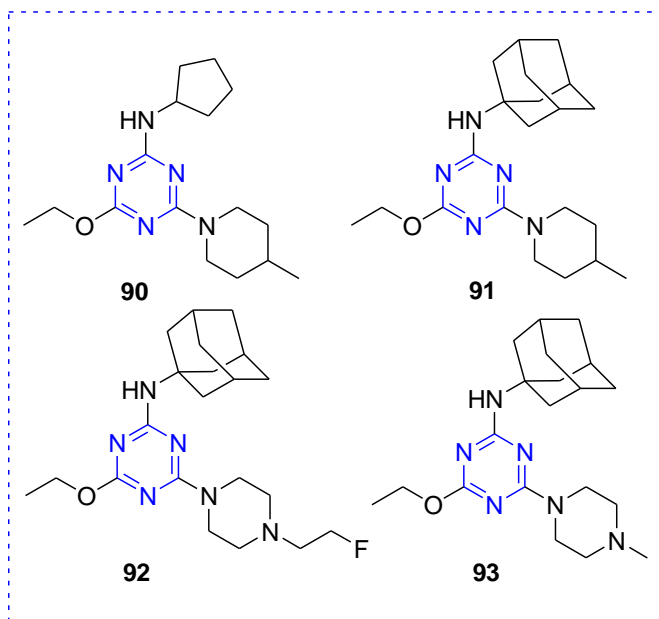
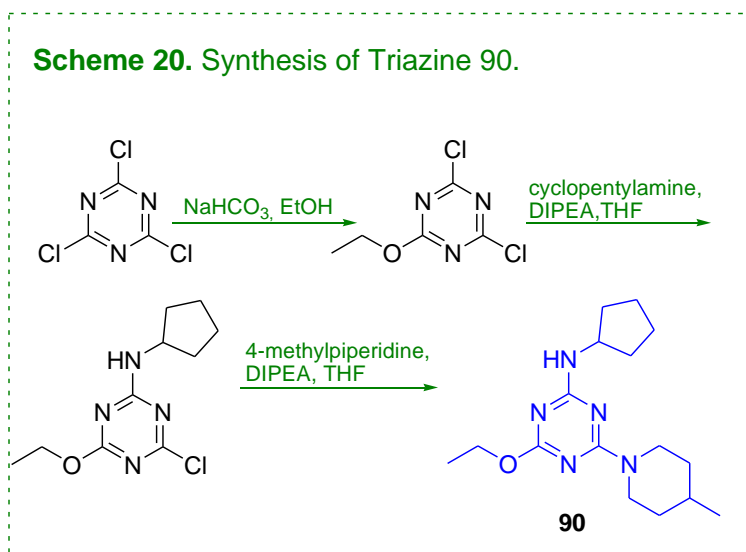


Figure 22. Triazine derivatives.



7.21. Analogs of HU-308

The discovery of CB₂-selective ligand HU-308 (hCB₂ K_i = 22 nM, rCB₁ K_i > 10000)²³¹ (**94**, Figure 23) supported the evidence of analgesic effects induced by the selective activation of CB₂ receptors.²³² More recently, a structurally similar compound labeled HU-910 (**95**, Figure 23) is reported as potent CB₂ agonist (hCB₂ K_i = 6 nM, hCB₁ K_i = 1400 nM), which may exert protective effects in several diseases related to inflammation and tissue injury.²⁹⁷ The enantiomer of HU-308 labelled as HU-433 (**96**, Figure 23) was also synthesized with higher potency than HU-308 in mouse models such as the release of ovariectomy-induced bone loss and ear inflammation. In addition, HU-433 has a lower affinity in comparison with HU-308 showing an inverse relationship between binding affinity and biological potency of the two enantiomers with two different binding conformations responsible for the affinity difference.²⁹⁸

Preparation of ((1R,4R,5S)-4-(2,6-dimethoxy-4-(2-methyloctan-2-yl)phenyl)-6,6 dimethylbicyclo [3.1.1]hept-2-en-2-yl)methanol (96). As shown in Scheme 21, (1R)-myrtenol was treated with pivaloyl chloride to give myrtenyl pivalate which was transformed to the 4-oxo-myrtenyl pivalate by means of chromium trioxide and *tert*-butyl hydroperoxide. The reduction with NaBH₄ yielded the 4-hydroxy-6,6-dimethylbicyclo intermediate which was reacted with 5-(1,1-dimethylheptyl)resorcinol in presence of *p*-toluenesulfonic acid to give 2-(3-myrtenyl pivalate)-5-(1,1-dimethylheptyl)resorcinol. O-alkylation with methyl iodides and successive reduction reaction with lithium aluminium hydride gave the desired product.

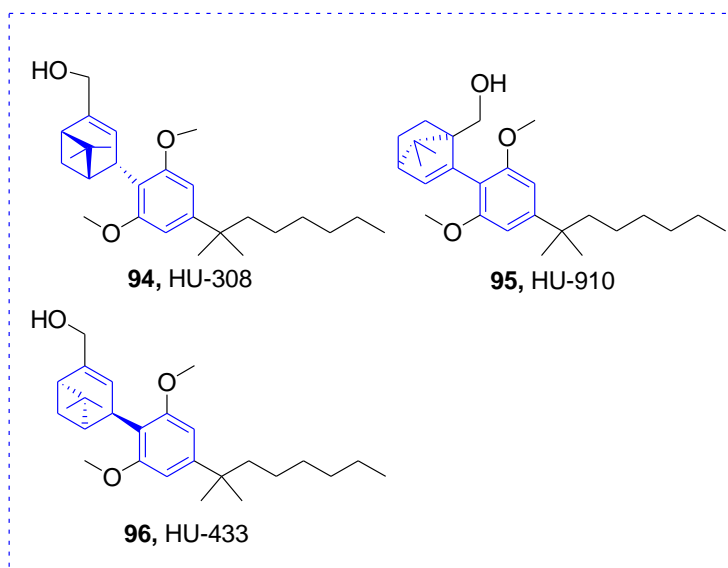
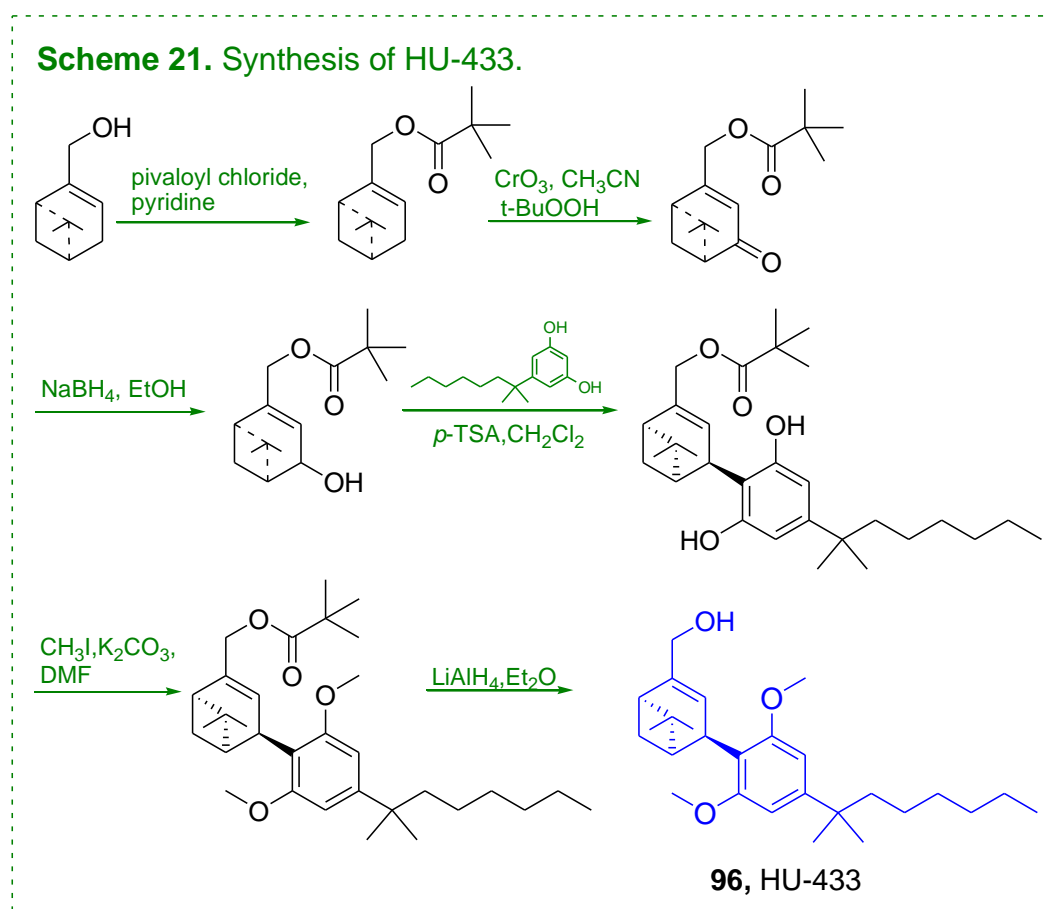


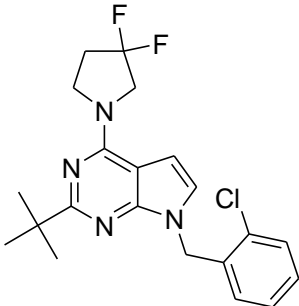
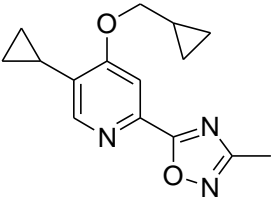
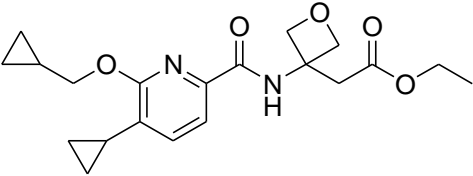
Figure 23. Analogs of HU-308.



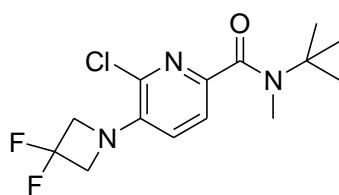
8. PATENTS

This section reports the patent literature from 2008 until today on novel molecules targeting CB₂ receptor that are currently studied in clinical trials or are candidates for future clinical evaluation in the management of diseases. The purpose is to show selected scenarios of chemical structures adopted in this field of research (Table 3).

Table 3. Scaffold-based CB₂ agonists in patent literature.

<i>Scaffold</i>	<i>Representative Compound</i>	<i>Biological Activity</i>	<i>Company name</i>
Pyrrolo[2,3d]pyrimidine ^a		EC ₅₀ = 10 nM	Hoffmann-La Roche
Pyridine ^b		EC ₅₀ = 26 nM	Hoffmann-La Roche
Pyridine ^c		K _i < 1 μM	Hoffmann-La Roche

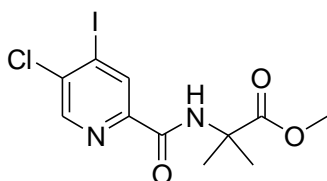
Pyridine^d



EC₅₀
= 5 nM

Hoffmann-La
Roche

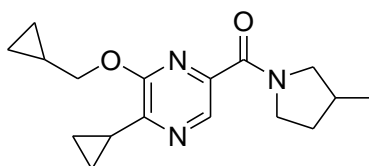
Pyridine^e



EC₅₀ = 36
nM

Hoffmann-La
Roche

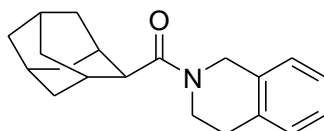
Pyrazinecarboxamide^f



EC₅₀ = 36
nM

Hoffmann-La
Roche

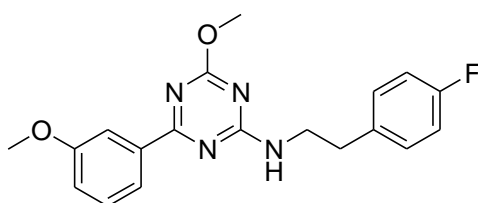
Adamantane^g



EC₅₀ = 7
nM

Hoffmann-La
Roche

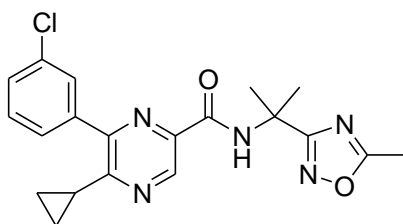
1,3,5-Triazine-2-amine^h



IC₅₀ < 500
nM

Sanofi

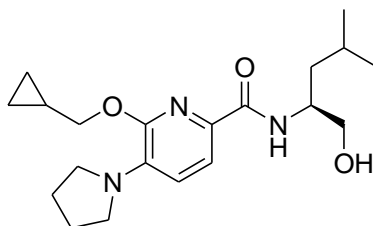
Pyrazineⁱ



IC₅₀ < 3
μM

Hoffmann-La
Roche

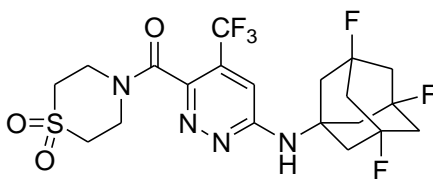
Pyridine^j



EC₅₀ = 3
nM

Hoffmann-La
Roche

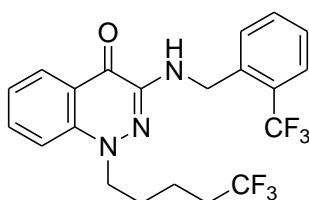
Pyridazin-3-amine^k



No data

Astellas Pharma

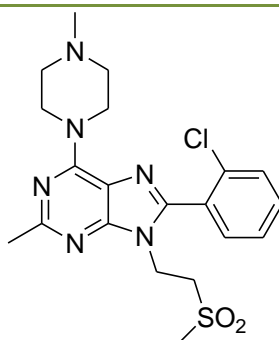
3-aminocinnolin-4(1H)-
one^l



IC₅₀ < 3
nM

Sanofi-Aventis

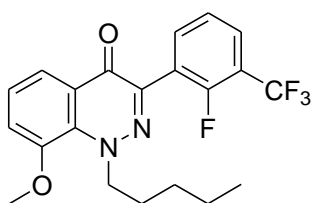
Purine^m



No data

Eli Lilly and
Company

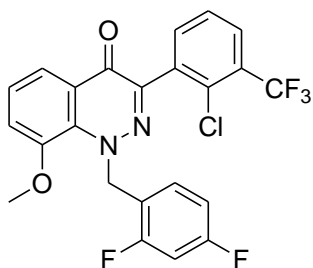
1-alkylcinnolin-4(1*H*)-
oneⁿ



IC₅₀ < 500
nM

Sanofi-Aventis

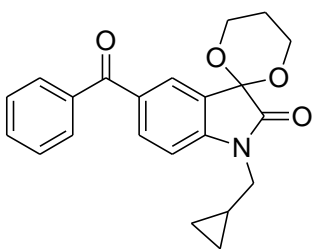
1-benzylcinnolin-4(1*H*)-
one^o



No data

Sanofi-Aventis

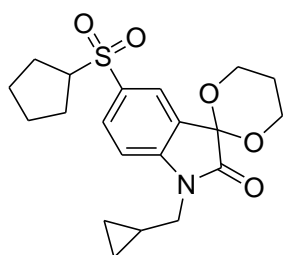
Oxindole^p



IC₅₀
ranging
from 0.062
to 20 μM

Wyeth LLC

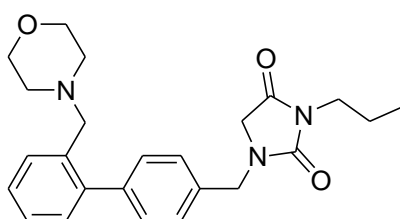
Oxindole^q



IC₅₀ = 54
nM

Wyeth LLC

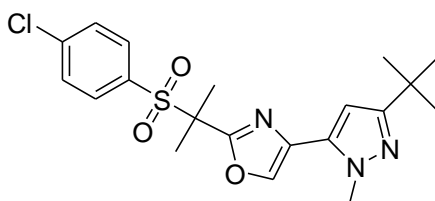
1-(biphenyl-4-
ylmethyl)imidazolidine-
2,4-dione^r



EC₅₀ < 0.4
nM

N.V. Organon

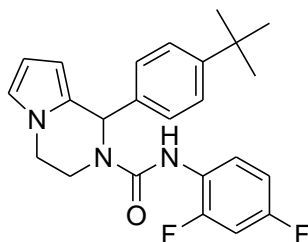
Sulfone^s



No data

Boehringer
Ingelheim
International
G.m.b.H.

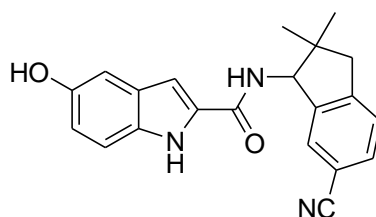
Diazepine^t



EC₅₀ < 1
μM

Kalypsys, Inc.

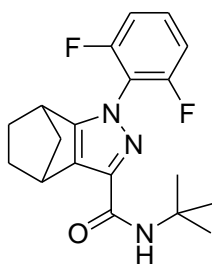
Indaylindole^u



EC₅₀ = 2.4
nM

Bristol-Myers
Squibb Company

Methanoindazole^v



No data

Glenmark
Pharmaceuticals,
S.A.

^aGrether, U.; Kimbara, A.; Nettekoven, M.; Roever, S.; Rogers-Evans, M.; Schulz-Gasch, T. WO2014177527, **2014**.^bGavelle, O.; Grether, U.; Kimbara, A.; Nettekoven, M.; Roever, S.; Rogers-Evans, M.; Rombach, D.; Schulz-Gasch, T. WO 2014154612, **2014**.^cFrei, B.; Gobbi, L.; Grether, U.; Kimbara, A.; Nettekoven, M.; Roever, S.; Rogers-Evans, M.; Schulz-Gasch, T. WO2014086705, **2014**.^dBendels, S.; Grether, U.; Kimbara, A.; Nettekoven, M.; Roever, S.; Rogers-Evans, M.; Schaffter, E.t; Schulz-Gasch, T. WO2014086806, **2014**.^eGrether, U.; Kimbara, A.; Nettekoven, M.; Ricklin, F.; Roever, S.; Rogers-Evans, M.; Rombach, D.; Schulz-Gasch, T.; Westphal, M. WO2014086805, **2014**.^fDhurwasulu, B.; Grether, U.; Nettekoven, M.; Roever, S.; Rogers-Evans, M.; Schulz-Gasch, T. WO2014086807, **2014**.^gKimbara, A.; Grether, U.; Nettekoven, M.; Puellmann, B.; Rogers-Evans, M.; Schulz-Gasch, T. WO2014005968, **2014**.^hArnaud, J.; Artiaga, M.; Barth, F.; Hortalá, L.; Martínez, S.; Roux, P. WO2013087643, **2013**.ⁱBissantz, C.; Dhurwasulu, B.; Grether, U.; Hazra, A.; Hebeisen, P.; Roever, S.; Rogers-Evans, M. WO2013060751, **2013**.^jBissantz, C.; Grether, U.; Hebeisen, P.; Kimbara, A.; Liu, Q.; Nettekoven, M.; Prunotto, M.; Roever, S.; Rogers-Evans, M.;

Schulz-Gasch, T.; Ullmer, C.; Wang, Z.; Yang, W. WO2012168350, **2012**. ^kMatsushima, Y.; Kubota, K.; Hamaguchi, T.; Okamoto, Y.; Terasawa, T.; Hondo, T.; Nishigaki, F. JP2012072067, **2012**. ^lLegeay, C.; Rinaldi-Carmona, M.; Roux, P.; Vernhet, C. WO2011033225, **2011**. ^mHollinshead, S. P. WO2011123482, **2011**. ⁿBarbagallo, E.; Legeay, C.; Rinaldi-Carmona, M.; Roux, P.; Vernhet, C. WO2010116084, **2010**. ^oBarbagallo, E.; Rinaldi-Carmona, M.; Roux, P.; Vernhet, C. WO2010004215, **2010**. ^pZhang, M.; Harrison, B. L.; Stanton, C. J.; Havran, L. M.; Chong, D. C.; Childers, W. E.; O'Neil, S. V. WO2010090680, **2010**. ^qDollings, P. J.; Donnell, A. F.; Gilbert, A. M.; Zhang, M.; Harrison, B. L.; Stanton, C. J.; O'Neil, S. V.; Havran, L. M.; Chong, D. C. WO2010077839, **2010**. ^rVan Der Stelt, M.; Cals, J. M. G. B. WO2010063721, **2010**. ^sHickey, E. R.; Riether, D.; Thomson, D. S.; Zindell, R. M.; Amouzegh, P.; Ermann, M.; Palmer, C. F.; Whittaker, M. WO2009140089, **2009**. ^tGahman, T. C.; Zhao, C.; Lang, H.; Massari, M. E. US20090062253, **2009**. ^uLiu, C.; Wroblewski, S. T.; Leftheris, K.; Wu, G.; Sher, P. M.; Ellsworth, B. A. US20090041722, **2009**. ^vShreedhar, N.; Shobha Rao, S. WO2008093194, **2008**.

9. POTENTIAL THERAPEUTIC APPLICATIONS OF CB₂ AGONISTS

The therapeutic value of CB₂ agonists has been well investigated for several years and medicinal chemistry research has identified a principal role of these compounds in the pharmacological treatment of pain and inflammation.^{11,13,105} Clinical applications of novel compounds have been complicated by several problems such as the presence of CB₂ receptors, that is very low in healthy but which significantly increases in the disease. It is, therefore, difficult to compare the effect of CB₂ agonists in animals with injury, illness or inflammatory conditions. Another problem is the CB₂ selectivity because, in the past, the older agonists at high doses were able to bind CB₁ receptors that are much more abundant compared to CB₂ receptors. As a consequence, an analysis of the CB₂ effects could be confused with CB₁ functional response, suggesting a possible explanation for the failure of the clinical trial. Moreover, it cannot be excluded that CB₂ agonists might interact with other membrane receptors involved in different diseases, like cancer or chronic pain.

In spite of all CB₂ agonists have progressed to various clinical trials by several pharmaceutical companies including PRS-211375 (Figure 24) from Pharmos Corporation; GW842166 (compound 1, Figure 3) and GW833972A (Figure 24) from GlaxoSmithKline; GRC10693 (Figure 24) from Glenmark Pharmaceuticals; JBT101 (Figure 24) from JB Therapeutics, Inc.; KHK6188 (structure not disclosed) from Kyowa Hakko Kirin; ABT521 (structure not disclosed) from Abbott Laboratories; LY2828360 (Figure 24) from Eli Lilly and Company; AZD1940 (Figure 24) from

Astra Zeneca; CR701 (structure not disclosed) from Cara Therapeutics, Inc.; APD 371 (structure not disclosed) from Arena Pharmaceuticals, Inc. and S-777469 (compound 7, Figure 3) from Shionogi & Company (Table 4).

A CB₂ selective cannabinoid library from Pharmos Corporation has shown promise in animal models for autoimmune inflammatory disorders, such as MS and inflammatory bowel disease. These compounds also have a good efficacy in animal models of neuropathic, inflammatory and chronic nociceptive pain. In selected preclinical models, these ligands have an analgesic activity equivalent better than Gabapentin. These CB₂-selective synthetic compounds belong to nonclassical CBs and display fewer of the undesired psychotropic and cardiovascular side-effects seen with some natural CBs. These compounds have been used in preclinical testing for neuropathic pain and the specific pharmacological properties have highlighted a wide range of beneficial therapeutic indications in common which could be useful, for example, as analgesic, neuroprotective, immunomodulatory and anti-inflammatory agents. As a consequence, CB₂ agonists could be important treatments in humans for analgesic activities and autoimmune diseases, such as MS and RA. Cannabinor (PRS-211375) was the first lead candidate to emerge from the chemical Pharmos technology used to experimental models of pain even if it failed in the inhibition of capsaicin-induced pain in Phase IIa trials.²⁹⁹ The effects of cannabinor were investigated in rats with partial urethral obstruction, treated daily for 14 days, and showed that the ability to empty the bladder was preserved and contraction frequency was low, if compared to those in controls, with a significant reduction in bladder weight.³⁰⁰ Detrusor preparations from cannabinor treated rats showed a higher response to nerve stimulation than those from controls, suggesting that cannabinor may be a novel principle to enable improved bladder function after partial urethral obstruction.³⁰¹ An additional study in Phase IIa trials was organized to evaluate CB₂ agonists used intravenously in a third molar dental extraction. Unfortunately, the results of this trial were confounding because this compound in the lowest dose showed significant effects, whereas higher doses failed to achieve significance.¹²

GlaxoSmithKline completed clinical trials by using selective CB₂ receptor agonists for dental pain, third molar tooth extraction and OA pain. Selective investigations of the structure activity relationships suggested an identification of several potent and selective agonists. These compounds showed antihyperalgesic effects in models of inflammatory and neuropathic pain, indicating a potential role in the treatment of chronic pain. A clinical program to evaluate the analgesic efficacy of GW842166 in acute dental pain failed to demonstrate either pharmacological and/or statistical significance.³⁰² In particular, the phase IIa study was conducted as a multi-centre study to evaluate the analgesic efficacy of pre-emptive doses of GW842166 following dental surgery after 3rd molar tooth extraction.³⁰³ Safety and tolerability were evaluated by adverse event monitoring, cardiovascular assessments and clinical laboratory tests such as haematology, clinical chemistry and urinalysis. Ibuprofen was significantly more effective than placebo across all endpoints and single doses of GW842166 (100 and 800 mg) failed to demonstrate clinically analgesia in the setting of acute dental pain. Moreover, GW833972A was able to inhibit Fos expression and reduces the number of trigeminal brainstem neurones activated by electrical tooth pulp stimulation caudally, but not rostrally, in animal models, suggesting an analgesic efficacy in dental pain. In addition, this compound inhibited capsaicin-induced depolarization in human and guinea-pig and prostaglandin E₂ (PGE₂) and hypertonic saline-induced depolarization of the guinea-pig isolated vagus nerve.¹⁶¹ Of interest, GW 833972A decreased citric acid-induced cough, but not plasma extravasation, in the guinea-pig, this effect being blocked by a CB₂ antagonist. This study highlights the role of CB₂ receptors in the modulation of sensory nerve activity elicited both by the exogenous ligands, capsaicin and hypertonic saline, but also by endogenous modulators, such as PGE₂ and low pH stimuli, suggesting that these receptors could be an interesting target for the treatment of chronic cough.¹⁶¹

Glenmark Pharmaceuticals reported successful completion of phase I trial in Europe with their candidate, GRC10693 (Tedralinab), demonstrating both safety and efficacy.³⁰⁴ GRC10693, a potent

and selective CB₂ agonist, is a drug developed for the treatment of OA and neuropathic pain. This compound has shown promise in the treatment of OA in phase I clinical trials. GRC 10693 was also investigated in neuropathic pain as primary indication in phase IIb clinical trials following oral route of administration.³⁰⁴

JB Therapeutics is a clinical stage biopharmaceutical company which has developed the first new class of analgesic agents in two decades. Their lead compound, JBT-101, is a potent CB agonist with demonstrated efficacy in several animal models of pain and inflammation. It has also completed a Phase II clinical study in chronic refractory neuropathic pain. JBT-101 has been developed as a potential disease modifying OA treatment. It has been demonstrated that the endocannabinoid system is up-regulated in pathologic fibrosis and that the modulation of CB receptors might limit the progression of uncontrolled fibrogenesis associated to scleroderma.³⁰⁵ JBT-101 also represents a platform for combination with other anti-inflammatory and analgesic agents as well as various drug delivery strategies including ointment and gels, topical patches, and timed release formulations.

A phase II Study of KHK6188 from Kyowa Hakko Kirin, a placebo-controlled trial, double blind in postherpetic neuralgia was carried out to evaluate the efficacy and safety of this orally administered compound.³⁰⁶ It is well reported that CBs could suppress neuropathic nociception in animal models of postherpetic neuralgia, and clinical studies largely affirm that neuropathic pain patients derive benefits from CB treatment.³⁰⁷

Abbott Company is investigating, in phase I clinical trials, the CB₂ agonist ABT-521 (possibly A-796260 or a similar compound with indole structure) for pain disorders.¹² Another CB₂ selective ligand A-836339 (Figure 24) was found to possess high hCB₂ affinity with a 425-fold selectivity versus hCB₁ but did not display significant affinity to other GPCRs.³⁰⁸ However, A-836339 exhibited selective analgesic, anti-inflammatory and anti-hyperalgesic effects even if typical cannabis-like effects appeared at higher doses despite its low binding affinity for CB₁ mediating a

significant decrease in spontaneous motor activity.^{309,310} It has been also reported that this compound could be a suitable target for the development of positron emission tomography (PET) radiotracers that could serve as imaging biomarkers in A β -induced neuroinflammation.³¹¹ The specificity of the PET signal was confirmed by the use of a specific CB₂ antagonist, AM630. The data collected in this study indicate that A β amyloidosis without concomitant tau pathology is sufficient to activate CB₂ receptors that are suitable as an imaging biomarker of neuroinflammation.⁸²

Eli Lilly and Company has carried out, in phase II clinical trials, the safety, efficacy and pharmacokinetics of a single daily oral dose of LY2828360 in OA knee pain.³¹² It is well reported that OA is a degenerative joint disease associated with articular cartilage degradation. The major clinical outcome of OA is a complex pain state that includes both nociceptive and neuropathic mechanisms. Currently, the therapeutic approaches for OA are limited to controlling disease progression and the analgesic treatment has restricted efficacy. Increasing evidence from preclinical studies supports the interest of the endocannabinoid system as an emerging therapeutic target for OA pain.¹⁰⁵ Indeed, pharmacological studies have shown the anti-nociceptive effects of CBs in different rodent models of OA, and compelling evidence suggests an active participation of these compounds in the disease pathophysiology. The ubiquitous distribution of CB receptors, together with the physiological role of the endocannabinoid system in the regulation of pain, inflammation and even joint function further support the therapeutic interest of CBs for OA.³¹²

From AstraZeneca derived a study that investigated the effects of AZD1940, a novel peripherally acting CB agonist, on capsaicin-induced pain and hyperalgesia, as well as on biomarkers of CNS effects.³¹³ This study was a randomized, double-blind, placebo-controlled, four-sequence, two-period, cross-over study in healthy male volunteers for postoperative pain. A phase II clinical trial showed the effects of two single oral doses of AZD1940 (400 and 800 μ g) that were compared with placebo. Pain intensity after intradermal capsaicin injections in the forearm was assessed on a

continuous visual analogue scale. Primary and secondary hyperalgesia induced by application of capsaicin cream on the calf were assessed by measuring heat pain thresholds and the area of mechanical allodynia, respectively. The CNS effects were assessed at baseline and up to 24 h after dosing using a visual analogue mood scale for various feeling. AZD1940 did not significantly attenuate ongoing pain or primary or secondary hyperalgesia compared with placebo. As part of the preclinical evaluation of AZD1940, a microdosing study with PET was conducted to assess brain exposure and the regional distribution of radioactivity within brain that was homogenous and low in comparison to peripheral organ exposure.³¹⁴ Treatment with AZD1940 mediated mild CNS effects while dose-dependent adverse events related to both CNS and gastrointestinal were reported. No evidence of analgesic efficacy for a peripherally acting CB agonist in human capsaicin pain model was found.¹⁰⁵

Cara Therapeutics Inc., a clinical-stage biopharmaceutical company, analyzed CR701 in a preclinical trial in order to treat neuropathic and inflammatory pain, and CR845, that has completed phase II clinical trials for the treatment of moderate-to-severe acute chronic pain.^{105,315}

APD371 from Arena Pharmaceutical Inc., an orally available agonist of CB₂ receptors, is a drug candidate intended for the treatment of pain.¹⁰⁵ Currently available CB agonists have been limited in utility by the psychotropic effects associated with the activation of CB₁, but not CB₂ receptors. Several novel, potent, CB₂-selective, lead compounds have been developed to retain the analgesic activity while avoiding the psychotropic side effects. There is a need for non-opioid (non-narcotic), non-steroidal anti-inflammatory drugs (NSAIDs) for novel approaches to acute or chronic pain. Narcotics involve problems of tolerance, addiction and abuse potential, whereas long term use of NSAIDs may be associated with stomach injury, bleeding, death, heart attacks, hypertension, and kidney injury.

Shionogi & Company investigated several novel CB₂ agonists and among them, the compound S-777469 or raclopride as a CB₂ selective agonist for clinical development. This compound is

considered to be a promising candidate for an orally active antipruritic effect since it significantly inhibited histamine-induced peripheral nerve firing in mice, suggesting that S-777469 produces its antipruritic effects by inhibiting itch signal transmission through CB₂ agonism.^{280,281} The metabolism and pharmacokinetics of S-777469 were investigated after a single oral administration of [¹⁴C]-S-777469 to healthy human subjects and showed a total radioactivity which was rapid and well absorbed in humans.³¹⁸

It is well reported that CB₂ receptors are abundant in splenocytes and leukocytes and responsible for many of the CB immunomodulatory and anti-inflammatory effects. CB₂ selective agonists are very attractive therapeutic agents without CB₁ mediated psychoactive effects because of their predominant peripheral distribution. Quantitative analysis of administered small molecule drugs such as S-777469 in biological tissues must be carried out through use of imaging mass spectrometry in order to understand the mechanisms underlying its therapeutic efficacy or toxicity.³¹⁹ Moreover, safety assessments of S-777469 have included adverse event monitoring, vital sign measurements, physical examination measurements, electrocardiograms assessments, and standard clinical laboratory safety tests such as hematology, blood chemistry, and urinalysis. Pharmacokinetic endpoints for each dose level of S-777469 based on the sampling schedule have been evaluated and suggest that this compound is rapidly and well absorbed with bioavailability values ranging from 50 to 70% in rats and dogs. Time required to achieve steady state and serum protein binding was determined for each dose level.³²⁰ Plasma and urine concentrations of S-777469 were determined for single dose or multiple doses in all examined cohorts. In vivo metabolism in rats and dogs showed good qualitative agreement with in vitro metabolism. Therefore, rats and dogs were thought to be appropriate species for non-clinical toxicity studies.³²¹ These data could be useful for the characterization of the pharmacokinetic properties of S-777469 and the estimation of its pharmacokinetic fate in humans. To assess the potential anti-pruritic and anti-inflammatory efficacy of S-777469 after single-dose administration and twice daily administration in subjects

with mild-to-moderate atopic dermatitis was investigated. The efficacy endpoints primarily included the physician's overall assessment and patient assessment of pruritus and indicated that the CB₂ selective agonists could have significant antipruritic effects.²⁴¹

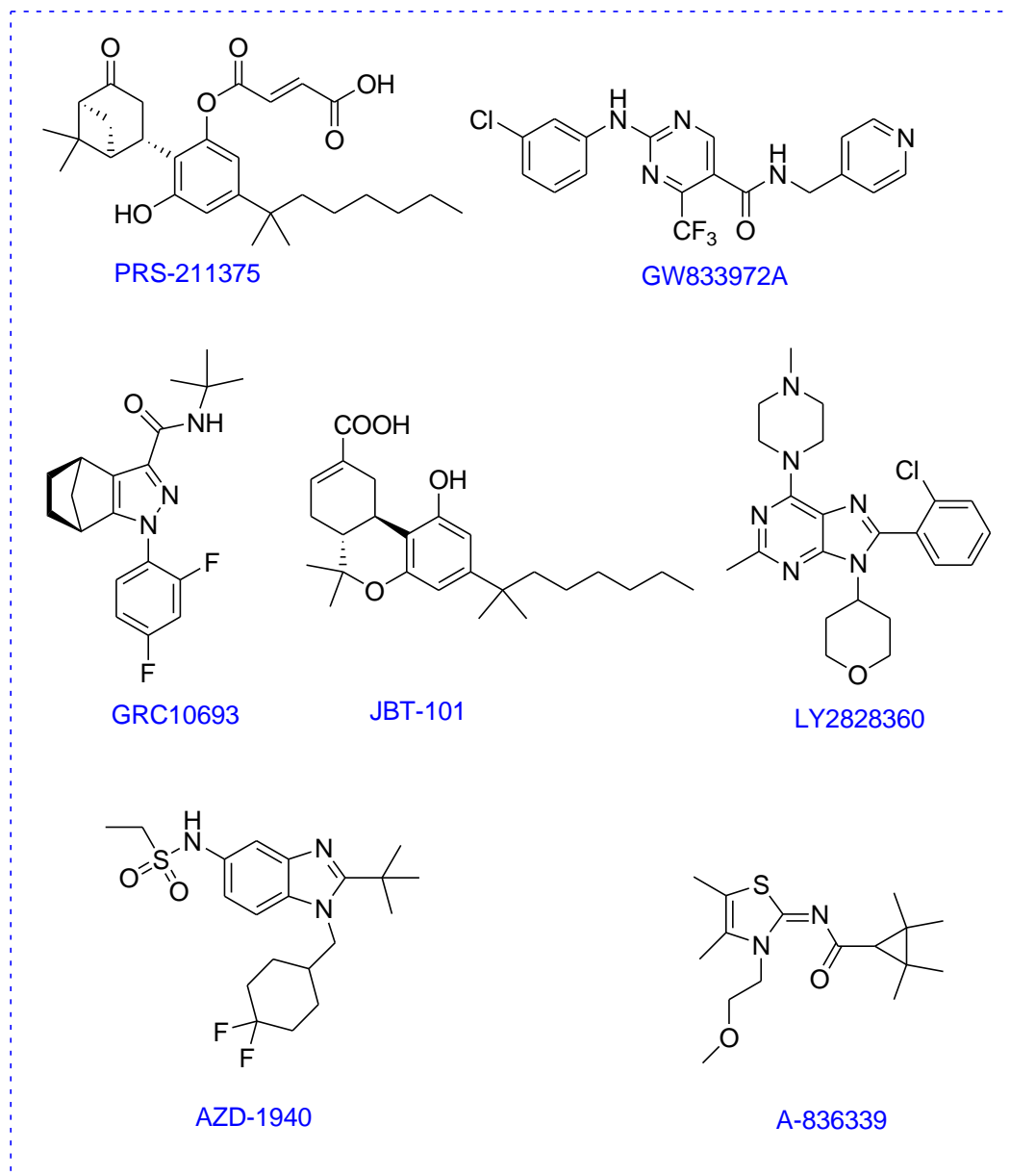


Figure 24. Selected CB₂ agonists in clinical development.

Table 4. Clinical studies of some of the most important CB₂ agonists.

<i>Compound</i>	<i>Therapeutic Implications</i>	<i>Clinical Status</i>	<i>Company</i>	<i>Source</i>
PRS-211375 (Cannabior)	Chronic pain	Phase 2a completed	Pharmos Corporation	WO 2006043260
GW842166	Osteoarthritis pain	Phase 2 completed	GlaxoSmithKline	NCT00447486
GRC10693 (Tedalinab)	Neuropathic pain, Osteoarthritis	Phase 1 Completed	Glenmark Pharmaceuticals	http://www.pipelinereview.com/
JBT101	Diffuse cutaneous systemic sclerosis	Phase 2	JB Therapeutics, CorbusPharmaceuticals	NCT02465437
KHK6188	Postherpetic neuralgia	Phase 2 completed	Kyowa Hakko Kirin	NCT01544296
ABT521	Pain	Phase 1	Abbott Laboratories	http://media.corporateir.net
LY 2828360	Knee pain, Osteoarthritis	Phase 2	Eli Lilly	NCT01319929
AZD1940	Pain	Phase 1	AstraZeneca	NCT00689780
CR701	Pain	Preclinical	Cara Therapeutics	http://www.caratherapeutics.com
CR845	Osteoarthritis	Phase 2	Cara Therapeutics	NCT02524197
APD 371	Pain	Phase 1	Arena Pharmaceuticals	http://www.arenapharma.com
S-777469	Atopic Dermatitis	Phase 2	Shionogi & Company	NCT00697710

10. FUTURE PERSPECTIVES AND CONCLUSIONS

The knowledge gained from several studies indicates the presence of CB₂ receptors in the brain and raises many questions about the possible roles that CB₂ receptors may play in the CNS. These results therefore extend previous evidence that CB₂ receptors play an important role in the immune function to other putative neuronal functions through their apparent presence in neuronal processes. As neuroinflammation is known to be associated with a number of autoimmune and neurological disorders, the close association of the immune system with CB₂ receptors and their functional expression in neurons requires a new evaluation of CB₂ receptors in mental disorders. Both CB₁ and CB₂ receptors seem likely to work both independently or in opposite directions and/or cooperatively in differing neuronal and/or glial cell populations to regulate important physiological activities in the CNS. Thus, more studies are required to determine the exact role of CB₂ receptors and the

nature of their interactions with other receptors or with CB₁ receptors in the brain, which may then determine the therapeutic utility of CB₂ ligands.

Taken together, the studies published in the literature point to potential therapeutic benefits of CB stimulation in the treatment of inflammatory status. Despite some differences in preclinical models and in the signaling of human and rodent CB₂ receptors, the development of selective CB₂ agonists may open new possibilities in therapeutic intervention. Such interventions would aim at reducing the release of pro-inflammatory mediators, particularly in chronic neuropathologic conditions, and delineate the therapeutic effects in neuropathic and chronic pain. Other potential applications for CB₂ agonists could be their use in atherosclerosis, allergic dermatitis, uveoretinitis, cough, gastrointestinal diseases and osteoporosis. A great interest appears to be the use of CB₂ agonists as anticancer agents in skin melanoma, colon cancer, breast and prostate tumor, scleroderma and different grades of lymphoma. Moreover, additional effects of CBs in cancer patients include inhibition of nausea and emesis associated to chemo- or radiotherapy, appetite stimulation, pain relief, mood elevation and insomnia reduction, all of which suggest meaningful clinical outcomes.

In conclusion, some issues remain to be clarified: a) the development of novel CB₂ agonists without CB₁ central side effects and their pharmacological and clinical characterization to specific pathological disorders; b) clinical and functional implication of neuronal CB₂ receptors in the brain and their contribution and interaction with CB₁ receptors in neuropsychiatry and in the neuroimmune activity; c) the mechanism and the presence of an allosteric site and the identification of allosteric modulator enhancers versus CB₂ receptors with the aim to limit the adverse side effects and to potentiate the effect of endocannabinoids; d) the characterization of homodimers and/or heterodimers for CB₂ receptors to better investigate drug-receptor interaction and the possible cross-talk between endocannabinoid system and other endogenous systems; e) identification of selected inhibitors on the endocannabinoid transport and enzyme inhibitors to augment the activation of CB₂ receptors from the endogenous CBs and highlight their autoprotective role in several disease states.

Several pharmaceutical companies and academic institutes have shown important scientific interest towards the chemical and pharmacological development of novel, potent and selective CB₂ agonists, in spite of the difficulties involved in marketing any new drug, which is expensive and full of regulatory and commercial obstacles. The relevant scientific and clinical advances illustrated in this review provide hope that CB₂ agonists will become manageable in some of the most common chronic diseases with the aim to improve human health.

AUTHOR INFORMATION

Corresponding Author

*E-mail: baraldi@unife.it. Phone: +39 0532455921. Fax: +39 0532455953

Notes

The authors declare no competing financial interest

Biographies



Mojgan Aghazadeh Tabrizi received her B.S. in Medicinal Chemistry and her Ph.D. in Pharmaceutical Sciences (1991) from Ferrara University, Italy. She has worked at the Department of Pharmaceutical Sciences of Ferrara University focusing her interests on the design and synthesis of adenosine receptor ligands and antitumor compounds in collaboration with the company King Pharmaceuticals (North Carolina, U.S.A.), now part of Pfizer. Since 2008, she has been part of a research project on the treatment of pain and inflammation involving the Endocannabinoid System. She is author/co-author of several scientific publications in the field of medicinal chemistry.



Pier Giovanni Baraldi received his degree in Chemistry in 1974 from the University of Ferrara where he is currently Full Professor of Medicinal Chemistry. He has published more than 400 scientific papers including about 50 patents focusing his research activity on the design and synthesis of DNA minor groove alkylating agents, combretastatin analogs, and ligands for ARs and TRP channel modulators. He has presented more than 90 lectures as speaker in international congresses. He was founder with Prof. Pier Andrea Borea of the spin off Pharmeste involved in the discovery of TRPV1 compounds.



Pier Andrea Borea received his degree in Chemistry from the University of Ferrara in 1967. He is currently President of the Evaluation Board of the University of Ferrara. He has published 400 publications in international journals and contributed about 20 chapters to International Books. He has participated in more than 200 congresses, presenting main, oral and poster communications. He is also named as inventor in about 20 Patents. He has been invited to give about 50 Plenary Lectures at International Symposia and at several Universities. His main field of interest is represented by the study of drug-receptor interactions at molecular level.



Katia Varani received her Doctor degree in Biology in 1988 and her Ph.D. degree in Cellular and Molecular Pharmacology in 1995 from the University of Ferrara. She is currently Associate Professor of Pharmacology at the Medical Sciences Department of the University of Ferrara. She is author/co-author of several scientific publications involving various topics such as the following: pharmacological, biochemical and molecular study of adenosine receptors; interactions of G-protein coupled receptors and human neurodegenerative diseases; chronic and neuropathic pain, cardiovascular disorders; inflammatory diseases and cancer.

REFERENCES

- (1) Di Marzo, V.; Bifulco, M.; De Petrocellis, L. The Endocannabinoid System and Its Therapeutic Exploitation. *Nat. Rev. Drug Discov.* **2004**, *3* (9), 771–784.
- (2) Ahn, K.; McKinney, M. K.; Cravatt, B. F. Enzymatic Pathways That Regulate Endocannabinoid Signaling in the Nervous System. *Chem. Rev.* **2008**, *108* (5), 1687–1707.
- (3) Di Marzo, V.; Bisogno, T.; De Petrocellis, L. Endocannabinoids and Related Compounds: Walking Back and Forth between Plant Natural Products and Animal Physiology. *Chem. Biol.* **2007**, *14* (7), 741–756.
- (4) Di Iorio, G.; Lupi, M.; Sarchione, F.; Matarazzo, I.; Santacroce, R.; Petrucci, F.; Martinotti, G.; Di Giannantonio, M. The Endocannabinoid System: A Putative Role in Neurodegenerative Diseases. *Int. J. high risk Behav. Addict.* **2013**, *2* (3), 100–106.
- (5) Makriyannis, A. 2012 Division of Medicinal Chemistry Award Address. Trekking the Cannabinoid Road: A Personal Perspective. *J. Med. Chem.* **2014**, *57* (10), 3891–3911.
- (6) Howlett, A. C.; Barth, F.; Bonner, T. I.; Cabral, G.; Casellas, P.; Devane, W. A.; Felder, C. C.; Herkenham, M.; Mackie, K.; Martin, B. R.; et al. International Union of Pharmacology. XXVII. Classification of Cannabinoid Receptors. *Pharmacol. Rev.* **2002**, *54* (2), 161–202.
- (7) Piomelli, D. The Molecular Logic of Endocannabinoid Signalling. *Nat. Rev. Neurosci.* **2003**, *4* (11), 873–884.
- (8) Whiteside, G. T.; Lee, G. P.; Valenzano, K. J. The Role of the Cannabinoid CB2 Receptor in Pain Transmission and Therapeutic Potential of Small Molecule CB2 Receptor Agonists. *Curr. Med. Chem.* **2007**, *14* (8), 917–936.
- (9) Malfitano, A. M.; Basu, S.; Maresz, K.; Bifulco, M.; Dittel, B. N. What We Know and Do Not Know about the Cannabinoid Receptor 2 (CB2). *Semin. Immunol.* **2014**, *26* (5), 369–379.
- (10) Pertwee, R. G. Targeting the Endocannabinoid System with Cannabinoid Receptor Agonists: Pharmacological Strategies and Therapeutic Possibilities. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* **2012**, *367* (1607), 3353–3363.
- (11) Picone, R. P.; Kendall, D. A. Minireview: From the Bench, toward the Clinic: Therapeutic Opportunities for Cannabinoid Receptor Modulation. *Mol. Endocrinol.* **2015**, *29* (6), 801–813.
- (12) Nevalainen, T. Recent Development of CB2 Selective and Peripheral CB1/CB2 Cannabinoid Receptor Ligands. *Curr. Med. Chem.* **2014**, *21* (2), 187–203.
- (13) Vemuri, V. K.; Makriyannis, A. Medicinal Chemistry of Cannabinoids. *Clin. Pharmacol. Ther.* **2015**, *97* (6), 553–558.

- (14) Baraldi, P. G.; Saponaro, G.; Moorman, A. R.; Romagnoli, R.; Preti, D.; Baraldi, S.; Ruggiero, E.; Varani, K.; Targa, M.; Vincenzi, F.; et al. 7-Oxo-[1,4]oxazino[2,3,4-Ij]quinoline-6-Carboxamides as Selective CB(2) Cannabinoid Receptor Ligands: Structural Investigations around a Novel Class of Full Agonists. *J. Med. Chem.* **2012**, *55* (14), 6608–6623.
- (15) Aghazadeh Tabrizi, M.; Baraldi, P. G.; Saponaro, G.; Moorman, A. R.; Romagnoli, R.; Preti, D.; Baraldi, S.; Corciulo, C.; Vincenzi, F.; Borea, P. A.; et al. Design, Synthesis, and Pharmacological Properties of New Heteroarylpyridine/heteroarylpyrimidine Derivatives as CB(2) Cannabinoid Receptor Partial Agonists. *J. Med. Chem.* **2012**, *56* (3), 1098–1112.
- (16) Aghazadeh Tabrizi, M.; Baraldi, P. G.; Saponaro, G.; Moorman, A. R.; Romagnoli, R.; Preti, D.; Baraldi, S.; Ruggiero, E.; Tintori, C.; Tuccinardi, T.; et al. Discovery of 7-oxopyrazolo[1,5-A]pyrimidine-6-Carboxamides as Potent and Selective CB(2) Cannabinoid Receptor Inverse Agonists. *J. Med. Chem.* **2013**, *56* (11), 4482–4496.
- (17) Onaivi, E. S.; Leonard, C. M.; Ishiguro, H.; Zhang, P. W.; Lin, Z.; Akinshola, B. E.; Uhl, G. R. Endocannabinoids and Cannabinoid Receptor Genetics. *Prog. Neurobiol.* **2002**, *66* (5), 307–344.
- (18) Turcotte, C.; Chouinard, F.; Lefebvre, J. S.; Flamand, N. Regulation of Inflammation by Cannabinoids, the Endocannabinoids 2-Arachidonoyl-Glycerol and Arachidonoyl-Ethanolamide, and Their Metabolites. *J. Leukoc. Biol.* **2015**, *97* (6), 1049–1070.
- (19) Fratta, W.; Fattore, L. Molecular Mechanisms of Cannabinoid Addiction. *Curr. Opin. Neurobiol.* **2013**, *23* (4), 487–492.
- (20) Pertwee, R. G.; Ross, R. A. Cannabinoid Receptors and Their Ligands. *Prostaglandins. Leukot. Essent. Fatty Acids* **2002**, *66* (2-3), 101–121.
- (21) Davies, S. N.; Pertwee, R. G.; Riedel, G. Functions of Cannabinoid Receptors in the Hippocampus. *Neuropharmacology* **2002**, *42* (8), 993–1007.
- (22) Pertwee, R. G.; Howlett, A. C.; Abood, M. E.; Alexander, S. P. H.; Di Marzo, V.; Elphick, M. R.; Greasley, P. J.; Hansen, H. S.; Kunos, G.; Mackie, K.; et al. International Union of Basic and Clinical Pharmacology. LXXIX. Cannabinoid Receptors and Their Ligands: Beyond CB₁ and CB₂. *Pharmacol. Rev.* **2010**, *62* (4), 588–631.
- (23) Howlett, A. C.; Barth, F.; Bonner, T. I.; Cabral, G.; Casellas, P.; Devane, W. A.; Felder, C. C.; Herkenham, M.; Mackie, K.; Martin, B. R.; et al. International Union of Pharmacology. XXVII. Classification of Cannabinoid Receptors. *Pharmacol. Rev.* **2002**, *54* (2), 161–202.
- (24) Griffin, G.; Tao, Q.; Abood, M. E. Cloning and Pharmacological Characterization of the Rat CB(2) Cannabinoid Receptor. *J. Pharmacol. Exp. Ther.* **2000**, *292* (3), 886–894.
- (25) Feng, W.; Song, Z. H. Functional Roles of the Tyrosine within the NP(X)(n)Y Motif and the Cysteines in the C-Terminal Juxtamembrane Region of the CB2 Cannabinoid Receptor. *FEBS Lett.* **2001**, *501* (2-3), 166–170.

- (26) Reggio, P. H. Cannabinoid Receptors and Their Ligands: Ligand-Ligand and Ligand-Receptor Modeling Approaches. *Handb. Exp. Pharmacol.* **2005**, No. 168, 247–281.
- (27) Liu, Q.-R.; Pan, C.-H.; Hishimoto, A.; Li, C.-Y.; Xi, Z.-X.; Llorente-Berzal, A.; Viveros, M.-P.; Ishiguro, H.; Arinami, T.; Onaivi, E. S.; et al. Species Differences in Cannabinoid Receptor 2 (CNR2 Gene): Identification of Novel Human and Rodent CB2 Isoforms, Differential Tissue Expression and Regulation by Cannabinoid Receptor Ligands. *Genes. Brain. Behav.* **2009**, 8 (5), 519–530.
- (28) Howlett, A. C.; Breivogel, C. S.; Childers, S. R.; Deadwyler, S. A.; Hampson, R. E.; Porrino, L. J. Cannabinoid Physiology and Pharmacology: 30 Years of Progress. *Neuropharmacology* **2004**, 47 Suppl 1, 345–358.
- (29) Howlett, A. C. The Cannabinoid Receptors. *Prostaglandins Other Lipid Mediat.* **2002**, 68-69, 619–631.
- (30) Dalton, G. D.; Bass, C. E.; Van Horn, C. G.; Howlett, A. C. Signal Transduction via Cannabinoid Receptors. *CNS Neurol. Disord. Drug Targets* **2009**, 8 (6), 422–431.
- (31) Thompson, M. D.; Cole, D. E. C.; Capra, V.; Siminovitch, K. A.; Rovati, G. E.; Burnham, W. M.; Rana, B. K. Pharmacogenetics of the G Protein-Coupled Receptors. *Methods Mol. Biol.* **2014**, 1175, 189–242.
- (32) Birdsall, N. J. M. Class A GPCR Heterodimers: Evidence from Binding Studies. *Trends Pharmacol. Sci.* **2010**, 31 (11), 499–508.
- (33) Perrey, D. A.; Gilmour, B. P.; Thomas, B. F.; Zhang, Y. Toward the Development of Bivalent Ligand Probes of Cannabinoid CB1 and Orexin OX1 Receptor Heterodimers. *ACS Med. Chem. Lett.* **2014**, 5 (6), 634–638.
- (34) Bagher, A. M.; Laprairie, R. B.; Kelly, M. E. M.; Denovan-Wright, E. M. Co-Expression of the Human Cannabinoid Receptor Coding Region Splice Variants (hCB₁) Affects the Function of hCB₁ Receptor Complexes. *Eur. J. Pharmacol.* **2013**, 721 (1-3), 341–354.
- (35) Bauer, M.; Chicca, A.; Tamborini, M.; Eisen, D.; Lerner, R.; Lutz, B.; Poetz, O.; Pluschke, G.; Gertsch, J. Identification and Quantification of a New Family of Peptide Endocannabinoids (Pepcans) Showing Negative Allosteric Modulation at CB1 Receptors. *J. Biol. Chem.* **2012**, 287 (44), 36944–36967.
- (36) Khurana, L.; Ali, H. I.; Olszewska, T.; Ahn, K. H.; Damaraju, A.; Kendall, D. A.; Lu, D. Optimization of Chemical Functionalities of Indole-2-Carboxamides to Improve Allosteric Parameters for the Cannabinoid Receptor 1 (CB1). *J. Med. Chem.* **2014**, 57 (7), 3040–3052.
- (37) Onaivi, E. S. Cannabinoid Receptors in Brain: Pharmacogenetics, Neuropharmacology, Neurotoxicology, and Potential Therapeutic Applications. *Int. Rev. Neurobiol.* **2009**, 88, 335–369.
- (38) Brown, S. M.; Wager-Miller, J.; Mackie, K. Cloning and Molecular Characterization of the Rat CB2 Cannabinoid Receptor. *Biochim. Biophys. Acta* **2002**, 1576 (3), 255–264.

- (39) Ofek, O.; Karsak, M.; Leclerc, N.; Fogel, M.; Frenkel, B.; Wright, K.; Tam, J.; Attar-Namdar, M.; Kram, V.; Shohami, E.; et al. Peripheral Cannabinoid Receptor, CB2, Regulates Bone Mass. *Proc. Natl. Acad. Sci. U. S. A.* **2006**, *103* (3), 696–701.
- (40) Julien, B.; Grenard, P.; Teixeira-Clerc, F.; Van Nhieu, J. T.; Li, L.; Karsak, M.; Zimmer, A.; Mallat, A.; Lotersztajn, S. Antifibrogenic Role of the Cannabinoid Receptor CB2 in the Liver. *Gastroenterology* **2005**, *128* (3), 742–755.
- (41) Duncan, M.; Mouihate, A.; Mackie, K.; Keenan, C. M.; Buckley, N. E.; Davison, J. S.; Patel, K. D.; Pittman, Q. J.; Sharkey, K. A. Cannabinoid CB2 Receptors in the Enteric Nervous System Modulate Gastrointestinal Contractility in Lipopolysaccharide-Treated Rats. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2008**, *295* (1), G78–G87.
- (42) Atwood, B. K.; Mackie, K. CB2: A Cannabinoid Receptor with an Identity Crisis. *Br. J. Pharmacol.* **2010**, *160* (3), 467–479.
- (43) Price, D. A.; Martinez, A. A.; Seillier, A.; Koek, W.; Acosta, Y.; Fernandez, E.; Strong, R.; Lutz, B.; Marsicano, G.; Roberts, J. L.; et al. WIN55,212-2, a Cannabinoid Receptor Agonist, Protects against Nigrostriatal Cell Loss in the 1-Methyl-4-Phenyl-1,2,3,6-Tetrahydropyridine Mouse Model of Parkinson's Disease. *Eur. J. Neurosci.* **2009**, *29* (11), 2177–2186.
- (44) Pietr, M.; Kozela, E.; Levy, R.; Rimmerman, N.; Lin, Y. H.; Stella, N.; Vogel, Z.; Juknat, A. Differential Changes in GPR55 during Microglial Cell Activation. *FEBS Lett.* **2009**, *583* (12), 2071–2076.
- (45) Berdyshev, E. V. Cannabinoid Receptors and the Regulation of Immune Response. *Chem. Phys. Lipids* **2000**, *108* (1-2), 169–190.
- (46) Oz, M. Receptor-Independent Effects of Endocannabinoids on Ion Channels. *Curr. Pharm. Des.* **2006**, *12* (2), 227–239.
- (47) Howlett, A. C. Cannabinoid Receptor Signaling. *Handb. Exp. Pharmacol.* **2005**, No. 168, 53–79.
- (48) Galve-Roperh, I.; Chiurchiù, V.; Díaz-Alonso, J.; Bari, M.; Guzmán, M.; Maccarrone, M. Cannabinoid Receptor Signaling in Progenitor/stem Cell Proliferation and Differentiation. *Prog. Lipid Res.* **2013**, *52* (4), 633–650.
- (49) Upham, B. L.; Rummel, A. M.; Carbone, J. M.; Trosko, J. E.; Ouyang, Y.; Crawford, R. B.; Kaminski, N. E. Cannabinoids Inhibit Gap Junctional Intercellular Communication and Activate ERK in a Rat Liver Epithelial Cell Line. *Int. J. Cancer* **2003**, *104* (1), 12–18.
- (50) Merighi, S.; Gessi, S.; Varani, K.; Simioni, C.; Fazzi, D.; Mirandola, P.; Borea, P. A. Cannabinoid CB(2) Receptors Modulate ERK-1/2 Kinase Signalling and NO Release in Microglial Cells Stimulated with Bacterial Lipopolysaccharide. *Br. J. Pharmacol.* **2012**, *165* (6), 1773–1788.

- (51) Cunha, T. M.; Roman-Campos, D.; Lotufo, C. M.; Duarte, H. L.; Souza, G. R.; Verri, W. A.; Funez, M. I.; Dias, Q. M.; Schivo, I. R.; Domingues, A. C.; et al. Morphine Peripheral Analgesia Depends on Activation of the PI3Kgamma/AKT/nNOS/NO/KATP Signaling Pathway. *Proc. Natl. Acad. Sci. U. S. A.* **2010**, *107* (9), 4442–4447.
- (52) Murphy, S. Production of Nitric Oxide by Glial Cells: Regulation and Potential Roles in the CNS. *Glia* **2000**, *29* (1), 1–13.
- (53) Merighi, S.; Gessi, S.; Varani, K.; Fazzi, D.; Mirandola, P.; Borea, P. A. Cannabinoid CB(2) Receptor Attenuates Morphine-Induced Inflammatory Responses in Activated Microglial Cells. *Br. J. Pharmacol.* **2012**, *166* (8), 2371–2385.
- (54) Guzmán, M.; Galve-Roperh, I.; Sánchez, C. Ceramide: A New Second Messenger of Cannabinoid Action. *Trends Pharmacol. Sci.* **2001**, *22* (1), 19–22.
- (55) Morel, L. J.; Giros, B.; Daugé, V. Adolescent Exposure to Chronic Delta-9-Tetrahydrocannabinol Blocks Opiate Dependence in Maternally Deprived Rats. *Neuropsychopharmacology* **2009**, *34* (11), 2469–2476.
- (56) Parolaro, D.; Rubino, T.; Viganò, D.; Massi, P.; Guidali, C.; Realini, N. Cellular Mechanisms Underlying the Interaction between Cannabinoid and Opioid System. *Curr. Drug Targets* **2010**, *11* (4), 393–405.
- (57) Zoppi, S.; Madrigal, J. L.; Caso, J. R.; García-Gutiérrez, M. S.; Manzanares, J.; Leza, J. C.; García-Bueno, B. Regulatory Role of the Cannabinoid CB2 Receptor in Stress-Induced Neuroinflammation in Mice. *Br. J. Pharmacol.* **2014**, *171* (11), 2814–2826.
- (58) Gui, H.; Liu, X.; Wang, Z.-W.; He, D.-Y.; Su, D.-F.; Dai, S.-M. Expression of Cannabinoid Receptor 2 and Its Inhibitory Effects on Synovial Fibroblasts in Rheumatoid Arthritis. *Rheumatology (Oxford)*. **2014**, *53* (5), 802–809.
- (59) Guillot, A.; Hamdaoui, N.; Bizy, A.; Zoltani, K.; Souktani, R.; Zafrani, E.-S.; Mallat, A.; Lotersztajn, S.; Lafdil, F. Cannabinoid Receptor 2 Counteracts Interleukin-17-Induced Immune and Fibrogenic Responses in Mouse Liver. *Hepatology* **2014**, *59* (1), 296–306.
- (60) Wilkerson, J. L.; Gentry, K. R.; Dengler, E. C.; Wallace, J. A.; Kerwin, A. A.; Armijo, L. M.; Kuhn, M. N.; Thakur, G. A.; Makriyannis, A.; Milligan, E. D. Intrathecal Cannabidiol CB(2)R Agonist, AM1710, Controls Pathological Pain and Restores Basal Cytokine Levels. *Pain* **2012**, *153* (5), 1091–1106.
- (61) Gao, F.; Zhang, L.-H.; Su, T.-F.; Li, L.; Zhou, R.; Peng, M.; Wu, C.-H.; Yuan, X.-C.; Sun, N.; Meng, X.-F.; et al. Signaling Mechanism of Cannabinoid Receptor-2 Activation-Induced B-Endorphin Release. *Mol. Neurobiol.* **2015**.
- (62) De Petrocellis, L.; Cascio, M. G.; Di Marzo, V. The Endocannabinoid System: A General View and Latest Additions. *Br. J. Pharmacol.* **2004**, *141* (5), 765–774.

- (63) Maccarrone, M.; Bab, I.; Bíró, T.; Cabral, G. A.; Dey, S. K.; Di Marzo, V.; Konje, J. C.; Kunos, G.; Mechoulam, R.; Pacher, P.; et al. Endocannabinoid Signaling at the Periphery: 50 Years after THC. *Trends Pharmacol. Sci.* **2015**, *36* (5), 277–296.
- (64) Pagotto, U.; Marsicano, G.; Cota, D.; Lutz, B.; Pasquali, R. The Emerging Role of the Endocannabinoid System in Endocrine Regulation and Energy Balance. *Endocr. Rev.* **2006**, *27* (1), 73–100.
- (65) Wang, J.; Ueda, N. Biology of Endocannabinoid Synthesis System. *Prostaglandins Other Lipid Mediat.* **2009**, *89* (3-4), 112–119.
- (66) Bisogno, T.; Ligresti, A.; Di Marzo, V. The Endocannabinoid Signalling System: Biochemical Aspects. *Pharmacol. Biochem. Behav.* **2005**, *81* (2), 224–238.
- (67) Schmid, H. H. O.; Berdyshev, E. V. Cannabinoid Receptor-Inactive N-Acylethanolamines and Other Fatty Acid Amides: Metabolism and Function. *Prostaglandins. Leukot. Essent. Fatty Acids* **2002**, *66* (2-3), 363–376.
- (68) Bisogno, T.; De Petrocellis, L.; Di Marzo, V. Fatty Acid Amide Hydrolase, an Enzyme with Many Bioactive Substrates. Possible Therapeutic Implications. *Curr. Pharm. Des.* **2002**, *8* (7), 533–547.
- (69) Serra, G.; Bogliolo, V.; Garzia, A.; Zazzara, F.; Koukopoulos, A.; Demontis, F. The Role of the Endocannabinoid System in the Pathophysiology of Mood Disorders. *J. Psychopathol.* **2011**, *17*, 78–91.
- (70) Fowler, C. J. The Cannabinoid System and Its Pharmacological Manipulation--a Review, with Emphasis upon the Uptake and Hydrolysis of Anandamide. *Fundam. Clin. Pharmacol.* **2006**, *20* (6), 549–562.
- (71) Naidoo, V.; Karanian, D. A.; Vadivel, S. K.; Locklear, J. R.; Wood, J. T.; Nasr, M.; Quizon, P. M. P.; Graves, E. E.; Shukla, V.; Makriyannis, A.; et al. Equipotent Inhibition of Fatty Acid Amide Hydrolase and Monoacylglycerol Lipase - Dual Targets of the Endocannabinoid System to Protect against Seizure Pathology. *Neurotherapeutics* **2012**, *9* (4), 801–813.
- (72) Hwang, J.; Adamson, C.; Butler, D.; Janero, D. R.; Makriyannis, A.; Bahr, B. A. Enhancement of Endocannabinoid Signaling by Fatty Acid Amide Hydrolase Inhibition: A Neuroprotective Therapeutic Modality. *Life Sci.* **2010**, *86* (15-16), 615–623.
- (73) Lichtman, A. H.; Leung, D.; Shelton, C. C.; Saghatelian, A.; Hardouin, C.; Boger, D. L.; Cravatt, B. F. Reversible Inhibitors of Fatty Acid Amide Hydrolase That Promote Analgesia: Evidence for an Unprecedented Combination of Potency and Selectivity. *J. Pharmacol. Exp. Ther.* **2004**, *311* (2), 441–448.
- (74) Ross, R. A. Anandamide and Vanilloid TRPV1 Receptors. *Br. J. Pharmacol.* **2003**, *140* (5), 790–801.
- (75) Di Marzo, V.; De Petrocellis, L. Why Do Cannabinoid Receptors Have More than One Endogenous Ligand? *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* **2012**, *367* (1607), 3216–3228.

- (76) Malek, N.; Popiolek-Barczyk, K.; Mika, J.; Przewlocka, B.; Starowicz, K. Anandamide, Acting via CB2 Receptors, Alleviates LPS-Induced Neuroinflammation in Rat Primary Microglial Cultures. *Neural Plast.* **2015**, *2015*, 130639.
- (77) Karaliota, S.; Siafaka-Kapadai, A.; Gontinou, C.; Psarra, K.; Mavri-Vavayanni, M. Anandamide Increases the Differentiation of Rat Adipocytes and Causes PPAR γ and CB1 Receptor Upregulation. *Obesity (Silver Spring)*. **2009**, *17* (10), 1830–1838.
- (78) Miller, A. M.; Stella, N. CB2 Receptor-Mediated Migration of Immune Cells: It Can Go Either Way. *Br. J. Pharmacol.* **2008**, *153* (2), 299–308.
- (79) Ronco, A. M.; Llanos, M.; Tamayo, D.; Hirsch, S. Anandamide Inhibits Endothelin-1 Production by Human Cultured Endothelial Cells: A New Vascular Action of This Endocannabinoid. *Pharmacology* **2007**, *79* (1), 12–16.
- (80) Maldonado, R.; Berrendero, F.; Ozaita, A.; Robledo, P. Neurochemical Basis of Cannabis Addiction. *Neuroscience* **2011**, *181*, 1–17.
- (81) Aso, E.; Ferrer, I. Cannabinoids for Treatment of Alzheimer's Disease: Moving toward the Clinic. *Front. Pharmacol.* **2014**, *5*, 37.
- (82) Savonenko, A. V.; Melnikova, T.; Wang, Y.; Ravert, H.; Gao, Y.; Koppel, J.; Lee, D.; Pletnikova, O.; Cho, E.; Sayyida, N.; et al. Cannabinoid CB2 Receptors in a Mouse Model of A β Amyloidosis: Immunohistochemical Analysis and Suitability as a PET Biomarker of Neuroinflammation. *PLoS One* **2015**, *10* (6), e0129618.
- (83) Rogers, N. Cannabinoid Receptor with an “Identity Crisis” Gets a Second Look. *Nat. Med.* **2015**, *21* (9), 966–967.
- (84) Sipe, J. C.; Arbour, N.; Gerber, A.; Beutler, E. Reduced Endocannabinoid Immune Modulation by a Common Cannabinoid 2 (CB2) Receptor Gene Polymorphism: Possible Risk for Autoimmune Disorders. *J. Leukoc. Biol.* **2005**, *78* (1), 231–238.
- (85) Karsak, M.; Cohen-Solal, M.; Freudenberg, J.; Ostertag, A.; Morieux, C.; Kornak, U.; Essig, J.; Erxlebe, E.; Bab, I.; Kubisch, C.; et al. Cannabinoid Receptor Type 2 Gene Is Associated with Human Osteoporosis. *Hum. Mol. Genet.* **2005**, *14* (22), 3389–3396.
- (86) Borea, P. A.; Dalpiaz, A.; Varani, K.; Gilli, P.; Gilli, G. Can Thermodynamic Measurements of Receptor Binding Yield Information on Drug Affinity and Efficacy? *Biochem. Pharmacol.* **2000**, *60* (11), 1549–1556.
- (87) Merighi, S.; Simioni, C.; Gessi, S.; Varani, K.; Borea, P. A. Binding Thermodynamics at the Human Cannabinoid CB1 and CB2 Receptors. *Biochem. Pharmacol.* **2010**, *79* (3), 471–477.
- (88) Varani, K.; Merighi, S.; Gessi, S.; Klotz, K. N.; Leung, E.; Baraldi, P. G.; Cacciari, B.; Romagnoli, R.; Spalluto, G.; Borea, P. A. [(3)H]MRE 3008F20: A Novel Antagonist Radioligand for the Pharmacological and Biochemical Characterization of Human A(3) Adenosine Receptors. *Mol. Pharmacol.* **2000**, *57* (5), 968–975.

- (89) Merighi, S.; Varani, K.; Gessi, S.; Klotz, K. N.; Leung, E.; Baraldi, P. G.; Borea, P. A. Binding Thermodynamics at the Human A(3) Adenosine Receptor. *Biochem. Pharmacol.* **2002**, *63* (2), 157–161.
- (90) Gessi, S.; Fogli, E.; Sacchetto, V.; Varani, K.; Merighi, S.; Leung, E.; Lennan, S. Mac; Borea, P. A. Thermodynamics of A2B Adenosine Receptor Binding Discriminates Agonistic from Antagonistic Behaviour. *Biochem. Pharmacol.* **2008**, *75* (2), 562–569.
- (91) McKallip, R. J.; Lombard, C.; Fisher, M.; Martin, B. R.; Ryu, S.; Grant, S.; Nagarkatti, P. S.; Nagarkatti, M. Targeting CB2 Cannabinoid Receptors as a Novel Therapy to Treat Malignant Lymphoblastic Disease. *Blood* **2002**, *100* (2), 627–634.
- (92) Robinson, R. H.; Meissler, J. J.; Breslow-Deckman, J. M.; Gaughan, J.; Adler, M. W.; Eisenstein, T. K. Cannabinoids Inhibit T-Cells via Cannabinoid Receptor 2 in an in Vitro Assay for Graft Rejection, the Mixed Lymphocyte Reaction. *J. Neuroimmune Pharmacol.* **2013**, *8* (5), 1239–1250.
- (93) Rieder, S. A.; Chauhan, A.; Singh, U.; Nagarkatti, M.; Nagarkatti, P. Cannabinoid-Induced Apoptosis in Immune Cells as a Pathway to Immunosuppression. *Immunobiology* **2010**, *215* (8), 598–605.
- (94) Jean-Gilles, L.; Braitch, M.; Latif, M. L.; Aram, J.; Fahey, A. J.; Edwards, L. J.; Robins, R. A.; Tanasescu, R.; Tighe, P. J.; Gran, B.; et al. Effects of pro-Inflammatory Cytokines on Cannabinoid CB1 and CB2 Receptors in Immune Cells. *Acta Physiol. (Oxf)*. **2015**, *214* (1), 63–74.
- (95) Chen, Y.; Buck, J. Cannabinoids Protect Cells from Oxidative Cell Death: A Receptor-Independent Mechanism. *J. Pharmacol. Exp. Ther.* **2000**, *293* (3), 807–812.
- (96) Martínez-Orgado, J.; Fernández-López, D.; Lizasoain, I.; Romero, J. The Seek of Neuroprotection: Introducing Cannabinoids. *Recent Pat. CNS Drug Discov.* **2007**, *2* (2), 131–139.
- (97) Mecha, M.; Torrao, A. S.; Mestre, L.; Carrillo-Salinas, F. J.; Mechoulam, R.; Guaza, C. Cannabidiol Protects Oligodendrocyte Progenitor Cells from Inflammation-Induced Apoptosis by Attenuating Endoplasmic Reticulum Stress. *Cell Death Dis.* **2012**, *3*, e331.
- (98) Patel, K. D.; Davison, J. S.; Pittman, Q. J.; Sharkey, K. A. Cannabinoid CB(2) Receptors in Health and Disease. *Curr. Med. Chem.* **2010**, *17* (14), 1393–1410.
- (99) Pereira, J. P.; An, J.; Xu, Y.; Huang, Y.; Cyster, J. G. Cannabinoid Receptor 2 Mediates the Retention of Immature B Cells in Bone Marrow Sinusoids. *Nat. Immunol.* **2009**, *10* (4), 403–411.
- (100) Sophocleous, A.; Börjesson, A. E.; Salter, D. M.; Ralston, S. H. The Type 2 Cannabinoid Receptor Regulates Susceptibility to Osteoarthritis in Mice. *Osteoarthritis Cartilage* **2015**.

- (101) Gui, H.; Tong, Q.; Qu, W.; Mao, C.-M.; Dai, S.-M. The Endocannabinoid System and Its Therapeutic Implications in Rheumatoid Arthritis. *Int. Immunopharmacol.* **2015**, *26* (1), 86–91.
- (102) Richardson, D.; Pearson, R. G.; Kurian, N.; Latif, M. L.; Garle, M. J.; Barrett, D. A.; Kendall, D. A.; Scammell, B. E.; Reeve, A. J.; Chapman, V. Characterisation of the Cannabinoid Receptor System in Synovial Tissue and Fluid in Patients with Osteoarthritis and Rheumatoid Arthritis. *Arthritis Res. Ther.* **2008**, *10* (2), R43.
- (103) Montecucco, F.; Burger, F.; Mach, F.; Steffens, S. CB2 Cannabinoid Receptor Agonist JWH-015 Modulates Human Monocyte Migration through Defined Intracellular Signaling Pathways. *Am. J. Physiol. Heart Circ. Physiol.* **2008**, *294* (3), H1145–H1155.
- (104) Gaffal, E.; Cron, M.; Glodde, N.; Tüting, T. Anti-Inflammatory Activity of Topical THC in DNFB-Mediated Mouse Allergic Contact Dermatitis Independent of CB1 and CB2 Receptors. *Allergy* **2013**, *68* (8), 994–1000.
- (105) Han, S.; Thatte, J.; Buzard, D. J.; Jones, R. M. Therapeutic Utility of Cannabinoid Receptor Type 2 (CB₂) Selective Agonists. *J. Med. Chem.* **2013**, *56* (21), 8224–8256.
- (106) Xu, H.; Cheng, C. L.; Chen, M.; Manivannan, A.; Cabay, L.; Pertwee, R. G.; Coutts, A.; Forrester, J. V. Anti-Inflammatory Property of the Cannabinoid Receptor-2-Selective Agonist JWH-133 in a Rodent Model of Autoimmune Uveoretinitis. *J. Leukoc. Biol.* **2007**, *82* (3), 532–541.
- (107) Carlisle, S. J.; Marciano-Cabral, F.; Staab, A.; Ludwick, C.; Cabral, G. A. Differential Expression of the CB₂ Cannabinoid Receptor by Rodent Macrophages and Macrophage-like Cells in Relation to Cell Activation. *Int. Immunopharmacol.* **2002**, *2* (1), 69–82.
- (108) Eljaschewitsch, E.; Witting, A.; Mawrin, C.; Lee, T.; Schmidt, P. M.; Wolf, S.; Hoertnagl, H.; Raine, C. S.; Schneider-Stock, R.; Nitsch, R.; et al. The Endocannabinoid Anandamide Protects Neurons during CNS Inflammation by Induction of MKP-1 in Microglial Cells. *Neuron* **2006**, *49* (1), 67–79.
- (109) Benito, C.; Tolón, R. M.; Pazos, M. R.; Núñez, E.; Castillo, A. I.; Romero, J. Cannabinoid CB₂ Receptors in Human Brain Inflammation. *Br. J. Pharmacol.* **2008**, *153* (2), 277–285.
- (110) Benito, C.; Kim, W.-K.; Chavarría, I.; Hillard, C. J.; Mackie, K.; Tolón, R. M.; Williams, K.; Williams, K.; Romero, J. A Glial Endogenous Cannabinoid System Is Upregulated in the Brains of Macaques with Simian Immunodeficiency Virus-Induced Encephalitis. *J. Neurosci.* **2005**, *25* (10), 2530–2536.
- (111) Concannon, R. M.; Okine, B. N.; Finn, D. P.; Dowd, E. Differential Upregulation of the Cannabinoid CB₂ Receptor in Neurotoxic and Inflammation-Driven Rat Models of Parkinson's Disease. *Exp. Neurol.* **2015**, *269*, 133–141.
- (112) Gómez-Gálvez, Y.; Palomo-Garo, C.; Fernández-Ruiz, J.; García, C. Potential of the Cannabinoid CB₂ Receptor as a Pharmacological Target against Inflammation in Parkinson's Disease. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2015**.

- (113) Rajesh, M.; Mukhopadhyay, P.; Haskó, G.; Huffman, J. W.; Mackie, K.; Pacher, P. CB2 Cannabinoid Receptor Agonists Attenuate TNF-Alpha-Induced Human Vascular Smooth Muscle Cell Proliferation and Migration. *Br. J. Pharmacol.* **2008**, *153* (2), 347–357.
- (114) Kurihara, R.; Tohyama, Y.; Matsusaka, S.; Naruse, H.; Kinoshita, E.; Tsujioka, T.; Katsumata, Y.; Yamamura, H. Effects of Peripheral Cannabinoid Receptor Ligands on Motility and Polarization in Neutrophil-like HL60 Cells and Human Neutrophils. *J. Biol. Chem.* **2006**, *281* (18), 12908–12918.
- (115) Mecha, M.; Feliú, A.; Carrillo-Salinas, F. J.; Rueda-Zubiaurre, A.; Ortega-Gutiérrez, S.; de Sola, R. G.; Guaza, C. Endocannabinoids Drive the Acquisition of an Alternative Phenotype in Microglia. *Brain. Behav. Immun.* **2015**.
- (116) Adhikary, S.; Kocieda, V. P.; Yen, J.-H.; Tuma, R. F.; Ganea, D. Signaling through Cannabinoid Receptor 2 Suppresses Murine Dendritic Cell Migration by Inhibiting Matrix Metalloproteinase 9 Expression. *Blood* **2012**, *120* (18), 3741–3749.
- (117) Golech, S. A.; McCarron, R. M.; Chen, Y.; Bembry, J.; Lenz, F.; Mechoulam, R.; Shohami, E.; Spatz, M. Human Brain Endothelium: Coexpression and Function of Vanilloid and Endocannabinoid Receptors. *Brain Res. Mol. Brain Res.* **2004**, *132* (1), 87–92.
- (118) Sospedra, M.; Martin, R. Immunology of Multiple Sclerosis. *Annu. Rev. Immunol.* **2005**, *23*, 683–747.
- (119) Maresz, K.; Carrier, E. J.; Ponomarev, E. D.; Hillard, C. J.; Dittel, B. N. Modulation of the Cannabinoid CB2 Receptor in Microglial Cells in Response to Inflammatory Stimuli. *J. Neurochem.* **2005**, *95* (2), 437–445.
- (120) Ehrhart, J.; Obregon, D.; Mori, T.; Hou, H.; Sun, N.; Bai, Y.; Klein, T.; Fernandez, F.; Tan, J.; Shytle, R. D. Stimulation of Cannabinoid Receptor 2 (CB2) Suppresses Microglial Activation. *J. Neuroinflammation* **2005**, *2*, 29.
- (121) Maresz, K.; Pryce, G.; Ponomarev, E. D.; Marsicano, G.; Croxford, J. L.; Shriver, L. P.; Ledent, C.; Cheng, X.; Carrier, E. J.; Mann, M. K.; et al. Direct Suppression of CNS Autoimmune Inflammation via the Cannabinoid Receptor CB1 on Neurons and CB2 on Autoreactive T Cells. *Nat. Med.* **2007**, *13* (4), 492–497.
- (122) Palazuelos, J.; Davoust, N.; Julien, B.; Hatterer, E.; Aguado, T.; Mechoulam, R.; Benito, C.; Romero, J.; Silva, A.; Guzmán, M.; et al. The CB(2) Cannabinoid Receptor Controls Myeloid Progenitor Trafficking: Involvement in the Pathogenesis of an Animal Model of Multiple Sclerosis. *J. Biol. Chem.* **2008**, *283* (19), 13320–13329.
- (123) Zhang, M.; Martin, B. R.; Adler, M. W.; Razdan, R. J.; Kong, W.; Ganea, D.; Tuma, R. F. Modulation of Cannabinoid Receptor Activation as a Neuroprotective Strategy for EAE and Stroke. *J. Neuroimmune Pharmacol.* **2009**, *4* (2), 249–259.
- (124) Pava, M. J.; Woodward, J. J. A Review of the Interactions between Alcohol and the Endocannabinoid System: Implications for Alcohol Dependence and Future Directions for Research. *Alcohol* **2012**, *46* (3), 185–204.

- (125) Pradier, B.; Erxlebe, E.; Markert, A.; Rácz, I. Interaction of Cannabinoid Receptor 2 and Social Environment Modulates Chronic Alcohol Consumption. *Behav. Brain Res.* **2015**, *287*, 163–171.
- (126) Onaivi, E. S.; Ishiguro, H.; Gong, J.-P.; Patel, S.; Meozzi, P. A.; Myers, L.; Perchuk, A.; Mora, Z.; Tagliaferro, P. A.; Gardner, E.; et al. Functional Expression of Brain Neuronal CB2 Cannabinoid Receptors Are Involved in the Effects of Drugs of Abuse and in Depression. *Ann. N. Y. Acad. Sci.* **2008**, *1139*, 434–449.
- (127) Onaivi, E. S.; Carpio, O.; Ishiguro, H.; Schanz, N.; Uhl, G. R.; Benno, R. Behavioral Effects of CB2 Cannabinoid Receptor Activation and Its Influence on Food and Alcohol Consumption. *Ann. N. Y. Acad. Sci.* **2008**, *1139*, 426–433.
- (128) Ishiguro, H.; Iwasaki, S.; Teasenfitz, L.; Higuchi, S.; Horiuchi, Y.; Saito, T.; Arinami, T.; Onaivi, E. S. Involvement of Cannabinoid CB2 Receptor in Alcohol Preference in Mice and Alcoholism in Humans. *Pharmacogenomics J.* **2007**, *7* (6), 380–385.
- (129) Gong, J.-P.; Onaivi, E. S.; Ishiguro, H.; Liu, Q.-R.; Tagliaferro, P. A.; Brusco, A.; Uhl, G. R. Cannabinoid CB2 Receptors: Immunohistochemical Localization in Rat Brain. *Brain Res.* **2006**, *1071* (1), 10–23.
- (130) Brusco, A.; Tagliaferro, P.; Saez, T.; Onaivi, E. S. Postsynaptic Localization of CB2 Cannabinoid Receptors in the Rat Hippocampus. *Synapse* **2008**, *62* (12), 944–949.
- (131) Ishiguro, H.; Carpio, O.; Horiuchi, Y.; Shu, A.; Higuchi, S.; Schanz, N.; Benno, R.; Arinami, T.; Onaivi, E. S. A Nonsynonymous Polymorphism in Cannabinoid CB2 Receptor Gene Is Associated with Eating Disorders in Humans and Food Intake Is Modified in Mice by Its Ligands. *Synapse* **2010**, *64* (1), 92–96.
- (132) Pacher, P.; Mukhopadhyay, P.; Mohanraj, R.; Godlewski, G.; Bátkai, S.; Kunos, G. Modulation of the Endocannabinoid System in Cardiovascular Disease: Therapeutic Potential and Limitations. *Hypertension* **2008**, *52* (4), 601–607.
- (133) Pacher, P.; Haskó, G. Endocannabinoids and Cannabinoid Receptors in Ischaemia-Reperfusion Injury and Preconditioning. *Br. J. Pharmacol.* **2008**, *153* (2), 252–262.
- (134) Moris, D.; Georgopoulos, S.; Felekouras, E.; Patsouris, E.; Theocharis, S. The Effect of Endocannabinoid System in Ischemia-Reperfusion Injury: A Friend or a Foe? *Expert Opin. Ther. Targets* **2015**, 1–15.
- (135) Hillard, C. J. Role of Cannabinoids and Endocannabinoids in Cerebral Ischemia. *Curr. Pharm. Des.* **2008**, *14* (23), 2347–2361.
- (136) Rajesh, M.; Pan, H.; Mukhopadhyay, P.; Bátkai, S.; Osei-Hyiaman, D.; Haskó, G.; Liaudet, L.; Gao, B.; Pacher, P. Cannabinoid-2 Receptor Agonist HU-308 Protects against Hepatic Ischemia/reperfusion Injury by Attenuating Oxidative Stress, Inflammatory Response, and Apoptosis. *J. Leukoc. Biol.* **2007**, *82* (6), 1382–1389.

- (137) Montecucco, F.; Lenglet, S.; Braunersreuther, V.; Burger, F.; Pelli, G.; Bertolotto, M.; Mach, F.; Steffens, S. CB(2) Cannabinoid Receptor Activation Is Cardioprotective in a Mouse Model of Ischemia/reperfusion. *J. Mol. Cell. Cardiol.* **2009**, *46* (5), 612–620.
- (138) Duerr, G.; Heinemann, J.; Kley, J.; Roell, W.; Ottersbach, A.; Zimmer, A.; Lutz, B.; Welz, A.; Dewald, O. P80 * Cannabinoid Receptor CB2 Prevents Development of Heart Failure in a Murine Model of Pressure Overload. *Cardiovasc. Res.* **2014**, *103* (suppl 1), S13–S13.
- (139) Duerr, G. D.; Heinemann, J. C.; Suchan, G.; Kolobara, E.; Wenzel, D.; Geisen, C.; Matthey, M.; Pässe-Tietjen, K.; Mahmud, W.; Ghanem, A.; et al. The Endocannabinoid-CB2 Receptor Axis Protects the Ischemic Heart at the Early Stage of Cardiomyopathy. *Basic Res. Cardiol.* **2014**, *109* (4), 425.
- (140) Di Filippo, C.; Rossi, F.; Rossi, S.; D'Amico, M. Cannabinoid CB2 Receptor Activation Reduces Mouse Myocardial Ischemia-Reperfusion Injury: Involvement of Cytokine/chemokines and PMN. *J. Leukoc. Biol.* **2004**, *75* (3), 453–459.
- (141) Hansson, G. K. Inflammation, Atherosclerosis, and Coronary Artery Disease. *N. Engl. J. Med.* **2005**, *352* (16), 1685–1695.
- (142) Steffens, S.; Veillard, N. R.; Arnaud, C.; Pelli, G.; Burger, F.; Staub, C.; Karsak, M.; Zimmer, A.; Frossard, J.-L.; Mach, F. Low Dose Oral Cannabinoid Therapy Reduces Progression of Atherosclerosis in Mice. *Nature* **2005**, *434* (7034), 782–786.
- (143) Arai, S.; Shelton, J. M.; Chen, M.; Bradley, M. N.; Castrillo, A.; Bookout, A. L.; Mak, P. A.; Edwards, P. A.; Mangelsdorf, D. J.; Tontonoz, P.; et al. A Role for the Apoptosis Inhibitory Factor AIM/Spalpa/Ap16 in Atherosclerosis Development. *Cell Metab.* **2005**, *1* (3), 201–213.
- (144) Chiurchiù, V.; Lanuti, M.; Catanzaro, G.; Fezza, F.; Rapino, C.; Maccarrone, M. Detailed Characterization of the Endocannabinoid System in Human Macrophages and Foam Cells, and Anti-Inflammatory Role of Type-2 Cannabinoid Receptor. *Atherosclerosis* **2014**, *233* (1), 55–63.
- (145) Liu, J.; Thewke, D. P.; Su, Y. R.; Linton, M. F.; Fazio, S.; Sinensky, M. S. Reduced Macrophage Apoptosis Is Associated with Accelerated Atherosclerosis in Low-Density Lipoprotein Receptor-Null Mice. *Arterioscler. Thromb. Vasc. Biol.* **2005**, *25* (1), 174–179.
- (146) Carbone, F.; Mach, F.; Vuilleumier, N.; Montecucco, F. Cannabinoid Receptor Type 2 Activation in Atherosclerosis and Acute Cardiovascular Diseases. *Curr. Med. Chem.* **2014**, *21* (35), 4046–4058.
- (147) Malik, Z.; Baik, D.; Schey, R. The Role of Cannabinoids in Regulation of Nausea and Vomiting, and Visceral Pain. *Curr. Gastroenterol. Rep.* **2015**, *17* (2), 429.
- (148) Storr, M.; Gaffal, E.; Saur, D.; Schusdziarra, V.; Allescher, H. D. Effect of Cannabinoids on Neural Transmission in Rat Gastric Fundus. *Can. J. Physiol. Pharmacol.* **2002**, *80* (1), 67–76.

- (149) Siegmund, S. V.; Schwabe, R. F. Endocannabinoids and Liver Disease. II. Endocannabinoids in the Pathogenesis and Treatment of Liver Fibrosis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2008**, *294* (2), G357–G362.
- (150) Xu, X.; Liu, Y.; Huang, S.; Liu, G.; Xie, C.; Zhou, J.; Fan, W.; Li, Q.; Wang, Q.; Zhong, D.; et al. Overexpression of Cannabinoid Receptors CB1 and CB2 Correlates with Improved Prognosis of Patients with Hepatocellular Carcinoma. *Cancer Genet. Cytogenet.* **2006**, *171* (1), 31–38.
- (151) Baldassarre, M.; Giannone, F. A.; Napoli, L.; Tovoli, A.; Ricci, C. S.; Tufoni, M.; Caraceni, P. The Endocannabinoid System in Advanced Liver Cirrhosis: Pathophysiological Implication and Future Perspectives. *Liver Int.* **2013**, *33* (9), 1298–1308.
- (152) Mulè, F.; Amato, A.; Baldassano, S.; Serio, R. Involvement of CB1 and CB2 Receptors in the Modulation of Cholinergic Neurotransmission in Mouse Gastric Preparations. *Pharmacol. Res.* **2007**, *56* (3), 185–192.
- (153) Kurjak, M.; Hamel, A. M.; Allescher, H. D.; Schusdziarra, V.; Storr, M. Differential Stimulatory Effects of Cannabinoids on VIP Release and NO Synthase Activity in Synaptosomal Fractions from Rat Ileum. *Neuropeptides* **2008**, *42* (5-6), 623–632.
- (154) Rhee, S. H.; Pothoulakis, C.; Mayer, E. A. Principles and Clinical Implications of the Brain-Gut-Enteric Microbiota Axis. *Nat. Rev. Gastroenterol. Hepatol.* **2009**, *6* (5), 306–314.
- (155) Ihenetu, K.; Molleman, A.; Parsons, M. E.; Whelan, C. J. Inhibition of Interleukin-8 Release in the Human Colonic Epithelial Cell Line HT-29 by Cannabinoids. *Eur. J. Pharmacol.* **2003**, *458* (1-2), 207–215.
- (156) Mayer, E. A.; Collins, S. M. Evolving Pathophysiologic Models of Functional Gastrointestinal Disorders. *Gastroenterology* **2002**, *122* (7), 2032–2048.
- (157) Fichna, J.; Bawa, M.; Thakur, G. A.; Tichkule, R.; Makriyannis, A.; McCafferty, D.-M.; Sharkey, K. A.; Storr, M. Cannabinoids Alleviate Experimentally Induced Intestinal Inflammation by Acting at Central and Peripheral Receptors. *PLoS One* **2014**, *9* (10), e109115.
- (158) Kimball, E. S.; Schneider, C. R.; Wallace, N. H.; Hornby, P. J. Agonists of Cannabinoid Receptor 1 and 2 Inhibit Experimental Colitis Induced by Oil of Mustard and by Dextran Sulfate Sodium. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2006**, *291* (2), G364–G371.
- (159) Borrelli, F.; Romano, B.; Petrosino, S.; Pagano, E.; Capasso, R.; Coppola, D.; Battista, G.; Orlando, P.; Di Marzo, V.; Izzo, A. A. Palmitoylethanolamide, a Naturally Occurring Lipid, Is an Orally Effective Intestinal Anti-Inflammatory Agent. *Br. J. Pharmacol.* **2015**, *172* (1), 142–158.
- (160) Cui, Y.-Y.; D'Agostino, B.; Risse, P.-A.; Marrocco, G.; Naline, E.; Zhang, Y.; Chen, H.-Z.; Finance, O.; Rinaldi-Carmona, M.; Rossi, F.; et al. Cannabinoid CB(2) Receptor Activation Prevents Bronchoconstriction and Airway Oedema in a Model of Gastro-Oesophageal Reflux. *Eur. J. Pharmacol.* **2007**, *573* (1-3), 206–213.

- (161) Belvisi, M. G.; Patel, H. J.; Freund-Michel, V.; Hele, D. J.; Crispino, N.; Birrell, M. A. Inhibitory Activity of the Novel CB2 Receptor Agonist, GW833972A, on Guinea-Pig and Human Sensory Nerve Function in the Airways. *Br. J. Pharmacol.* **2008**, *155* (4), 547–557.
- (162) Gkoumassi, E.; Dekkers, B. G. J.; Dröge, M. J.; Elzinga, C. R. S.; Schmidt, M.; Meurs, H.; Zaagsma, J.; Nelemans, S. A. Virodhamine and CP55,940 Modulate cAMP Production and IL-8 Release in Human Bronchial Epithelial Cells. *Br. J. Pharmacol.* **2007**, *151* (7), 1041–1048.
- (163) Maher, S. A.; Dubuis, E. D.; Belvisi, M. G. G-Protein Coupled Receptors Regulating Cough. *Curr. Opin. Pharmacol.* **2011**, *11* (3), 248–253.
- (164) Barnes, P. J. The Problem of Cough and Development of Novel Antitussives. *Pulm. Pharmacol. Ther.* **2007**, *20* (4), 416–422.
- (165) Liu, M.; Su, M.; Wang, Y.; Wei, W.; Qin, L.; Liu, X.; Tian, M.; Qian, C. Effect of Melilotus Extract on Lung Injury by Upregulating the Expression of Cannabinoid CB2 Receptors in Septic Rats. *BMC Complement. Altern. Med.* **2014**, *14*, 94.
- (166) Pacher, P.; Mechoulam, R. Is Lipid Signaling through Cannabinoid 2 Receptors Part of a Protective System? *Prog. Lipid Res.* **2011**, *50* (2), 193–211.
- (167) Sun, Y.-X.; Xu, A.-H.; Yang, Y.; Zhang, J.-X.; Yu, A.-W. Activation of Cannabinoid Receptor 2 Enhances Osteogenic Differentiation of Bone Marrow Derived Mesenchymal Stem Cells. *Biomed Res. Int.* **2015**, *2015*, 874982.
- (168) Rossi, F.; Bellini, G.; Tortora, C.; Bernardo, M. E.; Luongo, L.; Conforti, A.; Starc, N.; Manzo, I.; Nobili, B.; Locatelli, F.; et al. CB2 and TRPV1 Receptors Oppositely Modulate in Vitro Human Osteoblast Activity. *Pharmacol. Res.* **2015**.
- (169) Idris, A. I.; van 't Hof, R. J.; Greig, I. R.; Ridge, S. A.; Baker, D.; Ross, R. A.; Ralston, S. H. Regulation of Bone Mass, Bone Loss and Osteoclast Activity by Cannabinoid Receptors. *Nat. Med.* **2005**, *11* (7), 774–779.
- (170) Zofková, I.; Matucha, P. New Insights into the Physiology of Bone Regulation: The Role of Neurohormones. *Physiol. Res.* **2014**, *63* (4), 421–427.
- (171) Smoum, R.; Baraghithy, S.; Chourasia, M.; Breuer, A.; Mussai, N.; Attar-Namdar, M.; Kogan, N. M.; Raphael, B.; Bolognini, D.; Cascio, M. G.; et al. CB2 Cannabinoid Receptor Agonist Enantiomers HU-433 and HU-308: An Inverse Relationship between Binding Affinity and Biological Potency. *Proc. Natl. Acad. Sci.* **2015**, 201503395.
- (172) Lunn, C. A.; Fine, J.; Rojas-Triana, A.; Jackson, J. V.; Lavey, B.; Kozlowski, J. A.; Hipkin, R. W.; Lundell, D. J.; Bober, L. Cannabinoid CB(2)-Selective Inverse Agonist Protects against Antigen-Induced Bone Loss. *Immunopharmacol. Immunotoxicol.* **2007**, *29* (3-4), 387–401.
- (173) Cridge, B. J.; Rosengren, R. J. Critical Appraisal of the Potential Use of Cannabinoids in Cancer Management. *Cancer Manag. Res.* **2013**, *5*, 301–313.

- (174) Guzmán, M. Cannabinoids: Potential Anticancer Agents. *Nat. Rev. Cancer* **2003**, *3* (10), 745–755.
- (175) Qamri, Z.; Preet, A.; Nasser, M. W.; Bass, C. E.; Leone, G.; Barsky, S. H.; Ganju, R. K. Synthetic Cannabinoid Receptor Agonists Inhibit Tumor Growth and Metastasis of Breast Cancer. *Mol. Cancer Ther.* **2009**, *8* (11), 3117–3129.
- (176) Sarfaraz, S.; Adhami, V. M.; Syed, D. N.; Afaq, F.; Mukhtar, H. Cannabinoids for Cancer Treatment: Progress and Promise. *Cancer Res.* **2008**, *68* (2), 339–342.
- (177) Hermanson, D. J.; Marnett, L. J. Cannabinoids, Endocannabinoids, and Cancer. *Cancer Metastasis Rev.* **2011**, *30* (3-4), 599–612.
- (178) Galve-Roperh, I.; Sánchez, C.; Cortés, M. L.; Gómez del Pulgar, T.; Izquierdo, M.; Guzmán, M. Anti-Tumoral Action of Cannabinoids: Involvement of Sustained Ceramide Accumulation and Extracellular Signal-Regulated Kinase Activation. *Nat. Med.* **2000**, *6* (3), 313–319.
- (179) Hall, W.; Christie, M.; Currow, D. Cannabinoids and Cancer: Causation, Remediation, and Palliation. *Lancet. Oncol.* **2005**, *6* (1), 35–42.
- (180) Ayakannu, T.; Taylor, A. H.; Marczylo, T. H.; Willets, J. M.; Konje, J. C. The Endocannabinoid System and Sex Steroid Hormone-Dependent Cancers. *Int. J. Endocrinol.* **2013**, *2013*, 259676.
- (181) Portella, G.; Laezza, C.; Laccetti, P.; De Petrocellis, L.; Di Marzo, V.; Bifulco, M. Inhibitory Effects of Cannabinoid CB1 Receptor Stimulation on Tumor Growth and Metastatic Spreading: Actions on Signals Involved in Angiogenesis and Metastasis. *FASEB J.* **2003**, *17* (12), 1771–1773.
- (182) Blázquez, C.; Casanova, M. L.; Planas, A.; Gómez Del Pulgar, T.; Villanueva, C.; Fernández-Aceñero, M. J.; Aragonés, J.; Huffman, J. W.; Jorcano, J. L.; Guzmán, M. Inhibition of Tumor Angiogenesis by Cannabinoids. *FASEB J.* **2003**, *17* (3), 529–531.
- (183) Casanova, M. L.; Blázquez, C.; Martínez-Palacio, J.; Villanueva, C.; Fernández-Aceñero, M. J.; Huffman, J. W.; Jorcano, J. L.; Guzmán, M. Inhibition of Skin Tumor Growth and Angiogenesis in Vivo by Activation of Cannabinoid Receptors. *J. Clin. Invest.* **2003**, *111* (1), 43–50.
- (184) Islam, T. C.; Asplund, A. C.; Lindvall, J. M.; Nygren, L.; Liden, J.; Kimby, E.; Christensson, B.; Smith, C. I. E.; Sander, B. High Level of Cannabinoid Receptor 1, Absence of Regulator of G Protein Signalling 13 and Differential Expression of Cyclin D1 in Mantle Cell Lymphoma. *Leukemia* **2003**, *17* (9), 1880–1890.
- (185) Caffarel, M. M.; Sarrió, D.; Palacios, J.; Guzmán, M.; Sánchez, C. Delta9-Tetrahydrocannabinol Inhibits Cell Cycle Progression in Human Breast Cancer Cells through Cdc2 Regulation. *Cancer Res.* **2006**, *66* (13), 6615–6621.

- (186) Alberich Jordà, M.; Rayman, N.; Tas, M.; Verbakel, S. E.; Battista, N.; van Lom, K.; Löwenberg, B.; Maccarrone, M.; Delwel, R. The Peripheral Cannabinoid Receptor Cb2, Frequently Expressed on AML Blasts, Either Induces a Neutrophilic Differentiation Block or Confers Abnormal Migration Properties in a Ligand-Dependent Manner. *Blood* **2004**, *104* (2), 526–534.
- (187) Sánchez, C.; de Ceballos, M. L.; Gomez del Pulgar, T.; Rueda, D.; Corbacho, C.; Velasco, G.; Galve-Roperh, I.; Huffman, J. W.; Ramón y Cajal, S.; Guzmán, M. Inhibition of Glioma Growth in Vivo by Selective Activation of the CB(2) Cannabinoid Receptor. *Cancer Res.* **2001**, *61* (15), 5784–5789.
- (188) Schley, M.; Ständer, S.; Kerner, J.; Vajkoczy, P.; Schüpfer, G.; Dusch, M.; Schmelz, M.; Konrad, C. Predominant CB2 Receptor Expression in Endothelial Cells of Glioblastoma in Humans. *Brain Res. Bull.* **2009**, *79* (5), 333–337.
- (189) Solinas, M.; Massi, P.; Cantelmo, A. R.; Cattaneo, M. G.; Cammarota, R.; Bartolini, D.; Cinquina, V.; Valenti, M.; Vicentini, L. M.; Noonan, D. M.; et al. Cannabidiol Inhibits Angiogenesis by Multiple Mechanisms. *Br. J. Pharmacol.* **2012**, *167* (6), 1218–1231.
- (190) Do, Y.; McKallip, R. J.; Nagarkatti, M.; Nagarkatti, P. S. Activation through Cannabinoid Receptors 1 and 2 on Dendritic Cells Triggers NF-kappaB-Dependent Apoptosis: Novel Role for Endogenous and Exogenous Cannabinoids in Immunoregulation. *J. Immunol.* **2004**, *173* (4), 2373–2382.
- (191) De Filippis, D.; Russo, A.; De Stefano, D.; Maiuri, M. C.; Esposito, G.; Cinelli, M. P.; Pietropaolo, C.; Carnuccio, R.; Russo, G.; Iuvone, T. Local Administration of WIN 55,212-2 Reduces Chronic Granuloma-Associated Angiogenesis in Rat by Inhibiting NF-kappaB Activation. *J. Mol. Med. (Berl)*. **2007**, *85* (6), 635–645.
- (192) Blázquez, C.; Carracedo, A.; Barrado, L.; Real, P. J.; Fernández-Luna, J. L.; Velasco, G.; Malumbres, M.; Guzmán, M. Cannabinoid Receptors as Novel Targets for the Treatment of Melanoma. *FASEB J.* **2006**, *20* (14), 2633–2635.
- (193) Bellacosa, A.; Kumar, C. C.; Di Cristofano, A.; Testa, J. R. Activation of AKT Kinases in Cancer: Implications for Therapeutic Targeting. *Adv. Cancer Res.* **2005**, *94*, 29–86.
- (194) Wang, D.; Wang, H.; Ning, W.; Backlund, M. G.; Dey, S. K.; DuBois, R. N. Loss of Cannabinoid Receptor 1 Accelerates Intestinal Tumor Growth. *Cancer Res.* **2008**, *68* (15), 6468–6476.
- (195) Romano, B.; Borrelli, F.; Pagano, E.; Cascio, M. G.; Pertwee, R. G.; Izzo, A. A. Inhibition of Colon Carcinogenesis by a Standardized Cannabis Sativa Extract with High Content of Cannabidiol. *Phytomedicine* **2014**, *21* (5), 631–639.
- (196) Izzo, A. The Cannabinoid CB(2) Receptor: A Good Friend in the Gut. *Neurogastroenterol. Motil.* **2007**, *19* (9), 704–708.
- (197) Pérez-Gómez, E.; Andradas, C.; Blasco-Benito, S.; Caffarel, M. M.; García-Taboada, E.; Villa-Morales, M.; Moreno, E.; Hamann, S.; Martín-Villar, E.; Flores, J. M.; et al. Role of

Cannabinoid Receptor CB2 in HER2 Pro-Oncogenic Signaling in Breast Cancer. *J. Natl. Cancer Inst.* **2015**, *107* (6), djv077.

- (198) Guindon, J.; Hohmann, A. G. The Endocannabinoid System and Cancer: Therapeutic Implication. *Br. J. Pharmacol.* **2011**, *163* (7), 1447–1463.
- (199) Orellana-Serradell, O.; Poblete, C. E.; Sanchez, C.; Castellón, E. A.; Gallegos, I.; Huidobro, C.; Llanos, M. N.; Contreras, H. R. Proapoptotic Effect of Endocannabinoids in Prostate Cancer Cells. *Oncol. Rep.* **2015**, *33* (4), 1599–1608.
- (200) Gilron, I.; Baron, R.; Jensen, T. Neuropathic Pain: Principles of Diagnosis and Treatment. *Mayo Clin. Proc.* **2015**, *90* (4), 532–545.
- (201) Tauben, D. Nonopioid Medications for Pain. *Phys. Med. Rehabil. Clin. N. Am.* **2015**, *26* (2), 219–248.
- (202) Woodhams, S. G.; Sagar, D. R.; Burston, J. J.; Chapman, V. The Role of the Endocannabinoid System in Pain. *Handb. Exp. Pharmacol.* **2015**, *227*, 119–143.
- (203) Devesa, I.; Ferrer-Montiel, A. Neurotrophins, Endocannabinoids and Thermo-Transient Receptor Potential: A Threesome in Pain Signalling. *Eur. J. Neurosci.* **2014**, *39* (3), 353–362.
- (204) Hill, K. P. Medical Marijuana for Treatment of Chronic Pain and Other Medical and Psychiatric Problems. *JAMA* **2015**, *313* (24), 2474–2483.
- (205) Whiting, P. F.; Wolff, R. F.; Deshpande, S.; Di Nisio, M.; Duffy, S.; Hernandez, A. V.; Keurentjes, J. C.; Lang, S.; Misso, K.; Ryder, S.; et al. Cannabinoids for Medical Use. *JAMA* **2015**, *313* (24), 2456–2473.
- (206) Gilron, I.; Dickenson, A. H. Emerging Drugs for Neuropathic Pain. *Expert Opin. Emerg. Drugs* **2014**, *19* (3), 329–341.
- (207) Lucchesi, V.; Parkkari, T.; Savinainen, J. R.; Malfitano, A. M.; Allarà, M.; Bertini, S.; Castelli, F.; Del Carlo, S.; Laezza, C.; Ligresti, A.; et al. 1,2-Dihydro-2-Oxopyridine-3-Carboxamides: The C-5 Substituent Is Responsible for Functionality Switch at CB2 Cannabinoid Receptor. *Eur. J. Med. Chem.* **2014**, *74*, 524–532.
- (208) Davis, M. P. Cannabinoids in Pain Management: CB1, CB2 and Non-Classic Receptor Ligands. *Expert Opin. Investig. Drugs* **2014**, *23* (8), 1123–1140.
- (209) Vincenzi, F.; Targa, M.; Corciulo, C.; Tabrizi, M. A.; Merighi, S.; Gessi, S.; Saponaro, G.; Baraldi, P. G.; Borea, P. A.; Varani, K. Antinociceptive Effects of the Selective CB2 Agonist MT178 in Inflammatory and Chronic Rodent Pain Models. *Pain* **2013**, *154* (6), 864–873.
- (210) Ikeda, H.; Ikegami, M.; Kai, M.; Ohsawa, M.; Kamei, J. Activation of Spinal Cannabinoid CB2 Receptors Inhibits Neuropathic Pain in Streptozotocin-Induced Diabetic Mice. *Neuroscience* **2013**, *250*, 446–454.

- (211) Grim, T. W.; Ghosh, S.; Hsu, K.-L.; Cravatt, B. F.; Kinsey, S. G.; Lichtman, A. H. Combined Inhibition of FAAH and COX Produces Enhanced Anti-Allodynic Effects in Mouse Neuropathic and Inflammatory Pain Models. *Pharmacol. Biochem. Behav.* **2014**, *124*, 405–411.
- (212) Deng, L.; Guindon, J.; Cornett, B. L.; Makriyannis, A.; Mackie, K.; Hohmann, A. G. Chronic Cannabinoid Receptor 2 Activation Reverses Paclitaxel Neuropathy without Tolerance or Cannabinoid Receptor 1-Dependent Withdrawal. *Biol. Psychiatry* **2015**, *77* (5), 475–487.
- (213) Chen, X.; Zheng, C.; Qian, J.; Sutton, S. W.; Wang, Z.; Lv, J.; Liu, C.; Zhou, N. Involvement of B-Arrestin-2 and Clathrin in Agonist-Mediated Internalization of the Human Cannabinoid CB2 Receptor. *Curr. Mol. Pharmacol.* **2014**, *7* (1), 67–80.
- (214) Petrov, R. R.; Knight, L.; Chen, S.-R.; Wager-Miller, J.; McDaniel, S. W.; Diaz, F.; Barth, F.; Pan, H.-L.; Mackie, K.; Cavaotto, C. N.; et al. Mastering Tricyclic Ring Systems for Desirable Functional Cannabinoid Activity. *Eur. J. Med. Chem.* **2013**, *69*, 881–907.
- (215) Jean-Gilles, L.; Braitch, M.; Latif, M. L.; Aram, J.; Fahey, A. J.; Edwards, L. J.; Robins, R. A.; Tanasescu, R.; Tighe, P. J.; Gran, B.; et al. Effects of pro-Inflammatory Cytokines on Cannabinoid CB1 and CB2 Receptors in Immune Cells. *Acta Physiol. (Oxf)*. **2015**, *214* (1), 63–74.
- (216) Gui, H.; Tong, Q.; Qu, W.; Mao, C.-M.; Dai, S.-M. The Endocannabinoid System and Its Therapeutic Implications in Rheumatoid Arthritis. *Int. Immunopharmacol.* **2015**, *26* (1), 86–91.
- (217) Han, S.; Thatte, J.; Buzard, D. J.; Jones, R. M. Therapeutic Utility of Cannabinoid Receptor Type 2 (CB(2)) Selective Agonists. *J. Med. Chem.* **2013**, *56* (21), 8224–8256.
- (218) Gilron, I.; Baron, R.; Jensen, T. Neuropathic Pain: Principles of Diagnosis and Treatment. *Mayo Clin. Proc.* **2015**, *90* (4), 532–545.
- (219) Tauben, D. Nonopioid Medications for Pain. *Phys. Med. Rehabil. Clin. N. Am.* **2015**, *26* (2), 219–248.
- (220) Moris, D.; Georgopoulos, S.; Felekouras, E.; Patsouris, E.; Theocharis, S. The Effect of Endocannabinoid System in Ischemia-Reperfusion Injury: A Friend or a Foe? *Expert Opin. Ther. Targets* **2015**, 1–15.
- (221) Malik, Z.; Baik, D.; Schey, R. The Role of Cannabinoids in Regulation of Nausea and Vomiting, and Visceral Pain. *Curr. Gastroenterol. Rep.* **2015**, *17* (2), 429.
- (222) Sun, Y.-X.; Xu, A.-H.; Yang, Y.; Zhang, J.-X.; Yu, A.-W. Activation of Cannabinoid Receptor 2 Enhances Osteogenic Differentiation of Bone Marrow Derived Mesenchymal Stem Cells. *Biomed Res. Int.* **2015**, *2015*, 874982.
- (223) Rossi, F.; Bellini, G.; Tortora, C.; Bernardo, M. E.; Luongo, L.; Conforti, A.; Starc, N.; Manzo, I.; Nobili, B.; Locatelli, F.; et al. CB2 and TRPV1 Receptors Oppositely Modulate in Vitro Human Osteoblast Activity. *Pharmacol. Res.* **2015**.

- (224) Zofková, I.; Matucha, P. New Insights into the Physiology of Bone Regulation: The Role of Neurohormones. *Physiol. Res.* **2014**, *63* (4), 421–427.
- (225) Huffman, J. W.; Yu, S.; Showalter, V.; Abood, M. E.; Wiley, J. L.; Compton, D. R.; Martin, B. R.; Bramblett, R. D.; Reggio, P. H. Synthesis and Pharmacology of a Very Potent Cannabinoid Lacking a Phenolic Hydroxyl with High Affinity for the CB2 Receptor. *J. Med. Chem.* **1996**, *39* (20), 3875–3877.
- (226) Elmes, S. J. R.; Jhaveri, M. D.; Smart, D.; Kendall, D. A.; Chapman, V. Cannabinoid CB2 Receptor Activation Inhibits Mechanically Evoked Responses of Wide Dynamic Range Dorsal Horn Neurons in Naïve Rats and in Rat Models of Inflammatory and Neuropathic Pain. *Eur. J. Neurosci.* **2004**, *20* (9), 2311–2320.
- (227) Elmes, S. J. R.; Winyard, L. A.; Medhurst, S. J.; Clayton, N. M.; Wilson, A. W.; Kendall, D. A.; Chapman, V. Activation of CB1 and CB2 Receptors Attenuates the Induction and Maintenance of Inflammatory Pain in the Rat. *Pain* **2005**, *118* (3), 327–335.
- (228) Michler, T.; Storr, M.; Kramer, J.; Ochs, S.; Malo, A.; Reu, S.; Göke, B.; Schäfer, C. Activation of Cannabinoid Receptor 2 Reduces Inflammation in Acute Experimental Pancreatitis via Intra-Acinar Activation of p38 and MK2-Dependent Mechanisms. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2013**, *304* (2), G181–G192.
- (229) Ramirez, S. H.; Haskó, J.; Skuba, A.; Fan, S.; Dykstra, H.; McCormick, R.; Reichenbach, N.; Krizbai, I.; Mahadevan, A.; Zhang, M.; et al. Activation of Cannabinoid Receptor 2 Attenuates Leukocyte-Endothelial Cell Interactions and Blood-Brain Barrier Dysfunction under Inflammatory Conditions. *J. Neurosci.* **2012**, *32* (12), 4004–4016.
- (230) Li, Q.; Wang, F.; Zhang, Y.-M.; Zhou, J.-J.; Zhang, Y. Activation of Cannabinoid Type 2 Receptor by JWH133 Protects Heart against Ischemia/reperfusion-Induced Apoptosis. *Cell. Physiol. Biochem.* **2013**, *31* (4-5), 693–702.
- (231) Hanus, A.; Tchilibon, S.; Shiloah, S.; Goldenberg, D.; Horowitz, M.; Pertwee, R. G.; Ross, R. A.; Mechoulam, R.; Frider, E. L.; Breuer, HU-308: A Specific Agonist for CB2, a Peripheral Cannabinoid Receptor. *Proc. Natl. Acad. Sci. U. S. A.* **1999**, *96* (25), 14228–14233.
- (232) LaBuda, C. J.; Kobilsh, M.; Little, P. J. Cannabinoid CB2 Receptor Agonist Activity in the Hindpaw Incision Model of Postoperative Pain. *Eur. J. Pharmacol.* **2005**, *527* (1-3), 172–174.
- (233) Malan, T. P.; Ibrahim, M. M.; Deng, H.; Liu, Q.; Mata, H. P.; Vanderah, T.; Porreca, F.; Makriyannis, A. CB2 Cannabinoid Receptor-Mediated Peripheral Antinociception. *Pain* **2001**, *93* (3), 239–245.
- (234) Ibrahim, M. M.; Deng, H.; Zvonok, A.; Cockayne, D. A.; Kwan, J.; Mata, H. P.; Vanderah, T. W.; Lai, J.; Porreca, F.; Makriyannis, A.; et al. Activation of CB2 Cannabinoid Receptors by AM1241 Inhibits Experimental Neuropathic Pain: Pain Inhibition by Receptors Not Present in the CNS. *Proc. Natl. Acad. Sci. U. S. A.* **2003**, *100* (18), 10529–10533.

- (235) Wilkerson, J. L.; Gentry, K. R.; Dengler, E. C.; Wallace, J. A.; Kerwin, A. A.; Kuhn, M. N.; Zvonok, A. M.; Thakur, G. A.; Makriyannis, A.; Milligan, E. D. Immunofluorescent Spectral Analysis Reveals the Intrathecal Cannabinoid Agonist, AM1241, Produces Spinal Anti-Inflammatory Cytokine Responses in Neuropathic Rats Exhibiting Relief from Allodynia. *Brain Behav.* **2012**, *2* (2), 155–177.
- (236) Giblin, G. M.; O'Shaughnessy, C. T.; Naylor, A.; Mitchell, W. L.; Eatherton, A. J.; Slingsby, B. P.; Rawlings, D. A.; Goldsmith, P.; Brown, A. J.; Haslam, C. P.; et al. Discovery of 2-[(2,4-Dichlorophenyl)amino]-N-[(tetrahydro-2H-Pyran-4-Yl)methyl]-4-(trifluoromethyl)-5-Pyrimidinecarboxamide, a Selective CB2 Receptor Agonist for the Treatment of Inflammatory Pain. *J. Med. Chem.* **2007**, *50* (11), 2597–2600.
- (237) Mitchell, W. L.; Giblin, G. M.; Naylor, A.; Eatherton, A. J.; Slingsby, B. P.; Rawlings, A. D.; Jandu, K. S.; Haslam, C. P.; Brown, A. J.; Goldsmith, P.; et al. Pyridine-3-Carboxamides as Novel CB(2) Agonists for Analgesia. *Bioorg. Med. Chem. Lett.* **2009**, *19* (1), 259–263.
- (238) Zindell, R.; Riether, D.; Bosanac, T.; Berry, A.; Gemkow, M. J.; Ebneith, A.; Lobbe, S.; Raymond, E. L.; Thome, D.; Shih, D. T.; et al. Morpholine Containing CB2 Selective Agonists. *Bioorg. Med. Chem. Lett.* **2009**, *19* (6), 1604–1609.
- (239) Gleave, R. J.; Beswick, P. J.; Brown, A. J.; Giblin, G. M. P.; Haslam, C. P.; Livermore, D.; Moses, A.; Nicholson, N. H.; Page, L. W.; Slingsby, B.; et al. 2-Amino-5-Aryl-Pyridines as Selective CB2 Agonists: Synthesis and Investigation of Structure-Activity Relationships. *Bioorg. Med. Chem. Lett.* **2009**, *19* (23), 6578–6581.
- (240) Gleave, R. J.; Beswick, P. J.; Brown, A. J.; Giblin, G. M.; Goldsmith, P.; Haslam, C. P.; Mitchell, W. L.; Nicholson, N. H.; Page, L. W.; Patel, S.; et al. Synthesis and Evaluation of 3-Amino-6-Aryl-Pyridazines as Selective CB(2) Agonists for the Treatment of Inflammatory Pain. *Bioorg Med Chem Lett* **2010**, *20* (2), 465–468.
- (241) Odan, M.; Ishizuka, N.; Hiramatsu, Y.; Inagaki, M.; Hashizume, H.; Fujii, Y.; Mitsumori, S.; Morioka, Y.; Soga, M.; Deguchi, M.; et al. Discovery of S-777469: An Orally Available CB2 Agonist as an Antipruritic Agent. *Bioorg. Med. Chem. Lett.* **2012**, *22* (8), 2803–2806.
- (242) Van der Stelt, M.; Cals, J.; Broeders-Josten, S.; Cottney, J.; van der Doelen, A. A.; Hermkens, M.; de Kimpe, V.; King, A.; Klomp, J.; Oosterom, J.; et al. Discovery and Optimization of 1-(4-(pyridin-2-Yl)benzyl)imidazolidine-2,4-Dione Derivatives as a Novel Class of Selective Cannabinoid CB2 Receptor Agonists. *J Med Chem* **2011**, *54* (20), 7350–7362.
- (243) Chu, G. H.; Saeui, C. T.; Worm, K.; Weaver, D. G.; Goodman, A. J.; Broadrup, R. L.; Cassel, J. A.; DeHaven, R. N.; LaBuda, C. J.; Koblisch, M.; et al. Novel Pyridine Derivatives as Potent and Selective CB2 Cannabinoid Receptor Agonists. *Bioorg Med Chem Lett* **2009**, *19* (20), 5931–5935.
- (244) Gavelle, O.; Grether, U.; Kimbara, A.; Nettekoven, M.; Roever, S.; Rogers-Evans, M.; Rombrach, D.; Schulz-Gasch, T. Novel Pyridine Derivatives. WO2014154612, 2014.

- (245) Worm, K.; Zhou, Q. J.; Saeui, C. T.; Green, R. C.; Cassel, J. A.; Stabley, G. J.; DeHaven, R. N.; Conway-James, N.; LaBuda, C. J.; Koblisch, M.; et al. Sulfamoyl Benzamides as Novel CB2 Cannabinoid Receptor Ligands. *Bioorg. Med. Chem. Lett.* **2008**, *18* (9), 2830–2835.
- (246) Goodman, A. J.; Ajello, C. W.; Worm, K.; Le Bourdonnec, B.; Savolainen, M. A.; O'Hare, H.; Cassel, J. A.; Stabley, G. J.; DeHaven, R. N.; LaBuda, C. J.; et al. CB(2) Selective Sulfamoyl Benzamides: Optimization of the Amide Functionality. *Bioorg. Med. Chem. Lett.* **2009**, *19* (2), 309–313.
- (247) Sellitto, I.; Le Bourdonnec, B.; Worm, K.; Goodman, A.; Savolainen, M. A.; Chu, G. H.; Ajello, C. W.; Saeui, C. T.; Leister, L. K.; Cassel, J. A.; et al. Novel Sulfamoyl Benzamides as Selective CB2 Agonists with Improved in Vitro Metabolic Stability. *Bioorg. Med. Chem. Lett.* **2010**, *20* (1), 387–391.
- (248) Ohta, H.; Ishizaka, T.; Tatsuzuki, M.; Yoshinaga, M.; Iida, I.; Tomishima, Y.; Toda, Y.; Saito, S. N-Alkylidenearylcaboxamides as New Potent and Selective CB2 Cannabinoid Receptor Agonists with Good Oral Bioavailability. *Bioorg. Med. Chem. Lett.* **2007**, *17* (22), 6299–6304.
- (249) Ohta, H.; Ishizaka, T.; Tatsuzuki, M.; Yoshinaga, M.; Iida, I.; Yamaguchi, T.; Tomishima, Y.; Futaki, N.; Todac, Y.; Saito, S. Imine Derivatives as New Potent and Selective CB2 Cannabinoid Receptor Agonists with an Analgesic Action. *Bioorg. Med. Chem.* **2008**, *16* (3), 1111–1124.
- (250) Cheng, Y.; Albrecht, B. K.; Brown, J.; Buchanan, J. L.; Buckner, W. H.; DiMauro, E. F.; Emkey, R.; Fremerey, R. T.; Harrange, J. C.; Hoffman, B. J.; et al. Discovery and Optimization of a Novel Series of N-Arylamide Oxadiazoles as Potent, Highly Selective and Orally Bioavailable Cannabinoid Receptor 2 (CB(2)) Agonists. *J. Med. Chem.* **2008**, *51* (16), 5019–5034.
- (251) DiMauro, E. F.; Buchanan, J. L.; Cheng, A.; Emkey, R.; Hitchcock, S. A.; Huang, L.; Huang, M. Y.; Janosky, B.; Lee, J. H.; Li, X.; et al. Structural Modifications of N-Arylamide Oxadiazoles: Identification of N-Arylpiperidine Oxadiazoles as Potent and Selective Agonists of CB(2). *Bioorg. Med. Chem. Lett.* **2008**, *18* (15), 4267–4274.
- (252) Marx, I. E.; DiMauro, E. F.; Cheng, A.; Emkey, R.; Hitchcock, S. A.; Huang, L. Y.; Huang, M. Y.; Human, J.; Lee, J. H.; Li, X. W.; et al. Discovery of Alpha-Amidosulfones as Potent and Selective Agonists of CB(2): Synthesis, SAR, and Pharmacokinetic Properties. *Bioorg. Med. Chem. Lett.* **2009**, *19* (1), 31–35.
- (253) Yang, S.-W.; Smotryski, J.; Matasi, J.; Ho, G.; Tulshian, D.; Greenlee, W. J.; Brusa, R.; Beltramo, M.; Cox, K. Structure-Activity Relationships of 2,4-Diphenyl-1H-Imidazole Analogs as CB2 Receptor Agonists for the Treatment of Chronic Pain. *Bioorg. Med. Chem. Lett.* **2011**, *21* (1), 182–185.
- (254) Page, D.; Balaux, E.; Boisvert, L.; Liu, Z.; Milburn, C.; Tremblay, M.; Wei, Z.; Woo, S.; Luo, X.; Cheng, Y. X.; et al. Novel Benzimidazole Derivatives as Selective CB2 Agonists. *Bioorg. Med. Chem. Lett.* **2008**, *18* (13), 3695–3700.

- (255) Ryckmans, T.; Edwards, M. P.; Horne, V. A.; Correia, A. M.; Owen, D. R.; Thompson, L. R.; Tran, I.; Tutt, M. F.; Young, T. Rapid Assessment of a Novel Series of Selective CB₂ Agonists Using Parallel Synthesis Protocols: A Lipophilic Efficiency (LipE) Analysis. *Bioorg. Med. Chem. Lett.* **2009**, *19* (15), 4406–4409.
- (256) Ando, K.; Iwata, Y. Preparation of Benzoimidazolylsulfone Derivatives for Use as CB₂ Receptor Agonists. WO 2010084767, 2010.
- (257) Iwata, Y.; Ando, K.; Taniguchi, K.; Koba, N.; Sugiura, A.; Sudo, M. Identification of a Highly Potent and Selective CB₂ Agonist, RQ-00202730, for the Treatment of Irritable Bowel Syndrome. *Bioorg. Med. Chem. Lett.* **2015**, *25* (2), 236–240.
- (258) Watson, C.; Owen, D. R.; Harding, D.; Kon, I. K.; Lewis, M. L.; Mason, H. J.; Matsumizu, M.; Mukaiyama, T.; Rodriguez-Lens, M.; Shima, A.; et al. Optimisation of a Novel Series of Selective CNS Penetrant CB₂ Agonists. *Bioorg. Med. Chem. Lett.* **2011**, *21* (14), 4284–4287.
- (259) Gijssen, H. J.; De Cleyn, M. A.; Surkyn, M.; Van Lommen, G. R.; Verbist, B. M.; Nijsen, M. J.; Meert, T.; Wauwe, J. V.; Aerssens, J. 5-Sulfonyl-Benzimidazoles as Selective CB₂ Agonists-Part 2. *Bioorg Med Chem Lett* **2012**, *22* (1), 547–552.
- (260) Verbist, B. M.; De Cleyn, M. A.; Surkyn, M.; Fraiponts, E.; Aerssens, J.; Nijsen, M. J.; Gijssen, H. J. 5-Sulfonyl-Benzimidazoles as Selective CB₂ Agonists. *Bioorg. Med. Chem. Lett.* **2008**, *18* (8), 2574–2579.
- (261) Yu, X. H.; Cao, C. Q.; Martino, G.; Puma, C.; Morinville, A.; St-Onge, S.; Lessard, E.; Perkins, M. N.; Laird, J. M. A Peripherally Restricted Cannabinoid Receptor Agonist Produces Robust Anti-Nociceptive Effects in Rodent Models of Inflammatory and Neuropathic Pain. *Pain* **2010**, *151* (2), 337–344.
- (262) Bilodeau, M. T.; Burgey, C. S.; Deng, Z. J.; Hartnett, J. C.; Kett, N. R.; Melamed, J.; Munson, P. M.; Nanda, K. K.; Thompson, W.; Trotter, B. W.; et al. Imidazopyridine Analogs as cb₂ Receptor Modulators, Useful in the Treatment of Pain, Respiratory and Non-Respiratory Diseases. WO 2008085302, 2008.
- (263) Nanda, K. K.; Henze, D. A.; Della Penna, K.; Desai, R.; Leitl, M.; Lemaire, W.; White, R. B.; Yeh, S.; Brouillette, J. N.; Hartman, G. D.; et al. Benzimidazole CB₂ Agonists: Design, Synthesis and SAR. *Bioorg. Med. Chem. Lett.* **2014**, *24* (4), 1218–1221.
- (264) Trotter, B. W.; Nanda, K. K.; Burgey, C. S.; Potteiger, C. M.; Deng, J. Z.; Green, A. I.; Hartnett, J. C.; Kett, N. R.; Wu, Z.; Henze, D. A.; et al. Imidazopyridine CB₂ Agonists: Optimization of CB₂/CB₁ Selectivity and Implications for in Vivo Analgesic Efficacy. *Bioorg Med Chem Lett* **2011**, *21* (8), 2354–2358.
- (265) Wu, Z.; Green, A. I.; Hartnett, J. C. CB₂ Receptor Ligands for the Treatment of Pain. WO 2009025785 **2009**.
- (266) Trotter, B. W.; Nanda, K. K.; Burgey, C. S.; Potteiger, C. M.; Deng, J. Z.; Green, A. I.; Hartnett, J. C.; Kett, N. R.; Wu, Z.; Henze, D. A.; et al. Imidazopyridine CB₂ Agonists:

Optimization of CB2/CB1 Selectivity and Implications for in Vivo Analgesic Efficacy. *Bioorg Med Chem Lett* **2011**, *21* (8), 2354–2358.

- (267) Hollinshead, S. P.; Astles, P. C.; Chambers, M. G.; Johnson, M. P.; Palmer, J.; Tidwell, M. W. Discovery and Optimization of Novel Purines as Potent and Selective CB2 Agonists. *Bioorg. Med. Chem. Lett.* **2012**, *22* (15), 4962–4966.
- (268) Hollinshead, S. P.; Tidwell, M. W.; Palmer, J.; Guidetti, R.; Sanderson, A.; Johnson, M. P.; Chambers, M. G.; Oskins, J.; Stratford, R.; Astles, P. C. Selective Cannabinoid Receptor Type 2 (CB2) Agonists: Optimization of a Series of Purines Leading to the Identification of a Clinical Candidate for the Treatment of Osteoarthritic Pain. *J. Med. Chem.* **2013**, *56* (14), 5722–5733.
- (269) Manera, C.; Cascio, M. G.; Benetti, V.; Allara, M.; Tuccinardi, T.; Martinelli, A.; Saccomanni, G.; Vivoli, E.; Ghelardini, C.; Di Marzo, V.; et al. New 1,8-Naphthyridine and Quinoline Derivatives as CB2 Selective Agonists. *Bioorg. Med. Chem. Lett.* **2007**, *17* (23), 6505–6510.
- (270) Stern, E.; Muccioli, G. G.; Millet, R.; Goossens, J.-F.; Farce, A.; Chavatte, P.; Poupaert, J. H.; Lambert, D. M.; Depreux, P.; Hénichart, J.-P. Novel 4-Oxo-1,4-Dihydroquinoline-3-Carboxamide Derivatives as New CB2 Cannabinoid Receptors Agonists: Synthesis, Pharmacological Properties and Molecular Modeling. *J. Med. Chem.* **2006**, *49* (1), 70–79.
- (271) Manera, C.; Saccomanni, G.; Adinolfi, B.; Benetti, V.; Ligresti, A.; Cascio, M. G.; Tuccinardi, T.; Lucchesi, V.; Martinelli, A.; Nieri, P.; et al. Rational Design, Synthesis, and Pharmacological Properties of New 1,8-Naphthyridin-2(1H)-on-3-Carboxamide Derivatives as Highly Selective Cannabinoid-2 Receptor Agonists. *J. Med. Chem.* **2009**, *52* (12), 3644–3651.
- (272) Lucchesi, V.; Hurst, D. P.; Shore, D. M.; Bertini, S.; Ehrmann, B. M.; Allarà, M.; Lawrence, L.; Ligresti, A.; Minutolo, F.; Saccomanni, G.; et al. CB2-Selective Cannabinoid Receptor Ligands: Synthesis, Pharmacological Evaluation, and Molecular Modeling Investigation of 1,8-Naphthyridin-2(1H)-One-3-Carboxamides. *J. Med. Chem.* **2014**, *57* (21), 8777–8791.
- (273) Cianchi, F.; Papucci, L.; Schiavone, N.; Lulli, M.; Magnelli, L.; Vinci, M. C.; Messerini, L.; Manera, C.; Ronconi, E.; Romagnani, P.; et al. Cannabinoid Receptor Activation Induces Apoptosis through Tumor Necrosis Factor Alpha-Mediated Ceramide de Novo Synthesis in Colon Cancer Cells. *Clin. Cancer Res.* **2008**, *14* (23), 7691–7700.
- (274) Pasquini, S.; Botta, L.; Semeraro, T.; Mugnaini, C.; Ligresti, A.; Palazzo, E.; Maione, S.; Di Marzo, V.; Corelli, F. Investigations on the 4-Quinolone-3-Carboxylic Acid Motif. 2. Synthesis and Structure-Activity Relationship of Potent and Selective Cannabinoid-2 Receptor Agonists Endowed with Analgesic Activity in Vivo. *J. Med. Chem.* **2008**, *51* (16), 5075–5084.
- (275) Pasquini, S.; Ligresti, A.; Mugnaini, C.; Semeraro, T.; Cicione, L.; De Rosa, M.; Guida, F.; Luongo, L.; De Chiaro, M.; Cascio, M. G.; et al. Investigations on the 4-Quinolone-3-Carboxylic Acid Motif. 3. Synthesis, Structure-Affinity Relationships, and Pharmacological

Characterization of 6-Substituted 4-Quinolone-3-Carboxamides as Highly Selective Cannabinoid-2 Receptor Ligands. *J. Med. Chem.* **2010**, *53* (16), 5915–5928.

- (276) Pasquini, S.; De Rosa, M.; Pedani, V.; Mugnaini, C.; Guida, F.; Luongo, L.; De Chiaro, M.; Maione, S.; Dragoni, S.; Frosini, M.; et al. Investigations on the 4-Quinolone-3-Carboxylic Acid Motif. 4. Identification of New Potent and Selective Ligands for the Cannabinoid Type 2 Receptor with Diverse Substitution Patterns and Antihyperalgesic Effects in Mice. *J. Med. Chem.* **2011**, *54* (15), 5444–5453.
- (277) El Bakali, J.; Muccioli, G. G.; Renault, N.; Pradal, D.; Body-Malapel, M.; Djouina, M.; Hamtiaux, L.; Andrzejak, V.; Desreumaux, P.; Chavatte, P.; et al. 4-Oxo-1,4-Dihydropyridines as Selective CB₂ Cannabinoid Receptor Ligands: Structural Insights into the Design of a Novel Inverse Agonist Series. *J. Med. Chem.* **2010**, *53* (22), 7918–7931.
- (278) El Bakali, J.; Gilleron, P.; Body-Malapel, M.; Mansouri, R.; Muccioli, G. G.; Djouina, M.; Barczyk, A.; Klupsch, F.; Andrzejak, V.; Lipka, E.; et al. 4-Oxo-1,4-Dihydropyridines as Selective CB₂ Cannabinoid Receptor Ligands. Part 2: Discovery of New Agonists Endowed with Protective Effect against Experimental Colitis. *J. Med. Chem.* **2012**, *55* (20), 8948–8952.
- (279) El Bakali, J.; Muccioli, G. G.; Body-Malapel, M.; Djouina, M.; Klupsch, F.; Ghinet, A.; Barczyk, A.; Renault, N.; Chavatte, P.; Desreumaux, P.; et al. Conformational Restriction Leading to a Selective CB₂ Cannabinoid Receptor Agonist Orally Active Against Colitis. *ACS Med. Chem. Lett.* **2015**, *6* (2), 198–203.
- (280) Manley, P. J.; Zartman, A.; Paone, D. V.; Burgey, C. S.; Henze, D. A.; Della Penna, K.; Desai, R.; Leitl, M. D.; Lemaire, W.; White, R. B.; et al. Decahydroquinoline Amides as Highly Selective CB₂ Agonists: Role of Selectivity on in Vivo Efficacy in a Rodent Model of Analgesia. *Bioorg. Med. Chem. Lett.* **2011**, *21* (8), 2359–2364.
- (281) Cheng, Y.-X.; Pourashraf, M.; Luo, X.; Srivastava, S.; Walpole, C.; Salois, D.; St-Onge, S.; Payza, K.; Lessard, E.; Yu, X. H.; et al. Γ -Carbolines: A Novel Class of Cannabinoid Agonists with High Aqueous Solubility and Restricted CNS Penetration. *Bioorg. Med. Chem. Lett.* **2012**, *22* (4), 1619–1624.
- (282) Huffman, J. W. The Search for Selective Ligands for the CB₂ Receptor. *Curr. Pharm. Des.* **2000**, *6* (13), 1323–1337.
- (283) Huffman, J. W.; Zengin, G.; Wu, M.-J.; Lu, J.; Hynd, G.; Bushell, K.; Thompson, A. L. S.; Bushell, S.; Tartal, C.; Hurst, D. P.; et al. Structure-Activity Relationships for 1-Alkyl-3-(1-Naphthoyl)indoles at the Cannabinoid CB(1) and CB(2) Receptors: Steric and Electronic Effects of Naphthoyl Substituents. New Highly Selective CB(2) Receptor Agonists. *Bioorg. Med. Chem.* **2005**, *13* (1), 89–112.
- (284) Frost, J. M.; Dart, M. J.; Tietje, K. R.; Garrison, T. R.; Grayson, G. K.; Daza, A. V.; El-Kouhen, O. F.; Yao, B. B.; Hsieh, G. C.; Pai, M.; et al. Indol-3-Ylcycloalkyl Ketones: Effects of N1 Substituted Indole Side Chain Variations on CB₂ Cannabinoid Receptor Activity. *J. Med. Chem.* **2010**, *53* (1), 295–315.

- (285) Yao, B. B.; Hsieh, G. C.; Frost, J. M.; Fan, Y.; Garrison, T. R.; Daza, A. V.; Grayson, G. K.; Zhu, C. Z.; Pai, M.; Chandran, P.; et al. In Vitro and in Vivo Characterization of A-796260: A Selective Cannabinoid CB2 Receptor Agonist Exhibiting Analgesic Activity in Rodent Pain Models. *Br. J. Pharmacol.* **2008**, *153* (2), 390–401.
- (286) Felder, C. C.; Joyce, K. E.; Briley, E. M.; Mansouri, J.; Mackie, K.; Blond, O.; Lai, Y.; Ma, A. L.; Mitchell, R. L. Comparison of the Pharmacology and Signal Transduction of the Human Cannabinoid CB1 and CB2 Receptors. *Mol. Pharmacol.* **1995**, *48* (3), 443–450.
- (287) Riether, D.; Wu, L.; Cirillo, P. F.; Berry, A.; Walker, E. R.; Ermann, M.; Noya-Marino, B.; Jenkins, J. E.; Albaugh, D.; Albrecht, C.; et al. 1,4-Diazepane Compounds as Potent and Selective CB2 Agonists: Optimization of Metabolic Stability. *Bioorg. Med. Chem. Lett.* **2011**, *21* (7), 2011–2016.
- (288) Zindell, R.; Walker, E. R.; Scott, J.; Amouzegh, P.; Wu, L.; Ermann, M.; Thomson, D.; Fisher, M. B.; Fullenwider, C. L.; Grbic, H.; et al. Aryl 1,4-Diazepane Compounds as Potent and Selective CB2 Agonists: Optimization of Drug-like Properties and Target Independent Parameters. *Bioorg. Med. Chem. Lett.* **2011**, *21* (14), 4276–4280.
- (289) Hickey, E. R.; Zindell, R.; Cirillo, P. F.; Wu, L.; Ermann, M.; Berry, A. K.; Thomson, D. S.; Albrecht, C.; Gemkow, M. J.; Riether, D. Selective CB2 Receptor Agonists. Part 1: The Identification of Novel Ligands through Computer-Aided Drug Design (CADD) Approaches. *Bioorg. Med. Chem. Lett.* **2015**, *25* (3), 575–580.
- (290) Riether, D.; Zindell, R.; Wu, L.; Betageri, R.; Jenkins, J. E.; Khor, S.; Berry, A. K.; Hickey, E. R.; Ermann, M.; Albrecht, C.; et al. Selective CB2 Receptor Agonists. Part 2: Structure-Activity Relationship Studies and Optimization of Proline-Based Compounds. *Bioorg. Med. Chem. Lett.* **2015**, *25* (3), 581–586.
- (291) Bartolozzi, A.; Cirillo, P. F.; Berry, A. K.; Hickey, E. R.; Thomson, D. S.; Wu, L.; Zindell, R.; Albrecht, C.; Ceci, A.; Gemkow, M. J.; et al. Selective CB2 Receptor Agonists. Part 3: The Optimization of a Piperidine-Based Series That Demonstrated Efficacy in an in Vivo Neuropathic Pain Model. *Bioorg. Med. Chem. Lett.* **2015**, *25* (3), 587–592.
- (292) Han, S.; Thoresen, L.; Zhu, X.; Narayanan, S.; Jung, J.-K.; Strah-Pleynet, S.; Decaire, M.; Choi, K.; Xiong, Y.; Yue, D.; et al. Discovery of 1a,2,5,5a-Tetrahydro-1H-2,3-Diazacyclopropa[a]pentalen-4-Carboxamides as Potent and Selective CB2 Receptor Agonists. *Bioorg. Med. Chem. Lett.* **2015**, *25* (2), 322–326.
- (293) Tourteau, A.; Andrzejak, V.; Body-Malapel, M.; Lemaire, L.; Lemoine, A.; Mansouri, R.; Djouina, M.; Renault, N.; El Bakali, J.; Desreumaux, P.; et al. 3-Carboxamido-5-Aryl-Isloxazoles as New CB2 Agonists for the Treatment of Colitis. *Bioorg. Med. Chem.* **2013**, *21* (17), 5383–5394.
- (294) Tourteau, A.; Leleu-Chavain, N.; Body-Malapel, M.; Andrzejak, V.; Barczyk, A.; Djouina, M.; Rigo, B.; Desreumaux, P.; Chavatte, P.; Millet, R. Switching Cannabinoid Response from CB(2) Agonists to FAAH Inhibitors. *Bioorg. Med. Chem. Lett.* **2014**, *24* (5), 1322–1326.

- (295) Yrjölä, S.; Kalliokoski, T.; Laitinen, T.; Poso, A.; Parkkari, T.; Nevalainen, T. Discovery of Novel Cannabinoid Receptor Ligands by a Virtual Screening Approach: Further Development of 2,4,6-Trisubstituted 1,3,5-Triazines as CB2 Agonists. *Eur. J. Pharm. Sci.* **2013**, *48* (1-2), 9–20.
- (296) Yrjölä, S.; Sarparanta, M.; Airaksinen, A. J.; Hytti, M.; Kauppinen, A.; Pasonen-Seppänen, S.; Adinolfi, B.; Nieri, P.; Manera, C.; Keinänen, O.; et al. Synthesis, in Vitro and in Vivo Evaluation of 1,3,5-Triazines as Cannabinoid CB2 Receptor Agonists. *Eur. J. Pharm. Sci.* **2015**, *67*, 85–96.
- (297) Horvath, B.; Magid, L.; Mukhopadhyay, P.; Batkai, S.; Rajesh, M.; Park, O.; Tanchian, G.; Gao, R. Y.; Goodfellow, C. E.; Glass, M.; et al. A New Cannabinoid CB2 Receptor Agonist HU-910 Attenuates Oxidative Stress, Inflammation and Cell Death Associated with Hepatic Ischaemia/reperfusion Injury. *Br. J. Pharmacol.* **2012**, *165* (8), 2462–2478.
- (298) Smoum, R.; Baraghithy, S.; Chourasia, M.; Breuer, A.; Mussai, N.; Attar-Namdar, M.; Kogan, N. M.; Raphael, B.; Bolognini, D.; Cascio, M. G.; et al. CB2 Cannabinoid Receptor Agonist Enantiomers HU-433 and HU-308: An Inverse Relationship between Binding Affinity and Biological Potency. *Proc. Natl. Acad. Sci.* **2015**, 201503395.
- (299) Yacovan, A.; Bar-Joseph, A.; Meilin, S.; Amselem, S. Orally Effective Cannabinoid Analogs. WO 2006043260, 2006.
- (300) Gratzke, C.; Streng, T.; Stief, C. G.; Alroy, I.; Limberg, B. J.; Downs, T. R.; Rosenbaum, J. S.; Hedlund, P.; Andersson, K.-E. Cannabinor, a Selective Cannabinoid-2 Receptor Agonist, Improves Bladder Emptying in Rats with Partial Urethral Obstruction. *J. Urol.* **2011**, *185* (2), 731–736.
- (301) Gratzke, C.; Streng, T.; Stief, C. G.; Downs, T. R.; Alroy, I.; Rosenbaum, J. S.; Andersson, K.-E.; Hedlund, P. Effects of Cannabinor, a Novel Selective Cannabinoid 2 Receptor Agonist, on Bladder Function in Normal Rats. *Eur. Urol.* **2010**, *57* (6), 1093–1100.
- (302) GlaxoSmithKline. Dental Pain 3rd Molar Tooth Extraction GW842166. <https://clinicaltrials.gov/ct2/show/NCT00444769>. Identifier: NCT00444769.
- (303) Ostfeld, T.; Price, J.; Albanese, M.; Bullman, J.; Guillard, F.; Meyer, I.; Leeson, R.; Costantin, C.; Ziviani, L.; Nocini, P. F.; et al. A Randomized, Controlled Study to Investigate the Analgesic Efficacy of Single Doses of the Cannabinoid Receptor-2 Agonist GW842166, Ibuprofen or Placebo in Patients with Acute Pain Following Third Molar Tooth Extraction. *Clin. J. Pain* **2011**, *27* (8), 668–676.
- (304) Muthuppalaniappan, M.; Balasubramanian, G.; Gullapalli, S.; Joshi, N. K.; Narayanan, S. Novel Cannabinoid Receptor Ligands, Pharmaceutical Compositions Containing Them, and Process for Their Preparation. WO 2006/129178., 2006.
- (305) Servettaz, A.; Kavian, N.; Nicco, C.; Deveaux, V.; Chéreau, C.; Wang, A.; Zimmer, A.; Lotersztajn, S.; Weill, B.; Batteux, F. Targeting the Cannabinoid Pathway Limits the Development of Fibrosis and Autoimmunity in a Mouse Model of Systemic Sclerosis. *Am. J. Pathol.* **2010**, *177* (1), 187–196.

- (306) Kyowa Hakko Kirin Company. A Comparative Study of KHK6188. <https://clinicaltrials.gov/ct2/show/NCT01544296>. Identifier: NCT01544296.
- (307) Rahn, E. J.; Hohmann, A. G. Cannabinoids as Pharmacotherapies for Neuropathic Pain: From the Bench to the Bedside. *Neurotherapeutics* **2009**, *6* (4), 713–737.
- (308) McGaraughty, S.; Chu, K. L.; Dart, M. J.; Yao, B. B.; Meyer, M. D. A CB(2) Receptor Agonist, A-836339, Modulates Wide Dynamic Range Neuronal Activity in Neuropathic Rats: Contributions of Spinal and Peripheral CB(2) Receptors. *Neuroscience* **2009**, *158* (4), 1652–1661.
- (309) Yao, B. B.; Hsieh, G.; Daza, A. V.; Fan, Y.; Grayson, G. K.; Garrison, T. R.; El Kouhen, O.; Hooker, B. A.; Pai, M.; Wensink, E. J.; et al. Characterization of a Cannabinoid CB₂ Receptor-Selective Agonist, A-836339 [2,2,3,3-Tetramethyl-Cyclopropanecarboxylic Acid [3-(2-Methoxy-Ethyl)-4,5-Dimethyl-3H-Thiazol-(2Z)-Ylidene]-Amide], Using in Vitro Pharmacological Assays, in Vivo Pain Models, and . *J. Pharmacol. Exp. Ther.* **2009**, *328* (1), 141–151.
- (310) Hsieh, G. C.; Pai, M.; Chandran, P.; Hooker, B. A.; Zhu, C. Z.; Salyers, A. K.; Wensink, E. J.; Zhan, C.; Carroll, W. A.; Dart, M. J.; et al. Central and Peripheral Sites of Action for CB₂ Receptor Mediated Analgesic Activity in Chronic Inflammatory and Neuropathic Pain Models in Rats. *Br. J. Pharmacol.* **2011**, *162* (2), 428–440.
- (311) Horti, A. G.; Gao, Y.; Ravert, H. T.; Finley, P.; Valentine, H.; Wong, D. F.; Endres, C. J.; Savonenko, A. V.; Dannals, R. F. Synthesis and Biodistribution of [¹¹C]A-836339, a New Potential Radioligand for PET Imaging of Cannabinoid Type 2 Receptors (CB₂). *Bioorg. Med. Chem.* **2010**, *18* (14), 5202–5207.
- (312) Eli Lilly and Company. A Study of LY2828360 in Patients With Osteoarthritic Knee Pain. <https://clinicaltrials.gov/ct2/show/NCT01319929>. Identifier: NCT01319929.
- (313) Kalliomäki, J.; Segerdahl, M.; Webster, L.; Reimfelt, A.; Huizar, K.; Annas, P.; Karlsten, R.; Quiding, H. Evaluation of the Analgesic Efficacy of AZD1940, a Novel Cannabinoid Agonist, on Post-Operative Pain after Lower Third Molar Surgical Removal. *Scand. J. Pain* **2013**, *4* (1), 17–22.
- (314) Sekiguchi, K.; Kanazu, T.; Takeuchi, M.; Hasegawa, H.; Yamaguchi, Y. Non-Clinical Evaluation of the Metabolism, Pharmacokinetics and Excretion of S-777469, a New Cannabinoid Receptor 2 Selective Agonist. *Xenobiotica.* **2014**, *44* (1), 48–58.
- (315) Cara Therapeutics, Inc. A Phase 2 Study to Evaluate Analgesic Effect of IV CR845 For Pain Following Bunionectomy Surgery. <https://clinicaltrials.gov/ct2/show/NCT01789476>. Identifier: NCT01789476.
- (316) Haruna, T.; Soga, M.; Morioka, Y.; Hikita, I.; Imura, K.; Furue, Y.; Yamamoto, M.; Imura, C.; Ikeda, M.; Yamauchi, A.; et al. S-777469, a Novel Cannabinoid Type 2 Receptor Agonist, Suppresses Itch-Associated Scratching Behavior in Rodents through Inhibition of Itch Signal Transmission. *Pharmacology* **2015**, *95* (1-2), 95–103.

- (317) Odan, M.; Ishizuka, N.; Hiramatsu, Y.; Inagaki, M.; Hashizume, H.; Fujii, Y.; Mitsumori, S.; Morioka, Y.; Soga, M.; Deguchi, M.; et al. Discovery of S-777469: An Orally Available CB2 Agonist as an Antipruritic Agent. *Bioorg. Med. Chem. Lett.* **2012**, *22* (8), 2803–2806.
- (318) Sekiguchi, K.; Fukumura, K.; Hasegawa, H.; Kanazu, T. The Metabolism and Pharmacokinetics of [14 C]-S-777469, a New Cannabinoid Receptor 2 Selective Agonist, in Healthy Human Subjects. *Xenobiotica* **2015**, *45* (2), 150–157.
- (319) Takai, N.; Tanaka, Y.; Saji, H. Quantification of Small Molecule Drugs in Biological Tissue Sections by Imaging Mass Spectrometry Using Surrogate Tissue-Based Calibration Standards. *Mass Spectrom. (Tokyo, Japan)* **2014**, *3* (1), A0025.
- (320) Shionogi Inc. A Phase Ib/IIa, Double-Blind, Randomized Study to Assess the Safety, Tolerability, Pharmacokinetics, and Pharmacodynamics of S-777469 in Subjects With Atopic Dermatitis. <https://clinicaltrials.gov/ct2/show/NCT00697710>. Identifier: NCT0069771.
- (321) Sekiguchi, K.; Kanazu, T.; Takeuchi, M.; Hasegawa, H.; Yamaguchi, Y. Non-Clinical Evaluation of the Metabolism, Pharmacokinetics and Excretion of S-777469, a New Cannabinoid Receptor 2 Selective Agonist. *Xenobiotica*. **2014**, *44* (1), 48–58.

Table of Contents Graphic

